

## Original Research Article

# Effect of Exogenous Melatonin and Different photoperiods on Serum Glucose and Total Serum Protein levels in Chhotanagpuri Ewe.

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## ABSTRACT

Forty two (42) apparently healthy, non-pregnant, non-lactating Chhotanagpuri ewe, having body weight ranging between 14.11±0.09 to 15.38±0.06 Kg were selected and selected ewes were isolated from rams 2 months prior to melatonin administration. The selected animals were allocated randomly into seven groups viz. Group-I [Normal control], Group-II [Long day (LD) control], Group-III [LD + Melatonin administration orally (3mg/day)], Group-IV [(LD + Melatonin administration subcutaneously (1mg/day)], Group-V [Short Day (SD) control], Group-VI [SD + Melatonin administration orally (3mg/day)] and Group-VII [SD + Melatonin administration subcutaneously (1mg/day)] comprising six animals in each group. Rams were then introduced into each group after completion of exogenous administration of melatonin. Blood sample without anticoagulant in vials was collected from each animal day before the start of experiment and there after every month up to fifth month. After administration of melatonin, serum glucose was decreased in group- III, IV, VI and VII in comparison to group-I, although this decrease was not statistically significant. An increasing trend of serum glucose was observed in all groups from third to fourth month. Total protein at first month was significantly (P<0.05) higher in group VI in comparison to 0 day. After administration of melatonin, total serum protein concentration was significantly (P<0.05) decreased in group VI in comparison to first month and continued up to third month. At fifth month, total serum protein was significantly (P<0.05) lower in group III, IV and VI in comparison to 0 day and first month.

## Keywords

Chhotanagpuri ewe,  
Exogenous melatonin,  
Photoperiod,  
Serum glucose,  
Total serum protein

## Introduction

Sheep husbandry is backbone of rural economy in India. The Chhotanagpuri sheep is the only recognized breed of sheep found in Jharkhand with a few numbers in Bihar

and West Bengal. This breed is maintained solely for mutton production. They produce coarse wool of poor quality. This is being utilized for carpet manufacturing. Sheep

which has a vital economy in India are mostly raised under harsh environment condition as it is seasonal breeders (Sangeetha and Rameshkumar, 2014).

Melatonin (N-acetyl-5-methoxy-tryptamine) was first identified and isolated from bovine pineal extract in 1958 as a neuro-hormone which is synthesized and secreted mainly from the pineal gland (Chakravarty and Rizvi, 2008). Since its discovery, further investigation has revealed that it is also produced by several other organs like the gastrointestinal tract, brain, eye, lungs, skin, kidney, liver, thyroid, thymus and pancreas (Fernando and Rombauts, 2014). Melatonin is an indoleamine, which is synthesized from the essential amino acid, tryptophan (Reiter *et al.*, 2013). Its production is dependent on ambient illumination, with release being suppressed by light. The supra-chiasmatic nucleus (SCN) which is the major circadian oscillator that receives light input from the retina through the retino-hypothalamic tract is the one that regulate the circadian melatonin production (Berson *et al.*, 2002).

Circadian clocks govern the timing of development, behavior, physiology, endocrinology and biochemistry, as well as photoperiodic events (Forster *et al.*, 2001). Also, circadian clocks and energy metabolism are linked because mutation in clock gene leads to metabolic syndrome in mice (Turek *et al.*, 2005). The circadian clock reportedly regulates metabolism and energy homeostasis in the liver and other peripheral tissues by mediating the expression or activity of various metabolic enzymes and transport system involved in glycogen and glucose metabolism (Green *et al.*, 2008; Lafleur, 2003; Zhang *et al.*, 2006). In addition, Challet *et al.*, (2004) observed the blood glucose increased during exposure to light and decreased during darkness in

rats. A Study has demonstrated that constant darkness elevated 5' adenosine mono phosphate (5'AMP) in the blood of mice. Hence, 5'AMP is a pivotal metabolic signal whose circulatory level determines the balance of the peripheral organ energy supply between glucose and glycogen (Zhang *et al.*, 2006). It is known that insulin secretion is not affected by photoperiod in cattle (Zinn *et al.*, 1986). In mammals, SCN activity is modulated by several neurotransmitter or neuro-hormone including melatonin (Armstrong *et al.*, 1986). However, the results of studies investigating melatonin effect on glucose metabolism in experimental animal are controversial, such as increase of blood glucose in rats (Csaba and Barath, 1971) or, on the contrary, reduction of blood glucose in rats (Lizuka, 1996).

