



Review Article

Halophiles-A Review

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ABSTRACT

Keywords

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Extremophiles, able to live in unusual habitats can serve as a potential source of novel microorganisms and their metabolites gain diverse applications and thus these microorganisms are extensively studied. Halophiles are a group of microorganisms that live in saline environments and in many cases require salinity to survive. Hypersaline habitats are tremendous sources of halophilic microorganisms and studies of these microorganisms are of special importance as they are capable of producing compounds of diverse industrial, pharmaceutical and biomedical potentials.

Introduction

Extremophiles are those microorganisms which require extreme environment for growth. The term extremophile was first used by MacElroy in 1974. Extreme environment is a relative term, since environments that are extreme for one organism may be essential for the survival of another organism. Extremophiles are those microorganism which live in extreme environment like high or low temperature, salt concentration, pH etc. Extremophiles live under conditions that would kill most other creatures and many cannot survive in the normal global conditions. On the bases of their habitation these extremophiles can be categorized as: acidophiles, alkalophiles, halophiles, thermophiles and psychrophiles. Among extremophiles halophiles are those which are able to survive in high salt

concentrations. These microorganisms inhabit the world's most saline environments, like hypersaline lake, solar saltern, evaporation pond, salt flats and tidal marine environments. Halophilic bacteria have the capacity to balance the osmotic pressure of the environment and resist the denaturing effects of salts. On the basis of their halotolerance or optimum salt concentration wherein these organisms grow they are categorized as slight (2-5% NaCl), moderate (5-15% NaCl) or extreme halophiles (15-30% NaCl).

To survive in higher saline environment halophilic microorganisms use different types of strategies, they synthesize compatible solutes in cells or possess the transporters that help them to survive in

such type of harsh condition. Osmoregulatory solutes such as potassium ion, glutamate, proline, ectoine and betaine have been reported in these bacteria (Madigan, 1999 and Galinski, 1993). Some halophiles produce acidic proteins that have the capacity to function in high salt concentration.

Examples of well-adapted and widely distributed extremely halophilic microorganisms include archaea for example

Halobacterium sp. NRC-1, cyanobacteria

such as *Aphanothece halophytica* and the green alga *Dunaliella salina*. Multicellular halophilic eukaryotic organisms include brine shrimp and the larvae of brine flies.

Occurrence and Distribution

Halophiles can be found in areas where the concentration of salt is five times greater than that salt concentration of the ocean. Saline and hypersaline environments are broadly dispersed all through world either salt lakes (The Dead Sea, The Great Salt Lakes etc.) or salt mines [Salzbad-Salzetnan (Austria), Slanic, Turda, Praid (Romania), Wieliczka (Poland), Nakhlichevan (Azerbaijan), Chon-Tous (Kirghizstan), Cave Berezniki in Perm(Russia), Sototvino (Ukraine) etc.]. These hypersaline environments are too harsh for normal life to exist, but a variety of microorganisms survive.

These microorganisms produce different bioactive compounds like enzymes, pigments, solutes, metabolites and exopolysaccharides which have the ability to

work in such type of saline conditions. These properties can be used for the development of techniques outlined essentially on the optimal condition of these biomolecules.

These properties could be exploited for the development of additional bio-industrial processes based on the optimal conditions of these biomolecules. To cope up with the high and often changing salinity of their environment, the aerobic halophilic bacteria, similar to all other microorganisms, need to balance their cytoplasm with the osmotic pressure exerted by the external medium (Madigan, 1999; Oren, 2008; Oren, 2010).

Important Characteristics Of Halophiles

Enzymes

Enzymes are catalysts which have potential applications in food, detergent formulations, metal recovery, leather processing and in several industries. Extremophile enzymes, especially extracellular, have been found to have potential industrial applications as they can survive and catalyze reactions in unusual environment.

Table 1 has listed different isolation sites from where halophilic microorganisms have been isolated and different enzymes which are assayed.

Nucleases

A nuclease (nuclease H) is reported from "*M. varians* subsp. *halophilus*" having both DNase and RNase activities. Another halophilic nuclease (an exonuclease, releasing 5'-mononucleotides from both DNA and RNA) was produced by *Bacillus halophilus* (Onishi et al., 1983).

Amylases

A few α -amylases were purified and characterized from halophilic microorganisms, *Acinetobacter* (Onishiet *et al.*, 1980), *Halobacterium salinarum* (Goodand Hartman1970, Patel *et al.*, 1993), *M. varians* subsp. *halophilus*" (Kobayashiet *et al.*, 1986), *N. halobia* (Onishiet *et al.*, 1991), *Natronococcus amylolyticus* (Kobayashi *et al.*, 1992), *Halomonas meridiana* (Coronado *et al.*, 2000), *Haloferax mediterranei* (Perez-Pomares *et al.*, 2003)etc.

The enzyme from *Haloarcula* sp. S-1 showed high tolerance to various organic solvents (Fukushima *et al.*, 2005). The effect of ionic strength on the amylase activity, has also been reported at various ratios of Na⁺ and Mg²⁺ concentrations (Enache *et al.*, 2009).

