

Original Research Article

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Histopathological Changes in the Gills Associated with *Dactylogyrus* spp. Infestation in Orange-spotted Grouper, *Epinephelus coioides* Reared in Sea Cages

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ABSTRACT

Keywords

Orange spotted grouper, Cage culture, Histopathology, Flukes, *Dactylogyrus* spp.

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A study was conducted to investigate the histological changes in the gills of orange-spotted grouper infested with flukes of *Dactylogyrus* spp. Several histological changes were observed in the gill which included complete fusion of secondary lamellae, dilatation of the venous sinus at the middle of the primary lamellae, oedema and complete loss of secondary lamellae, etc.

Introduction

E. coioides is a potential species for mariculture in India. These are generally cultured in floating net cages or earthen ponds (FAO, 2010). It is also an important food fish for livelihoods of small and large-scale coastal fish farmers. Groupers are considered as a high-value species with a high market

demand in foreign countries (FAO, 2010). At the same time due to intensification or high stocking densities of *E. coioides* in cage culture cause stressful conditions and it may lead to expression of diseases in *E. coioides*. Fish under normal condition is in a state of equilibrium with its environment and infectious organisms. Any alterations in the environment, which will disturb equilibrium

state, subsequently it may cause stress to the fish and they become more vulnerable to disease producing organism.

Disease is one of the major constraints to intensification of any culture. In improved farming systems, nursery operators have a tendency to overstock fish, which ultimately causes disease. High stocking densities of fry and fingerlings during nursery operation generally increases stress, and subsequently, fish become more susceptible to infectious disease (Snieszko, 1974).

Thousands of fish parasite species are already known and many more remain to be discovered. However very few are seriously harmful to their host. The presence of dense populations of fish kept in particular environmental conditions may favor certain parasites, necessary to cause harm to a fish, it varies considerably with the species and size of the fish and its health status (Ramudu, 2013). The major infectious agents are associated with gill, most of the pathogens are passing through gill and will enter into the host. Among the all parasites, monogenean parasites are the most important pathogens to cause severe stress in *E. coioides*, in addition to that reduce the growth and when it become more and more severe, in the last stage it can be leads to mortality, which will be resulted in serious economic losses in farmer. The disease caused by monogenean parasites, causes serious problems in aquaculture reported so many authors (Okamoto, 1963; Ogawa and Inouye, 1997; Yoshinaga *et al.*, 2000, 2001, 2009; Mushiake *et al.*, 2001; Nakayasu *et al.*, 2002) with an obvious pathogenicity.

Various methods are used to assess the health status of aquatic organisms. Among them, the parasitological and histopathological diagnoses are presented as tools for analysis and prevention of diseases (Resende, 2009;

Sampaio *et al.*, 2013). Histopathological study is a good indicator of fish health status, and the relevance of each lesion depends on how it affects organ function and the fish's ability to survive (Bernet *et al.*, 1999). In this context, the present study focuses on histopathological investigations which have been proved to be a sensitive tool to detect histopathological changes in gill of *E. coioides* due to infestation of *Dactylogyrus* spp. parasites.

Materials and Methods

A total number of 15 freshly dead or ailing *E. coioides* (4.7-8.4cm length; 5.2-8.7g weight,) were collected from marine cages during the month of March, 2018 and transported to the laboratory for further analysis. Swabs were taken aseptically from kidney, spleen and liver and streaked onto Zobells marine agar (ZMA; Himedia, India), Thiosulfate Citrate Bile salt Sucrose agar (TCBS), Himedia, India) and brain heart infusion agar and incubated at 28 °C for 48 h. Further, organs like gills, skin, fins, intestine and kidney were examined under microscope for the presence of parasites. Water temperature, salinity, pH were measured at weekly intervals using portable instruments. Ammonia 0.02 mg/L, Nitrite 0.01 mg/L and Nitrate 0.01mg/L) was estimated according to procedure framed by APHA (1998). Representative samples of gills were fixed in 10% NBF, dehydrated in series of alcohol, cleared in xylene, embedded in paraffin wax and sectioned (6 μ) using a rotary microtome. The sections were stained with using Hematoxylin and Eosin, micro photographed (Roberts, 2001).