The major site of synthesis of the serum proteins is the liver. The second major site is the immune system consisting of the monocyte-macrophage system, lymphoid and plasma cells. Structural, functional, and enzyme proteins that are synthesized in all body cells and tissues are present in plasma in minor quantities as a result of cell turnover. A seasonal rhythm was found for the serum proteins in albumin and alb/glob for sheep with acrophase between July and September for sheep (Piccione *et al.*, 2011). These changes are connected to the changes of light and temperature throughout the year (Alila-Johansson *et al.*, 2004). In fact, the hypothalamic pacemaker, directly innervated from retina, results as the principle factor involved in determining circadian rhythms (Moore-Ede, 1986). This pacemaker regulates transcriptional process also, and, in particular, makes this like an autonomous pacemaker. On the basis of the results obtained by Piccione *et al.*, (2011) it is conceivable that albumin production from liver is strictly dependent on this circadian

clock and, may be, on melatonin release (Alila-Johansson *et al.*, 2001 ; Piccione *et al.*, 2008).

There is paucity of information regarding serum glucose and total serum protein status in Chhotanagpuri sheep. Therefore, a comprehensive study was designed to observe the base line study of serum glucose and total serum protein of indigenous ewe by exogenous administration of melatonin and also the effect of different photoperiod.

### **Materials and Methods**

The present study was conducted in Department of veterinary physiology, College of veterinary science and A.H., Birsa Agricultural University, Kanke, Ranchi-6, Jharkhand located at 23.36<sup>o</sup>N latitude and 85.33<sup>o</sup>E longitude with an altitude of 651 m above mean sea level. Design of experiment was approved by the Institutional Animal Ethics Committee vide letter no.-139/528/RVC/IAEC. Forty two (42) apparently healthy, non-pregnant, non-lactating Chhotanagpuri ewes, having body weight ranging between 14.11±0.09 to 15.38±0.06 Kg reared under uniform managerial husbandry practices were selected. Selected ewes were isolated from rams prior to melatonin administration. The selected animals were allocated randomly into seven groups (viz. group-I to group-VII) comprising six animals in each group. Group-I [Normal control]:- The animals in this group were exposed to normal variation in day length. Group-II [Long day (LD) control]:- In this group in addition to natural sun light, artificial light was provided to animal for maintaining 16-18 hours of light every day for the period of one month and considered as long day control. Group-III [LD + Melatonin administration orally (3mg/day)]:- In this group in addition to natural sun light, artificial light was

provided for maintaining 16-18 hours of light every day for the period of one month and after that 3 mg melatonin was administered orally for one month to each animal. Group-IV [(LD + Melatonin administration subcutaneously (1mg/day))]:- In this group in addition to natural sun light, artificial light was provided for maintaining 16-18 hours of light every day for the period of one month and after that 1 mg melatonin was administered subcutaneously for one month to each animal. Group-V [Short Day (SD) control]:- Animals in this group were provided only 8 hours natural day light and were then kept in a light-proof shed for 16 hours exposure in dark every day for the period of one month. This group served as short day control. Group-VI [SD + Melatonin administration orally (3mg/day)]:- Animals in this group were provided only 8 hours natural day light and were then kept in a light-proof shed for 16 hours exposure in dark every day for the period of one month and after that 3 mg melatonin was administered orally for one month to each animal. Group-VII [SD + Melatonin administration subcutaneously (1mg/day)]:- Animals in this group were provided only 8 hours natural day light and then were kept in a light-proof shed for 16 hours exposure in dark every day for the period of one month and after that 1 mg melatonin was administered subcutaneously for one month to each animal. Rams were introduced into each group after completion of exogenous administration of melatonin. Blood samples without anticoagulant in vials were collected from each animal day before the start of experiment and there after every month up to fifth month. Without anticoagulant added blood sample was allowed to clot at room temperature in centrifuge tubes for 2-3 hours. The clots was eased away from the edge of the vessel and allowed to shrink overnight at 4<sup>o</sup>C. The serum was pippered off, centrifuged at 3000