Proteases

Halophilic microorganisms produce proteases which can have novel applications (Margesin *et al.*, 2001) because of its high stability at saturated salt concentrations or organic solvent tolerance.

Halobacterium salinarum (Ryu *et al.*, 1994) produce an extra cellular serine protease that has potential to be used for peptide synthesis. The other serine proteases were isolated from *Natronococcus soccultus* (Studdert *et al.*,1997), *Natronomonas pharaonis* (Stan-Lotter *et al.*, 1999), *Natrialba magadii* (Gimenez *et al.*,2000). Some other proteases were purified and characterized from *Natrialba asiatica* (Kamekura *et al.*, 1992) and *Haloferax mediterranei* (Kamekura *et al.*,1992 and Kamekura *et al.*,1996) and other halophilic isolates (Norberg and Hofsten, 1969; Kamekura and Onishi, 1974; Kamekura and Seno, 1990).

In recent years, a number of studies have been conducted to characterize extracellular proteases from *Salinivibrio* genus (Lama *et al.*,2005 and Amoozegar *et al.*, 2007), *Pseudoalteromonas* sp. strain CP76 (Sa'nchez-Porro *et al.*, 2003), *Natrialba magadii* (Gime'nez *et al.*, 2000), *Halobacterium mediterranei* (Stepanov *et al.* 1992), *Bacillus clausii* (Kumar *et al.* 2004). There is also a report that a chymotrypsinogen B-like protease was isolated from the haloalkaliphilic archaeon *Natronomonas pharaonis* (Stan-Lotteret *et al.*, 1999).

Lipases

Lipase is one of the most important hydrolytic enzymes with potential in various fields of pharmaceutical industry and agriculture. Various moderately or extremely halophilic microorganisms have been reported for the production of lipase which are also stable at high temperature - *Salinivibrio* sp. (Amoozegar *et al.*, 2008), *Natronococcus* sp. (Boutaiba *et al.*, 2006), haloarchaeal strains (Ozcan *et al.*, 2009) etc.

Cellulose-Degrading Enzymes

Bolobova *et al.*, (1992) first reported a cellulose-utilizing, extremely halophilic bacterium. The obligate anaerobic organism named *Halocella cellulolytica* is able to utilize cellulose as a sole carbon source. Another work has shown that many cellulose-utilizing extremely halophilic Archaea are present in subsurface salt formation (Vreeland *et al.*,1998). A preliminary work done by Cojoc *et al.*, (2009) on extracellular hydrolytic enzymes of halophilic microorganisms from subterranean rock salt revealed the presence of cellulose.

Pigments

Halophilic microorganisms are a great source of diverse natural products. Carotenoid pigments are one of these natural products responsible for the yellow, orange, red, and purple colors in a wide variety of plants, animals, and microorganisms (Li Z *et al.*, 2012 and Cabral *et al.*, 2011). Carotenoid pigments are particularly prominent in hypersaline environment. Red and orangish color of hypersaline habitats is due to the presence of pigmented microorganisms, including *Dunaliella*, rich in β -carotene, Haloarchaea whose main production is bacterioruberin, and halophilic bacteria, such as *Salinibacter ruber* producing a carotenoid called salinixanthin (El-Banna Aaet *et al.*, 2012 and Jehlicka *et al.*, 2013). An extremely halophilic archaeon isolated from Urmia Lake, *Halorubrum* sp. TBZ126 is reported as potential producer of carotenoids (Naziri *et al.*, 2014).

Melanin is nearly a ubiquitous pigment having immense application potentials in the field of agriculture, cosmetics and pharmaceutical industries (photoprotection and mosquitocidal activity isolated from *Streptomyces*). Rani *et al.*, (2013) reported a halophilic black yeast, *Hortaea werneckii* which produced a diffusible dark pigment on potato dextrose agar. It also showed inhibitory activity against potential pathogens and activity was observed in *Salmonella typhi* and *Vibrio parahaemolyticus*.

Resistance to Antibiotics

The emergence of antibiotic resistance is an evolutionary process that is based on selection for organisms that have enhanced ability to survive doses of antibiotics that would have previously been lethal (Cowen, 2008). Pearson and Carol (2008) reported different antibiotics likes penicillin,

erythromycin and gentamycin which used to be one of the important cures are now less effective because bacteria have become more resistant.

Halophilic bacteria isolated from different region were found resistant against different antibiotics :Halobacteriaceae (Hilpert, 1981), *Halomonas elongate* (Boneloet *et al.*, 1984), *Bacillus cereus* SIU1 (Singhet *et al.*, 2010).

Antimicrobial Activity

Foreshore soil of Daecheon Beach and Saemangeum Sea of Korea represents an untapped source of bacterial biodiversity and also that most actinobacterial isolates are capable of antibacterial and antifungal metabolite production. The halophiles isolated from Ratnagiri coastal area (marine environments) having antibacterial and antifungal activity. The antibacterial and antifungal assays of halophiles (protein crude extract) have shown that, the marine environments represent a potential source of new antimicrobial and antifungal agents (Todkar *et al.*, 2012).