Results and Discussion

The infected fishes were lethargic, off feed, surfacing and with erratic swimming. Gills were pale in colour with excessive mucus.

Liver and other internal organs were normal grossly. No bacteria could be grown on plates after an incubation at 28°C for 48 hours.

The present study revealed the presence of *Dactylogyrus* spp. in the gills of all the *E. coioides* examined. *Dactylogyrus* spp. were identified from the gill filaments of *E. coioides* with an overall prevalence of 100%. The gill lesions are summarised in Table 1.

Briefly, the microscopy gill lesions showed severe morphological changes such as hyperplasia of secondary lamellae and their fusion, dilatation of the venous sinus at the middle of the primary lamellae, oedema (Figure 1B), complete loss of secondary lamellae due to attachment of parasite with marginal hooks (Figure 1C), vacuolation at the base of the primary lamellae (Figure 1D), telangiectasis of secondary lamellae (Figure

1E), and hypertrophy of the mucus cells and necrosis (Figure 1F). Curling of the secondary lamellae, degenerative and necrotic changes in the epithelium of gill filaments (Figure 2A), vacuole formation at the top of the secondary lamellae due to detachment of epithelial cells of secondary lamella (Figure 2B and F) were also observed in gill histology. Hypertrophy of the blood cells and vacuole formation at base of the secondary lamellae replacing the chloride cells was also observed (Figure 2C). Hyperplasia of secondary lamellar epithelial cells due to parasite attachment to the gill and dilatation of the venous sinus observed at the middle of the primary lamellae (Figure 2D). Oxygenated blood flow in to the primary lamellae and hypertrophy of blood cells and Hyperplasia of secondary lamellar epithelial cells (Figure 2E) has been found.

Table.1 Histopathological alterations of gill tissue due to *Dactylogyrus* spp. of *E. coioides*, the histopathological alterations were represented in symbol based on their severity.

Gill tissues morphological Anomalies	Normal gill tissue	<i>Dactylogyrus</i> spp. infested gill tissue
Lamellar fusion	-	++
Completely Loss of Secondary lamella	-	++
Gill lamellar hyperplasia	-	+
Degenerative and necrotic changes in the epithelium of gill filaments	-	+
Oedema	-	+
Vacuolization in gill filament base	-	++
Haemorrhage at the base of the secondary lamella	-	+
Epithelial lifting	-	++
Telangiectasis	-	+
Blood congestion	-	+
Dilatation of the venous sinus, observed at the Middle of the primary lamellae	-	+
Hypertrophy of Mucus cells	-	+
Hypertrophy of Blood cells	-	++
Curling	-	+

Note: The anomalies visible in the gill morphology were divided based on the severity into three grades (-) no anomalies; (+) anomalies in <20% of the fields; (++) anomalies in <20–60% of the fields.

Figure.1 Photomicrographs of the gill tissue infested with *Dactylogyrus* spp. parasites (P); A. Photomicrographs of the normal gill tissue (H and E ;400x) showing Gill filament with primary lamellae (PL) and Secondary lamellae (SL), Mucus cells (MC) and Chloride cells (CC); B. Dilatation of the venous sinus observed at the Middle of the primary lamellae (DVS); Oedema (ED), Complete fusion of secondary lamellae (FSL) (H and E;100x); C. Completely loss of secondary lamellae (LSL) due to attachment of parasite with marginal hooks (MH) (H and E; 400x); D. Vacuole (VC) formation at base of the primary lamellae; with higher magnification showing secondary lamellar epithelial hyperplasia (HP) with secondary lamellar fusion (FSL) (H and E;400x); E. Telangiectasia of secondary lamellae (TG, H and E;400x); F. Hypertrophy of the Mucus cells (HT); Necrosis (N) observed at the apex of all secondary lamellae (H and E;400x).