rpm for 10 minutes and stored at  $-20^{\circ}\text{C}$  for further analysis of biochemical parameters. Serum glucose and Total serum protein were estimated by Spectrophotometer by using GOD/POD method as described by Trinder (1969) and Biuret Method as described by Gornall *et al.*, (1949) respectively. Data were statistically analyzed as per methods described by Snedecor & Cochran (2004).

## Results and Discussion

Carbohydrate in the form of glucose is the principle source of energy for the life processes of the mammalian cells. All cells require a constant supply of this indispensable nutrient and only relatively small changes are tolerated without adverse effects on the health of the animal (Kaneko *et al.*, 1997). In present experiment, the value of serum glucose has been presented in Table-1. The mean of serum glucose was  $55.81\pm 1.96$ ,  $56.27\pm 2.08$ ,  $56.11\pm 2.11$ ,  $55.73\pm 2.35$ ,  $56.49\pm 2.72$ ,  $55.99\pm 2.37$  and  $55.67\pm 2.25$  mg/dl in group I – VII, respectively on day 0. After administration of melatonin, serum glucose was decreased in group III, IV, VI and VII in comparison to group I although this decrease was not statistically significant. An increasing trend of serum glucose was observed in all groups from third to fourth month.

At fifth month, serum glucose was decreased in all groups in comparison to fourth month, but this decrease was not statistically significant. Prunet – Marcassus *et al.*, (2003) reported, blood glucose was lower when, melatonin was administered. Kassayova *et al.*, (2006) reported, prolonged melatonin administration decreases serum glucose concentration in female rats. Mahmud and Mahmud (2013) also reported melatonin (120mg/kg diet) decreases the blood glucose, which is in agreement with our findings. However, the results of studies

investigating melatonin effect on glucose metabolism in experimental animal are controversial, such as increase of blood glucose in rat (Csaba and Barath, 1971) or, on the contrary, reduction of blood glucose in rats (Lizuka, 1996). Chaiyabutr *et al.*, (1982) reported during early and mid-pregnancy in goats the demand of fetus was minimal resulting little utilization while, the rate of glucose turnover increased in late pregnancy which support our finding. The values are within normal range as reported by several workers (Oshiro *et al.*, 1978 and Kaneko *et al.*, 1997). El-Sherif and Assad (2001) reported significant increased in blood glucose level in different stage of pregnancy did not agree with our findings.

Blood serum proteins serve important function in the body for maintaining colloid osmotic pressure, blood pressure, acid-base balance and transport of nutrients, hormones, enzyme etc. The major site of synthesis of the serum proteins is the liver and the second major site is the immune system consisting of the monocyte-macrophase system, lymphoid and plasma cells.

In present study, the value of total serum protein (g/dl) has been presented in Table-2. On 0 day, the mean of total protein estimated was  $8.04\pm 0.16$ ,  $8.04\pm 0.15$ ,  $7.85\pm 0.24$ ,  $7.92\pm 0.38$ ,  $8.06\pm 0.29$ ,  $8.07\pm 0.21$  and  $8.00\pm 0.28$  g/dl in group I – VII, respectively. At fifth month, total serum protein was significantly ( $P<0.05$ ) lower in groups III, IV and VI in comparison to 0 day and first month. A decreasing trend of total protein was observed in all remaining groups from second month to fifth month may be due to pregnancy, but this decrease was not statistically significant. Kassim *et al.*, (2008) observed the effect of length photoperiod on serum total protein and melatonin.