Microbacterium oxydans and *Streptomyces fradiae* showed antibacterial activity against all tested pathogenic bacteria and yeasts (Irshad *et al.*, 2013). Sawale *et al.*, (2014) collected soil samples from coastal area of Arabic ocean (Mumbai) and reported antibiotics production from two bacterial strains *Bacillus pumilus* (NKCM 8905) and *Bacillus pumilus* (AB211228).

Applications

Biotechnological Application

Moderately halophilic bacteria have the potential for exciting and promising applications. Not only do many of them produce compounds of industrial interest

(enzymes, polymers and osmoprotectants), but also they possess useful physiological properties which can facilitate their exploitation for commercial purposes. Here, we discuss some of the current industrial applications of these halophiles and emphasize some expected future developments.

Fermented Foods

Different fermented food products are prepared at a high salt concentration. For the preparation of such type of food products different microorganism are useful which can survive in such type of condition. For example in preparation of Thai fish sauce (nam pla) the first halophilic archaeon isolated resembles *Halobacterium salinarum* (Thongthai and Suntinanalert, 1991 and Thongthai *et al.*, 1992) and *Halococcus thailandensis* and *Natrinema gari*, were two new species recently isolated (Namwong *et al.*, 2007 and Tapingkae *et al.*, 2008). In fermentation of soy sauce halophilic lactococci (*Tetragenococcus halophilus*) are used as starters (Kushner *et al.*, 1998). In the preparation of fermented liver sauce *Tetragenococcus muritianus* is involved. *Halalkalicoccus jeotgali* is a novel isolate obtained from shrimp jeotgal (Roh *et al.*, 2007).

Halophilic Enzymes

Some of the halophilic bacteria produce hydrolytic enzymes which are stable at high salt concentration. These enzymes have diverse application in different fields. Some of these enzymes are proteases which have broad application in detergents, leather industry, food industry, pharmaceutical industry and bioremediation processes (Anwar and Saleemuddin, 1998 and Gupta *et al.*, 2002). Bacteria of *Halobacterium* species like *Haloferax mediteranei*, *Natrial baasiatica*, *Natrial bamagadii*, *Natrono*

coccusoccultus and *Natrono monaspharaonis* show proteolytic activity with potential industrial applications.

Amylases are used industrially in the production of high fructose corn syrup (hydrolysis of corn starch), the textile industry and are also used in laundry detergents. *Halomonas meridian*, *Haloarcula hispanica* and *Natronococcus amylolyticu* are some of halophilic bacteria from which amylases have been characterized.

Halophilic restriction endonuclease have diverse application. For the industrial production of the flavoring agent 5'-inosinic acid and 5'-guanylic acid Nuclease H produced by "*M. varians* subsp. *halophilus*" is useful (Kamekura *M et al.*, 1982). Some of endonucleases *HacI*, *HcuI*, *HhII* and *Hsal* have been isolated and produced on commercial scale from *Halococcus acetoinfaciens*, *Halobacterium cutirubrum*, *H. halobium* and *Halobacterium salinarium* respectively.

Esterases and lipases are widely used as biocatalysts and are also useful ingredients in laundry detergents. The lipase from *Thermomyces lanuginosus* is commercially produced and sold as Lipolase by Novozymes, Denmark since 1989. An esterase from *Haloarculam arismortui* has also been purified and characterized.

Cellulase is used for commercial food processing in coffee and in textile industry, laundry detergents, in pulp and paper industry and they are even used for pharmaceutical applications (Cavaco- Paulo, 1998; Tolan and Foody, 1999 and Ikeda *et al.*, 2006).

Compatible Solutes

To adapt to the osmotic stress and to keep up positive turgor pressure halophiles low molecular weight organic compounds. Some compatible solutes like glycine, betaine and ectoines, may be used as stress protectants and stabilizers of enzymes, nucleic acids, membranes and whole cells. The modern utilization of these compounds in enzyme technology (biosensor technology, PCR, and

so forth) and for pharmaceuticals and cosmetics is a most encouraging field (Galinski and Tindall, 1992 and Ventosa *et al.*, 1995). It may also enable the transfer of genes for salt and drought tolerance from moderate halophiles to agricultural crops, such as wheat, rice and barley, enabling them to grow in more saline soils.

Table.1 Microorganisms able to Produce Hydrolytic Enzymes Isolated from Different Hypersaline Environments