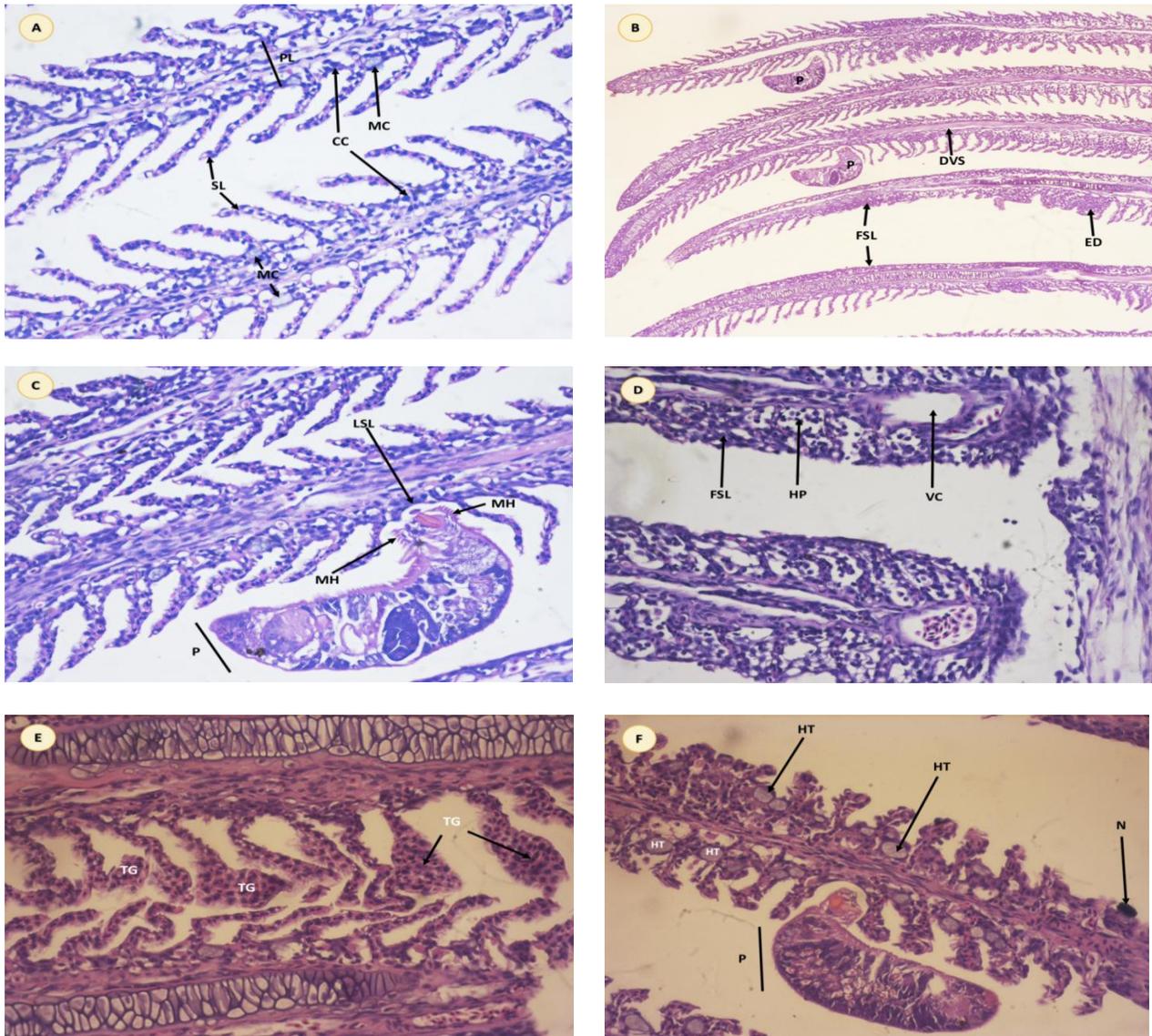