**Table.1** Serum glucose concentration (mg/dl) of Chhotanagpuri ewes in different groups at different periods (Mean ± SE)

	<b>0 Day</b>	<b>First Month</b>	<b>Second Month</b>	<b>Third month</b>	<b>Fourth Month</b>	<b>Fifth Month</b>
Group – I	55.81 ±1.96	55.80 ±1.88	55.84 ±1.90	56.87 ±1.88	57.63 ±1.95	56.80 ±1.87
Group – II	56.27 ±2.08	56.57 ±1.88	56.48 ±1.87	57.57 ±1.48	58.32 ±1.24	57.53 ±1.47
Group – III	56.11 ±2.11	56.33 ±1.91	55.57 ±1.92	57.62 ±1.95	59.14 ±1.94	57.45 ±1.95
Group – IV	55.73 ±2.35	56.04 ±2.26	55.19 ±2.22	56.89 ±2.30	58.15 ±2.42	56.81 ±2.31
Group – V	56.49 ±2.72	56.16 ±2.61	56.36± 2.61	57.15 ±2.47	57.68 ±2.46	57.09 ±2.44
Group – VI	55.99 ±2.37	55.75 ±2.24	54.81 ±2.24	56.94 ±2.18	58.45 ±2.19	56.71 ±2.14
Group – VII	55.67 ±2.25	55.31 ±2.31	54.34 ±2.21	55.76 ±2.23	56.80 ±2.30	55.68 ±2.24

Means did not vary significantly between the periods and groups.

**Table.2** Total serum protein concentration (g/dl) of Chhotanagpuri ewes in different groups at different periods (Mean ± SE)

	<b>0 Day</b>	<b>First Month</b>	<b>Second Month</b>	<b>Third month</b>	<b>Fourth month</b>	<b>Fifth Month</b>
Group – I	8.04 <sup>a</sup> ±0.16	8.05 <sup>ABa</sup> ±0.17	7.96 <sup>ab</sup> ±0.14	7.58 <sup>ABab</sup> ±0.17	7.51 <sup>ABb</sup> ±0.19	7.48 <sup>ABb</sup> ±0.20
Group – II	8.04 ±0.15	8.01 <sup>AB</sup> ±0.29	7.93 ±0.22	7.55 <sup>AB</sup> ±0.19	7.48 <sup>AB</sup> ±0.21	7.42 <sup>AB</sup> ±0.22
Group – III	7.85 <sup>ab</sup> ±0.24	8.10 <sup>ABa</sup> ±0.25	8.06 <sup>a</sup> ±0.16	7.44 <sup>ABbc</sup> ±0.16	7.34 <sup>ABbc</sup> ±0.16	7.22 <sup>ABc</sup> ±0.16
Group – IV	7.92 <sup>a</sup> ±0.38	7.84 <sup>Ba</sup> ±0.21	7.71 <sup>ab</sup> ±0.26	7.28 <sup>Bab</sup> ±0.19	7.21 <sup>Bab</sup> ±0.19	7.12 <sup>Bb</sup> ±0.17
Group – V	8.06 ±0.29	8.02 <sup>AB</sup> ±0.29	7.99 ±0.23	7.81 <sup>A</sup> ±0.20	7.76 <sup>A</sup> ±0.21	7.72 <sup>A</sup> ±0.22
Group – VI	8.07 <sup>b</sup> ±0.21	8.11 <sup>Aa</sup> ±0.23	8.04 <sup>b</sup> ±0.15	7.56 <sup>ABc</sup> ±0.13	7.45 <sup>ABc</sup> ±0.13	7.34 <sup>ABc</sup> ±0.13
Group – VII	8.00 ±0.28	8.01 <sup>AB</sup> ±0.20	7.98 ±0.16	7.65 <sup>AB</sup> ±0.17	7.56 <sup>AB</sup> ±0.19	7.49 <sup>AB</sup> ±0.20

Means bearing different superscript vary significantly (p<0.05) within the groups (a, b, c) and between the groups (A, B).

Melatonin hormone concentration recorded sharp decline, when buffalo heifer was exposed to long photoperiod, but total serum

protein did not show any significant difference like our finding. Oqetark *et al.*, (2004) reported melatonin treatment did not

cause a significant change in total protein levels in rat. The similar pattern was observed in our finding. Kumar (2003), Batavani *et al.*, (2006) and Verma (2012) reported decline in the protein level during pregnancy in sheep and goat, which are in agreement with our findings. The values obtained in the present experiment are within the normal range as reported by Dukes (1970), Somvanshi *et al.*, (1989) and Dutta *et al.*, (1996). Kumar (2003) reported the total serum protein concentration decrease, due to synthesis of hormones which is produced during gestation period and also due to demand of protein by growing fetus.