Isolation Site	Hydrolytic Activity Assayed	Most Abundant Hydrolytic Activity	Isolate Affiliation	References
Salterns in Almeria, Cadiz and Huelva (Spain)	Amylase protease lipase DNase Pullulanase	Amylase	<i>Salinivibrio</i> <i>Halomonas</i> <i>Chromohalobacter</i> <i>Bacillus</i> - <i>Salibacillus</i> <i>Salinicoccus</i> <i>Marinococcus</i>	Sánchez-Porroet <i>al.</i> ,(2003)
Saltern in Huelva (Spain)	Lipase protease amylase nuclease	Amylase	<i>Halorubrum</i> <i>Haloarcula</i> <i>Halobacterium</i> <i>Salicola</i> <i>Salinibacter</i> <i>Pseudomonas</i>	Morenoet <i>al.</i> ,(2009)
HowzSoltan Lake (Iran)	Lipase amylase protease xylanase DNase inulinase pectinase cellulase pulullanase	Lipase	<i>Salicola</i> <i>Halovibrio</i> <i>Halomonas</i> <i>Oceanobacillus</i> <i>Thalassobacillus</i> <i>Halobacillus</i> <i>Virgibacillus</i> <i>Gracilibacillus</i> <i>Salinicoccus</i> <i>Piscibacillus</i>	Rohban <i>et al.</i> ,(2009)
Deep-sea sediments of the Southern Okinawa Trough (China)	amylase protease lipase DNase	Amylase	<i>Alcanivorax</i> <i>Bacillus</i> <i>Cobetia</i> <i>Halomonas</i> <i>Methylarcula</i> <i>Micrococcus</i> <i>Myroides</i> <i>Paracoccus</i> <i>Planococcus</i> <i>Pseudomonas</i>	Danget <i>al.</i> ,(2009)

			<i>Psychrobacter</i> <i>Sporosarcina</i> <i>Sufflavibacter</i> <i>Wangia</i>	
SlanicPrahova salt mine (Romania)	amylase gelatinase lipase protease cellulase xylanase	Lipase Protease	ND	Cojoc <i>et al.</i> , (2009)
Maharlu Salt Lake (Iran)	protease lipase	Not defined	<i>Bacillus</i> <i>Paenibacillus</i> <i>Halobacterium</i> <i>Aeromonas</i> <i>Staphylococcus</i>	Ghasemi <i>et al.</i> , (2011 a,b)
Saline desert “Indian Wild Ass Sanctuary” (India)	Amylase	Not defined	<i>Bacillus</i>	Khunt <i>et al.</i> , (2011)
Atacama Desert (Chile)	amylase protease lipase DNase xylanase pullulanase	DNase	DNase <i>Bacillus</i> <i>Halobacillus</i> <i>Pseudomonas</i> <i>Halomonas</i> <i>Staphylococcus</i>	Moreno <i>et al.</i> , (2012)
Arabal soil of west coast of Karnataka	Protease Amylase Lipase gelatinase inulinase	ND	<i>Virgibacillus</i> <i>Salinicoccus</i>	Jayachandra <i>et al.</i> , (2012a, b)
Balta Albă salt lake from Romania	Protease Amylase Cellulase Esterase Gelatinase	ND	ND	Neagu <i>et al.</i> , (2013)
Solar salterns of Orissa and West Bengal, India	amylase glutaminase asparaginase xylanase cellulose gelatinase inulinase caseinase pectinase urease lipase	glutaminase asparaginase lipase caseinase	<i>Halomonas</i> <i>Cobetia</i> <i>Halococcus</i>	Biswas and Paul (2013)
Salt lakes of Romania	alanine- aminopeptidase α -glucosidase β -glucosidase	α -glucosidase β -glucosidase	ND	Păceșilăi <i>et al.</i> , (2014)

Degradation of Toxic Compounds

Hypersaline waters and soils are often contaminated with heavy metals or other

toxic compounds from different sources. Hypersaline wastewaters are generated during the manufacture of chemicals such as pesticides, pharmaceuticals and herbicides and during oil and gas recovery processes. Conventional microbiological treatment processes do not function at high salt concentrations and therefore the use of moderately halophilic bacteria should be considered (Oren *et al.* 1992,1993).

Halophiles which are able to degrade hydrocarbon have been isolated from different sites like Great Salt Lake (Ward and Brock, 1978) and Antarctic saline lakes (McMeekin *et al.*, 1993). Treatment of hypersaline wastewaters containing phenol was reported by Woolard and Irvine (1992) using a biofilm of a moderately halophilic bacterium. *H. halodurans* reported to cleave benzoate and other aromatic compounds. DeFrank *et al.*, (1991,1993) isolated halophilic bacteria, which were showing hydrolytic activity against several organophosphorus compounds. Halophiles belonging to the family *Halomonadaceae* have been recently isolated from highly saline sites contaminated with the herbicide 2, 4-dichlorophenoxyacetic acid, they were able to utilize chloroaromatic compounds as sources of carbon and energy.

Other Applications of Halophiles

Moderately halophilic bacteria may have various other uses in biotechnology, yet to be exploited. (i) Moderate halophiles could be used in the recovery of hypersaline waste brines derived from the olive oil industry and leather- or fur-curing processes. (iii) They may be screened for the production of bioactive compounds such as antibiotics. (iv) Biological surfactants derived from moderate halophiles may have a variety of

applications in industry. Thus, Yakimov *et al.*, (1996) recently isolated a moderate halophile which synthesizes a novel glycolipid belonging to a powerful novel class of biosurfactants. (vi) Many moderate halophiles produce orange or pink colonies, probably due to the production of carotenoids as a protective mechanism against photooxidation processes. Carotenoids have major applications in the food industry as food-coloring agents and as additives in health food products. Therefore, investigations of the utilization of moderate halophiles as producers of carotenoids could be of great interest.