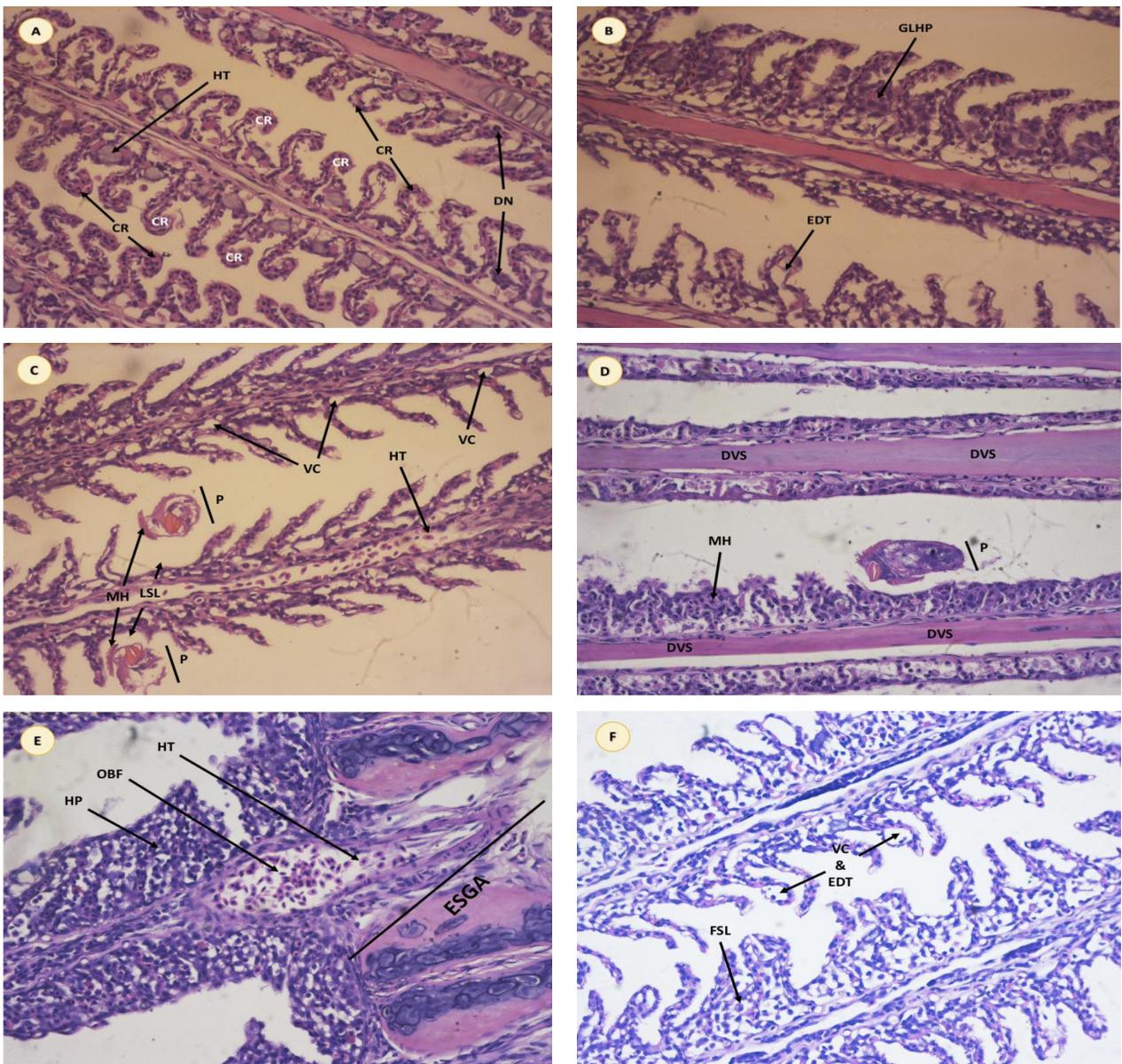