## References

- Alila-Johansson, A., Eriksson, L., Soveri, T. and Laakso, M.L. 2001. Seasonal variation in endogenous serum melatonin profiles in goats: a difference between spring and fall. *J. BiolRhythms.*, 16: 254-263.
- Alila-Johansson, A., Eriksson, L., Soveri, T. and Laakso, M.L. 2004. Daily and annual variation of free fatty acid, glycerol and leptin plasma concentrations in goats (*Capra hircus*) under different photoperiods. *Comp Biochem Physiol.*, 138: 119-131.
- Armstrong, S.M., Cassone, V.M., Chesworth, M.J., Redman, J.R., and Short, R.V. 1986. Synchronization of mammalian circadian rhythms by melatonin. *J. Neural. Transm. Suppl.*, 21: 375-394.
- Batavani, R.A., Ansari, M.H. and Asri, S. 2006. Concentrations of serum total protein and protein fractions during diestrus and pregnancy in Makuuii ewes. *Comparative clinical pathology*, 15: 227-230.
- Berson, D.M., Dunn, F.A. and Takao, M. 2002. Photo-transduction by retinal ganglion cells that set the circadian clock. *Scienc*, 295: 1070-1073.
- Chaiyabutr, N., Anne Faulkner and Peaker, M. 1982. Glucose metabolism in vivo in fed and 48 hours starved goats during pregnancy and lactation. *Br. J.Nutr.*, 47: 87-94.
- Challet, E., Malan, A., Turek, F.W. and Van Reeth, O. 2004. Daily variation of blood glucose, acid-base state and Pco<sub>2</sub> in rats: effect of light exposure. *Neurosci. Lett.*, 355: 131-135.
- Csaba, G. and Barath, P. 1971. Are langerhans islets influenced by the pineal body. *Experientia*, 27: 962.
- Dukes, H.H. 1970. Physiology of domestic animals, 8<sup>th</sup> edn. London, Comstock.
- Dutta, A., Surmah, S. and Rajkhowa, N.K. 1996. Haematological and biochemical studies in sheep of Assam. *Indian Vet. J.*, 73:402-405.
- EI-Sherif, M.M.A. and Assad, F. 2001. Changes in some blood constituents of Barki ewes during pregnancy and lactation under semi-arid conditions. *Small Ruminant Research.*, 40(3): 269-277.
- Fernando, S., and Rombauts, L. 2014. Melatonin: shedding light on infertility- a review of the recent literature. *Journal of ovarian research*, 7: 98.
- Forster, C.H., Winter, C., Hofbauer, A., Hall, J.C. and Stanewsky, R. 2001. The circadian clock of fruit flies is blind after elimination of all known photoreceptor. *J. Neuron*, 30: 249-261.
- Gornall, A., Bardawill, C. and David, M. 1949. Determination of serum proteins by means of the biuret reaction. *J. Biol. Chem.*, 177: 751 - 766.
- Green, C.B., Takahashi, J.S. and Bass, J. 2008. The meter of metabolism, *Cell*, 134: 728-742.
- Kaneko, J.J., Harvey, J.W. and Bruss, M.L. 1997. Clinical biochemistry of