In conclusion, comparative studies of micro flora in the extreme environments results in better understanding of the ecosystem and can benefit in designing the applications. Hypersaline environments represent a valuable source of different bioactive compounds with great economical potential in industrial, agricultural, chemical, pharmaceuticals and biotechnological applications. The microbial diversity can prove to be a valuable future resource in various industrial and biotechnological processes. Such microbes can also be used as a source of gene(s) that can increase salt tolerance in different crop species through genetic transformation.

Reference

- Amoozegar MA, Salehghamari E, Khajeh K, Kabiri M and Naddaf S. (2008). Production of an extracellular thermohalophilic lipase from a moderately halophilic bacterium, *Salinivibrio sp.* strain SA-2. *J. Basic Microbiol.*48:160–167.
- Anwar A and Saleemuddin M. (1998). Alkaline proteases. A Review. *Bioresource Technol.* 6: 175-183.

- Biswas J and Paul A K. (2013). Production of extracellular enzymes by halophilic bacteria isolated from solar salterns. *Int. J. of App. Biol and Pharma Technol.* 4 :30-36.
- Bolobova AV, Simankova M C and Markovich N A. (1992). Cellulase complex of a new halophilic bacterium *Halocella cellulolytica*. *Microbiol.* 61: 557–562.
- Bonelo G, Ventosa A, Megias M and Ruiz-Berraquero F. (1984). The sensitivity of halobacteria to antibiotics. *FEMS Microbiol Lett.* 21: 341-345.
- Boutaiba S, Bhatnagar T, Hacene H, Mitchell D A and Baratti J C. (2006). Preliminary characterization of a lipolytic activity from an extremely halophilic archaeon, *Natronococcus sp.* *J. Mol. Catal. B-Enzym.* 41: 21–26.
- Cabral MMS, Cence K, Zeni J, Tsai S M, Durrer A and Foltran LL. (2011). Carotenoids production from a newly isolated *Sporidiobolus pararoseus* strain by submerged fermentation. *Eur Food Res Technol.* 233(1):159–66.
- Cánovas D, Vargas C, Iglesias-Guerra F, Csonka L, Rhodes D, Ventosa A and Nieto J J. (1997). Isolation and characterization of salt-sensitive mutants of the moderate halophile *Halomonas elongata* and cloning of the ectoine synthesis genes. *J. Biol. Chem.* 272:25794–25801.
- Cavaco-Paulo A. (1998). Mechanism of cellulase action in textile processes. *Carbohydr. Polym.* 37: 273-277.
- Cojoc R, Merciu S, Popescu G, Dumitru L, Kamekura M and Enache M. (2009). Extracellular hydrolytic enzymes of halophilic bacteria isolated from a subterranean rock salt crystal, Rom. *Biotechnol. Lett.* 14: 4658–4664.
- Coronado MJ, Vargas C, Hofemeister J, Ventosa A and Nieto J. (2000). Production and biochemical characterization of an α -amylase from the moderate halophile *Halomonas meridian*. *FEMS Microbiol. Lett.* 183: 67–71.
- Cowen L E. (2008). The evolution of fungal drug resistance: Modulating the trajectory from genotype to phenotype. *Nat. Rev. Microbiol.* 6:187-198.
- Dang H, Zhu H, Wang J and Li T. (2009). Extracellular hydrolytic enzyme screening of culturable heterotrophic bacteria from deep-sea sediments of the Southern Okinawa Trough. *World J. Microbiol. Biotechnol.* 25: 71-79.
- DeFrank J J and Cheng T. (1991). Purification and properties of an organophosphorus acid anhydrase from a halophilic bacterial isolate. *J. Bacteriol.* 173:1938–1943.
- DeFrank J J, Beaudry W T, Cheng T C, Harvey S P, Stroup A N and Szafraniec L L. (1993). Screening of halophilic bacteria and *Alteromonas sp.* for organophosphorus hydrolyzing enzyme activity. *Chem. Biol. Interact.* 87:141–148.
- El-Banna AaE-R, El-Razek AM A and El-Mahdy AR. (2012). Isolation, identification and screening of carotenoid-producing strains of *Rhodotorula glutinis*. *Food Nutr (Roma)*. 3(5):627–33.
- Enache M, Popescu G, Dumitru L and Kamekura M. (2009). The effect of $\text{Na}^+/\text{Mg}^{2+}$ ratio on the amylase activity of haloarchaea isolated from Techirghiol lake, Romania, a low salt environment. *Proc. Rom. Acad. Series B.* 11: 3–7.
- Fukushima T, Mizuki T, Echigo A, Inoue A and Usami R. (2005). Organic solvent tolerance of halophilic α -