Fig.2 Photomicrographs of the gill tissue infested with *Dactylogyrus* spp. parasites (P); A. Curling of the secondary lamellae (CR) and Hypertrophy of the Mucus cells (HT), Degenerative and necrotic changes in the epithelium of gill filaments (DN)(H and E;400x); B. Gill lamellar hyperplasia (GLHP) and epithelial detachment (EDT)(H and E;400x); C. Hypertrophy of the blood cells (HT); Vacuole (VC) formation at base of the secondary lamellae in replace of Chloride cells, Completely loss of secondary lamellae (LSL) due to attachment of parasite with marginal hooks (MH) (H and E;400x). D. Hyperplasia (HP) of secondary lamellar epithelial cells due to parasite attachment to the gill and Dilatation of the venous sinus observed at the Middle of the primary lamellae (DVS) (H and E;400x). E. Oxygenated blood flow (OBF) in to the primary lamellae and hypertrophy (HT) of blood cells and Hyperplasia (HP) of secondary lamellar epithelial cells (H and E;400x). F. Vacuole (VC) formation at the top of the secondary lamellae due to detachment of epithelial (EDT) cells of secondary lamella



Histopathological study of the gills revealed a typical gill arch with complete loss of secondary lamellae due to attachment of parasite with marginal hooks. The parasite normally attaches to the gill lamellae of the host by inserting marginal hooklets (Turkayoztuk and Ahmet ozer, 2014) into the epithelium. Feeding activities of the parasites is associated with injuries to the epithelial cells of gill lamellae. The tissue alterations in the present study can be comparable to previous studies (Woo and Buchmann (2012) and Raissy and Ansari, 2011). Exacerbated swelling of gill arch with hyperplasia of secondary lamellar epithelial cells is due to parasite attachment to the gill, which is congruent with previous studies (Ramudu and Dash, 2015); Raissy and Ansari, 2011). The mucus cells of gill respond to the parasites with abnormal production of mucus and due to this hypertrophy of the mucus cells occurs as reported by Ramudu and Dash (2015) in Indian Major Carps and Jithendran *et al.*, (2005) in the gill of *Epinephelus tauvina* due to monogenean infestation. Vacuole formation at base of the gills e secondary lamellae in the present study could be due to the absence or reduced numbers of chloride cells as appreciated by histopathology.

Parasitic infestations, especially those associated with *Dactylogyru*s spp., are the most devastating in nature affecting the morphology of the gills and physiological functions of the fish in general and gills in particular (Roberts, 2012). Present study revealed dilatation of the venous sinus at the middle of the primary lamellae. Similar abnormalities were found in Indian major caps with multiple infestation of parasites as reported by Ramudu and Dash (2015).

At the apex of secondary lamellae, gill exhibited necrosis which possibly leads to suffocation these were similar with Leong, (2001). Vacuole formation at base of the

primary lamellae; with higher magnification showing secondary lamellar epithelial hyperplasia along with secondary lamellar fusion. In addition, telangiectasia were also observed in secondary lamellae of in gill histology, similar abnormalities in the gills were observed in fishes exposed to lead acetate in *Cyprinus carpio* reported by Mustafa *et al.*, (2017) and Dezfuli *et al.*, (2007) reported same anomalies in gill of European sea bass, *Dicentrarchus labrax* due infestation of *Diplectanum aequans*.

Curling of the secondary lamellae, Degenerative and necrotic changes in the epithelium of gill filaments, gill lamellar hyperplasia, vacuole formation at the top of the secondary lamellae due to detachment of epithelial cells of secondary lamella was observed in gill histology of *E.coioides*, similar anomalies were also observed in gills of *Cyprinus carpio*, here fishes were exposed to antimalarial drug chloroquine (CQ) studied by Ramesha *et al.*, (2018).

It is concluded in the present study that most of the *Dactylogyru*s spp. (Gill flukes) were found on the gill filaments and lamellae of the *E. coioides*. Several histological alterations were observed in the gill, which was shown above. However, destruction of the gill filaments and lamellae has been recorded. Blood and mucus was feeding by the parasite and which leads to hyperplasia and destruction of the epithelial cells which leads to reducing the surface area for oxygen exchange for effective respiration. Such pathological changes can also cause severe stress in the fish, or can even lead to death due to lack of oxygen. Gill damage could result in loss of gill surface area for respiration, which would lead to suffocation particularly at high water temperatures in the early morning. The failure of osmoregulation is also an important issue, since the parasite piercing its marginal hooks in to the gill

filaments and lamellae, finally exposes the surface of the gill to natural environment. According to the present study, the histopathological changes induced by the infestation of *Dactylogyrus* spp. would adversely affect the proper functioning of the gills of these infected fish.

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