- domestic animals. 5<sup>th</sup> edition, Harcourt Brace & Company, Singapore.
- Kassayova, M., Markova, M., Bojkova, B., Adamekova, E., Kubatka, P., Ahlersova, E. and Ahlers, I. 2006. The influence of long term melatonin administration on basic physiological and metabolic variables of young Wistar: Han rats. *Biologia*, Bratislava., 63(3): 313-320.
- Kassim, N.S.I., Afify, A.A. and Hoda, Z.H. 2008. Effect of photoperiod length on some reproductive traits and hormonal profiles in buffalo heifers. *American-Eurasian J. Agric. & Environ. Sci.*, 3(4): 646-655.
- Kumar, M. 2003. Induction of estrus in Anestrus goats. M.V.Sc. Thesis submitted to Birsa Agri. Univ., Ranchi, Jharkhand.
- Lafleur, S.E. 2003. Daily rhythms in glucose metabolism: suprachiasmatic nucleus output to peripheral tissue. *J. Neuroendocrinol.*, 15: 315-322.
- Lizuka, Y. 1996. Effect of melatonin on serum glucose and insulin levels in rats. *Med.Biol.*, 133: 65-67.
- Mahmud, S.A. and Mahmud, A.M.R. 2013. Physiological effects of melatonin on leptin, testosterone and biochemical parameters in Albino rats. *IOSR Journal of Pharmacy*, 3(4): 48-53.
- Moore-Ede, M.C. 1986. Physiology of the circadian timing system: predictive versus reactive homeostasis. *Am J Physiol Regul Integr Comp Physiol.*, 250: 737-752.
- Oqetark, M., Kus, I., Kavaki, A., Zararsiz, I., Iihan, N. and Sarsilmaz, M. 2004. Effect of melatonin on carbontetrachloride-induced changes in rat serum. *J. Physiol. Biochem.*, 60(3): 205-210.
- Oshiro, S., Shinjo, A. and Takahasi, H. 1978. Comparative studies on the plasma composition of buffaloes, cattle and goats. *Science Bull Coll. Agri. Univ. Ryukyus, Okinawa*, 25: 383-387.
- Piccione, G., Fazio, F., Caola, G. and Refinetti, R. 2008. Daily rhythmicity of glycemia in four species of domestic animals under various feeding regimes. *J Physiol Sci.*, 58: 271-275.
- Piccione, G., Messina, V., Giannetto, C., Casella, S., Assenza, A. and Fazio, F. 2011. Seasonal variations of the serum proteins in sheep and goats. *Archiv Tierzucht*, 54(4): 399-405.
- Prunet-Marcassus, B., Desbazeille, M., Bros, A., Louche, K., Delagrang, P., Renard, P., Casteilla, L. and Penicaud, L. 2003. Melatonin reduces body weight gain in sprague dawley rats with diet induced obesity. *Endocrinology*, 144(12): 5347-5352.
- Reiter, R., Rosales-Corral, S., Manchester, L. and Tan, D. 2013. Peripheral reproductive organ health and melatonin: ready for prime time. *Int. J. Mol. Sci.*, 14: 7231-7272.
- Sangeetha, P. and Rameshkumar, K. 2014. Observation of biochemical variations in sheep faeces during different reproductive phases. *Res.J.Animal, Veterinary and Fishery Scie.*, 2(2): 13-16.
- Snedecor, G.W. and Cochran, W.G. 2004. *Statistical method* 8<sup>th</sup> edn. The Iowa State University.
- Somvanshi, R., Biswas, J.C., Sharma, B. and Koul, G.L. 1989. Haematological studies on Indian pashmina goats. *Res.Vet.Sci.*, 42(I): 124-126.
- Trinder, P., 1969. *Ann. Clin. Biochem.* 6: 24.
- Turek, F.W., Joshu, C., Kohsaka, A., Line, E., Ivanova, G., McDearmorn, E., Lapoky, A., Losee-olson, S., Easton, A., Jensen, D.R., Eckel, R.H., Takahashi, J.S. and Bass, J. 2005.

- Obesity and metabolic syndrome in circadian clock mutant mice. *Science*, 308:1043-1045.
- Verma, R.K. 2012. Effect of temperature variation on reproductive hormones and blood biochemical profile during various states of reproduction in Chhotanagpuri ewes, Ph.D. Thesis submitted to Birsa Agricultural University, Kanke, Ranchi, Jharkhand.
- Zhang, J., Kaasik, K., Blackburn, M.R. and Lee, C.C. 2006. Constant darkness is a circadian metabolic signal in mammals. *Nature*, 439: 340-343.
- Zinn, S.A., Purchas, R.W., Chapin, L.T., Petittree, D., Markel, R.A., Bergen, W.G. and Tucker, H.A. 1986. Effect of photoperiod on growth, carcass composition, prolactin, growth hormone and cortisol in pre-pubertal and post-pubertal Holstein heifers. *J.Anim. Sci.*, 63: 1804-1815.