- amylase from a haloarchaeon, *Haloarcula* sp. strain S-1. *Extremophiles*.9:85–89.
- Galinski E A and Tindall B J. (1992). Biotechnological prospects for halophiles and halotolerant microorganisms. In: Herbert R A, Sharp R J, editors. Molecular biology and biotechnology of extremophiles. Glasgow, United Kingdom: Blackie. 76–114.
- Galinski EA. (1993). Compatible solutes of halophilic eubacteria: Molecular principles, water-solute interaction, stress protection. *Experientia*.49:487-496.
- Ghasemi Y, Rasoul-Amini S, Ebrahiminezhad A, Kazemi A, Shahbazia Mand Talebniia N. (2011a). Screening and Isolation of Extracellular Protease Producing Bacteria from the Maharloo Salt Lake. *Iran J. Pharm. Sci*.7:175-180.
- Ghasemi Y, Rasoul-Amini S, Kazemi A, Zarrini G, Morowvat M T and Kargar M. (2011b). Isolation and Characterization of Some Moderately Halophilic Bacteria with Lipase Activity. *Microbiol*.80, 483-487.
- Gimenez MI, Studdert CA, Sanchez J J and De Castro RE. (2000). Extracellular protease of *Natrialba magadii*: purification and biochemical characterization. *Extremophiles*.4,181–188.
- Good W A and Hartman P A.(1970). Properties of the amylase from *Halobacterium halobium*. *J. Bacteriol*.104:601–603.
- Gupta R, Beg Q K and Lorenz P. (2002). Bacterial alkaline proteases: molecular approaches and industrial applications. *App. Microbiol and Biotechnol*.59: 15-32.
- Hilpert R, Winter J, Hammes W and Kandler O. (1981). The sensitivity of archaeobacteria to antibiotics. *Zbl Baktl Hyg 1 Abt OrigC*.2: 11-20.
- Ikeda Y, Park E Y and Okida N. (2006). Bioconversion of waste office paper to gluconic acid in a turbine blade reactor by the filamentous fungus *Aspergillus niger*. *Bioresour. Technol*. 97: 1030-1035.
- Irshad A, Ahmad I and Kim S B. (2013). Isolation, characterization and antimicrobial activity of halophilic bacteria in foreshore soils. *African J. Microbiol Research*.7: 164-173.
- Jayachandra S Y, Parameshwar A B, Mohan Reddy K and Sulochana M B. (2012a). Characterization of extracellular hydrolytic enzymes producing extremely halophilic bacterium *Virgibacillus* sp. JS5. *World J. of Sci. and Technol*. 2(2):23-26.
- Jayachandra S Y, Anil Kumar S, Merley D P and Sulochana M B. (2012b). Isolation and characterization of extreme halophilic bacterium *Salinicoccus* sp. JAS4 producing extracellular hydrolytic enzymes. *Recent Research in Sci. and Technol* 2012, 4(4): 46-49
- Jehlicka J, Edwards H G and Oren A.(2013). Bacterioruberin and salinixanthin carotenoids of extremely halophilic Archaea and Bacteria: a Raman spectroscopic study. *Spectrochim Acta. A Mol. Biomol. Spectrosc*.106:99–103.
- Kamekura M and Onishi H.(1974). Protease formation by a moderately halophilic *Bacillus* strain. *Appl. Microbiol*. 27: 809–810
- Kamekura M, Hamakawa T and Onishi H. (1982). Application of halophilic nuclease H of *Micrococcus varians* subsp. *halophilus* to commercial production of flavoring agent 5'-

- GMP. *Appl. Environ. Microbiol.* 44:994–995.
- Kamekura M and Seno Y.(1990). A halophilic extracellular protease from a halophilic archaebacterium 172 P1.*Biochem. Cell Biol.* 68: 352–359.
- Kamekura M, Seno Y, Holmes M L and Dyall-Smith M L. (1992). Molecular cloning and sequencing of the gene for a halophilic alkaline serine protease (halolysin) from an unidentified halophilic archaea strain (172P1) and expression of the gene in *Haloferax volcanii*.*J. Bacteriol.* 174: 736–742.
- Kamekura M, Seno Y and Dyall-Smith ML. (1996). Halolysin R4, a serine proteinase from the halophilic archaeon *Haloferax mediterranei*; gene cloning, expression and structural studies. *Biochim. Biophys. Acta.*1294:159–167.
- Khunt M, Pandhi N and Rana A. (2011). Amylase from moderate halophiles isolated from wild ass excreta. *Int. J. Pharm. Bio. Sci.* 1: 586-592.
- Kobayashi T, Kamekura M, Kanlayakrit W and Onishi H. (1986). Production, purification, and characterization of an amylase from the moderate halophile, *Micrococcus varians* subsp. *halophilus*. *Microbios* .46:165–177.
- Kobayashi T, Kamai H, Aono R, Horikoshi K and Kudo T. (1992). Haloalkaliphilic maltotriose-forming α -amylase from the archaebacterium *Natronococcus* sp. strain Ah-36. *J. Bacteriol.*174: 3439–3444.
- Kumar C G, Joo H S, Koo Y M, Paik S R and. Chang C S.(2004). Thermostable alkaline protease from a novel marine haloalkaliphilic *Bacillus clausii* isolate. *World J. Microbiol. Biotechnol.*20: 351–357.
- Lama L, Romano I, Calandrelli V, Nicolaus B and Gambacorta A .(2005). Purification and characterization of a protease produced by an aerobic haloalkaliphilic species belonging to the *Salinivibrio* genus. *Res. Microbiol.* 156:S478–484
- Li Z, Sun M, Li Q, Li A and Zhang C. (2012). Profiling of carotenoids in six microalgae (Eustigmatophyceae) and assessment of their beta-carotene productions in bubble column photobioreactor. *Biotechnol. Lett.*34(11):2049–53
- MacElroy R D.(1974). Some comments on evolution of Extremophiles. *Biosystems.*6: 75-75
- Madigan M T and Oren A. (1999). Thermophilic and halophilic extremophiles. *Curr. Opin. Microbiol.*2:265–269.
- Margesin R and Schinner F. (2001). Potential of halotolerant and halophilic microorganisms for biotechnology. *Extremophiles.*5:73–83.
- McMeekin T A, Nichols P D, Nichols S D, Juhasz A and Franzmann P D. (1993). Biology and biotechnological potential of halotolerant bacteria from Antarctic saline lakes. *Experientia.*49:1042–1046.
- Moreno M L, García M T, Ventosa A and Mellado E. (2009). Characterization of *Salicola* sp. IC10, a lipase- and protease-producing extreme halophile. *FEMS Microbiol. Ecol.*68: 59-71.
- Moreno M L, Piubeli F, Bonfá M R, García M T , Durrant L and Mellado E. (2012). Analysis and characterization of cultivable extremophilic hydrolytic bacterial community in heavy-metal-contaminated soils from the Atacama Desert and their

- biotechnological potentials. *J. Appl. Microbiol.*113: 550-559.
- Namwong S, Tanasupawat S, Visessanguan W, Kudo T and Itoh T. (2007).*Halococcus thailandensis* sp. nov., from fish sauce in Thailand. *Int. J. Syst. Evol. Microbiol.* 57: 2199–2203.
- Naziri D, Hamidi M, Hassanzadeh S, Tarhriz V, Zanjani B M, Nazemyieh H, Hejazi M A and Hejazi M S.(2014). Analysis of Carotenoid Production by *Halorubrum* sp. TBZ126; an Extremely Halophilic Archeon from Urmia Lake. *Adv. Pharma. Bull.* 4(1): 61-67.
- Neagu S, Enache M and Cojoc R. (2013). Extracellular hydrolytic activities of halophilic microorganisms isolated from Balta Albă salt lake. *Romanian Biotechnol. Letters.* 19:8951-8958.
- Norberg P and Hofsten B.(1969). Proteolytic enzymes from extremely halophilic bacteria. *J. Gen. Microbiol.* 55: 251–256.
- Onishi H, Fuchi H, Konomi K, Hidaka O and Kamekura M. (1980). Isolation and distribution of a variety of halophilic bacteria and their classification by salt-response. *Agric. Biol. Chem.*44:1253–1258.
- Onishi H, Mori T, Takeuchi S, Tani K, Kobayashi T and Kamekura M. (1983). Halophilic nuclease of a moderately halophilic *Bacillus* sp.: production, purification and characteristics. *Appl. Environ. Microbiol.*45:24–30.
- Onishi H, Yokoi H and Kamekura M. (1991).An application of a bioreactor with flocculated cells of halophilic *Micrococcus varians* subsp. *halophilus* which preferentially adsorbed halophilic nuclease H to 5'-nucleotide production. In: Rodriguez-Valera F, editor. General and applied aspects of halophilic microorganisms. New York, N.Y: Plenum Press. 341–349.
- Oren A, Gurevich P, Azachi M and Henis Y. (1992). Microbial degradation of pollutants at high salt concentrations. *Biodegradation.*3:387–398.
- Oren A, Gurevich P, Azachi M and Henis Y. (1993). Microbial degradation of pollutants at high salt concentrations. In: Rosenberg E, editor. Microorganisms to combat pollution. Dordrecht, The Netherlands: Kluwer Academic Publishers. 263–274.
- Oren A. (2008). Microbial life at high salt concentrations: phylogenetic and metabolic diversity. *Saline Systems.*
- Oren A.(2010). Industrial and environmental applications of halophilic microorganisms. *Environ. Technol.* 3(1): 825-834.
- Ozcan B, Ozyilmaz G, Cokmus C and Caliskan M.(2009). Characterization of extracellular esterase and lipase activities from five halophilic archaeal strains. *J. Ind. Microbiol. Biotechnol.*36:105–110.
- Păceșilă I, Cojoc R and Enache M.(2014). Evaluation of halobacterial extracellular hydrolytic activities in several natural saline and hypersaline lakes from Romania. *British Biotechnol J.* 4(5): 541-550.
- Patel S, Jain N and Madamwar D. (1993). Production of α -amylase from *Halobacterium halobium*. *World J. Microbiol. Biotechnol.*9:25–28.
- Pearson and Carol. (2008). Antibiotic resistance fast-growing problem worldwide. *Voice of America.* 12-29.
- Perez-Pomares F, Bautista V, Ferrer J, Pire C, Marhuenda-Egea F C and Bonete MJ. (2003). α -Amylase activity from the halophilic archaeon *Haloferax mediterranei*. *Extremophiles.* 7:299–306.

- Rani M H S, Ramesh T, Subramanian J and Kalaiselvam M. (2013). Production and Characterization of Melanin Pigment from Halophilic Black Yeast *Hortaea werneckii*. *Int. J. of Pharma. Res. & Review*. 2(8):9-17.
- Roh S W, Nam Y D, Chang H W, Sung Y, Kim K H, Oh H M and Bae J W.(2007). *Halalkalicoccus jeotgali* sp. nov., a halophilic archaeon from shrimp jeotgal, a traditional Korean fermented seafood,*Int. J. Syst. Evol. Microbiol.* 57: 2296–2298.
- Rohban R, Amoozegar M A and Ventosa A. (2009). Screening and isolation of halophilic bacteria producing extracellular hydrolysis from Howz Soltan Lake, Iran. *J. Ind. Microbiol. Biotechnol.*36: 333-340.
- Ryu K, Kim J and Dordick JS. (1994). Catalytic properties and potential of an extracellular protease from an extreme halophile.*Enzyme Microbiol. Technol.* 16: 266–275.
- Sánchez-Porro C, Martín S, Mellado E and Ventosa A. (2003). Diversity of moderately halophilic bacteria producing extracellular hydrolytic enzymes. *J. Appl. Microbiol.* 94: 295-300.
- Sawale A, Kadam T A, Karale M A and Kadam O A. (2014).Antimicrobial Activity of Secondary Metabolites from Halophilic *Bacillus pumilus* sp.*Int.J.Curr.Microbiol.Appl.Sci.* 3: 506-512.
- Singh SK, Tripathi VR, Jain RK, Vikram S and Garg SK. (2010). An antibiotic, heavy metal resistant and halotolerant *Bacillus cereus* SIU1 and its thermoalkaline protease. *Microb Cell Fact.* 10.1186/1475-2859-9-59 .
- Stan-Lotter H, Doppler E, Jarosch M, Radax C, Gruber C and Inatomi K. (1999). Isolation of a chymotrypsinogen B-like enzyme from the archaeon *Natronomonas pharaonis* and other halobacteria.*Extremophiles*.3: 153–161.
- Stepanov V M, Rudenskaya G N, Revina L P, Gryaznova Y B, Lysogorskaya E N, Filippova I Y, Ivanova I I. (1992). A serine proteinase of an archaeobacterium *Halobacterium mediterranei* homologue of eubacterial subtilisins. *Biochem. J.* 285: 281–286.
- Studdert CA, De Castro RE, Seitz K H and Sanchez J J.(1997).Detection and preliminary characterization of extracellular proteolytic activities of the haloalkaliphilic archaeon *Natronococcus occultus*. *Arch. Microbiol.*168: 532–535.
- Tapingkae W, Tanasupawat S, Itoh T, Parkin KL, Benjakul S, Visessanguan W and Valyasevi R. (2008). *Natrinema gari* sp. nov., a halophilic archaeon isolated from fish sauce in Thailand.*Int. J. Syst. Evol. Microbiol.* 58: 2378–2383.
- Thongthai C and Suntinanalert P. (1991). Halophiles in Thai fish sauce (nam pla) In: Rodriguez-Valera F, editor. General and applied aspects of halophilic microorganisms. *New York, N.Y: Plenum Press.*381–388.
- Thongthai C, McGenity T J, Suntinanalert P and Grant W D. (1992). Isolation and characterization of an extremely halophilic archaeobacterium from traditionally fermented Thai fish sauce (nam pla).*Lett. Appl. Microbiol.* 14: 111–114.
- Todkar S, Todkar R, Kowale L, Karmarkar K and Kulkarni A. (2012). Isolation and Screening of Antibiotic producing Halophiles from Ratnagri coastal area, State of Maharashtra. *International Journal of Scientific*

- and Research Publications*. 2:2250-3153.
- Tolan J S and Foody B. (1999). Cellulase from submerged fermentation. In: Tsao GT (ed.). Recent Progress in Bioconversion of Lignocellulosics. Springer-Verlag, Berlin. *Adv. Biochem. Eng. Biotechnol.* 41-67.
- Ventosa A and Nieto J J. (1995). Biotechnological applications and potentialities of halophilic microorganisms. *World J. Microbiol. Biotechnol.* 11:85-94.
- Vreeland R H, Piselli Jr A F, McDonnough S and Meyers S S. (1998). Distribution and diversity of halophilic bacteria in a subsurface salt formation. *Extremophiles*. 2: 321-331.
- Ward D M and Brock T D. (1978). Hydrocarbon biodegradation in hypersaline environments. *Appl. Environ. Microbiol.* 35:353-359.
- Woolard C R and Irvine R L. (1992). Abstracts of the Annual Water Environmental Federation Conference. Biological treatment of hypersaline wastewater by a biofilm of halophilic bacteria. *Abstr.* 14.
- Yakimov M M, Golyshin P N, Lang S, Wagner F, Moore E, Abraham W R and Timmis K N. (1996). Abstracts of the First International Congress on Extremophiles. New moderate halophilic marine strain MM1 produces novel class of biosurfactants. *Abstr.* 182.