Original Research Article  

Different Garden Pea Varieties Health Status Analysis Procured from Various Locations of Odisha

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A B S T R A C T

Garden pea (Pisum sativum L.) is an important vegetable crop grown during winter season in Odisha. A number of seed borne pathogens affect seed which reduce seed quality and cause seed borne diseases in the field, adversely affecting yield and quality of produce. A study on thirty-six farmers’ seed samples collected from eight districts to analyse the health status of Garden pea were found to possess seed germination 10-15% which is below IMSCS (i.e. 75%) and carried 40.1-46% fungal count on seeds. Aspergillus flavus, A. niger, Fusarium sp. and Pencillium sp. were fungi frequently associated with seeds which caused 24.4-55.6% reduction in germination and caused rotting of seed and seedling shoot and root.

Keywords
Garden pea, Germination, Health status, Seed, Fungi, Pathogenicity

Introduction

Garden pea (Pisum sativum L.) is an important vegetable crop grown for its green pods and seeds. It is a frost-hardy, cool-season, nutritious leguminous vegetable which is widely cultivated throughout the world. In India, it is grown as a winter vegetable in the plains of north India and as a summer vegetable in the hills. It is a rich source of protein (25%), amino acids, sugars (12%), carbohydrate, vitamins A and C, calcium and phosphorus, apart from having a small quantity of iron. Incidence of disease outbreak is one of the major biotic factor responsible for low yield. Seed and seedling disease, root rot and stem rot disease caused by fungi like Fusarium spp, Rhizoctonia solani, Sclerotium rolfsi, Pythium species and powdery mildew are important disease of pea in India which affects the crop every year. However, lack of quality seed continues to be one of the greatest impediments bridging the vast yield gap. Therefore, to approach the potentially realizable yield of a cultivar, production and distribution of quality seed is essential. Present study was carried out to critically analyse quality status of garden pea.
seeds used by farmers in Odisha with special references to know the seed health status.

Materials and Methods

Collection of seed samples

The seeds of different garden pea varieties grown during 2013-14 winter season were collected from farmers of different locations. The seeds were collected from local market in properly labelled polythene packets along with information on variety, month of harvest, storage container, storage place etc. and stored in seed cabinet for further use.

Study of Seed health status of collected samples

The seeds collected from the farmers were first visually observed to record the visible symptoms of diseases like patches of discoloration, mycelial mats and fungal fructifications attached to it, etc. To ascertain the association of fungal pathogens with the sample, the seeds were subjected to seed health test following standard moist blotter method (ISTA, 1985). For this test, clean and sterilized 11cm diameter plastic petridishes were used. Three pieces of filter paper discs were dipped in sterile distilled water and fitted to the base of the bottom Petridish. The seeds were placed equidistantly from each other on the moist blotter maintaining ten seeds per plate. The lid was placed on it and properly labelled. The plates were incubated inside BOD incubator at 25°C for seven days. Observations were recorded on number of seeds showing growth of fungal colonies on them and percentage of infected seeds was calculated.

Isolation and identification of fungi associated with the seed

The fungi associated with the infected seeds were isolated in pure culture following standard methods of isolation. The fungi developing on seeds in moist blotters were transferred to the PDA slants aseptically. Further purification was made by single spore isolation following dilution plate method.

The colonies developing on the slants were examined microscopically to further ascertain their purity. The pure cultures thus obtained were maintained on PDA slants by sub culturing at 15 days interval.

The fungi isolated from the seeds were identified by taking observations on their colony characters, mycelia characters, colour, size and shape of spores and conidiophores. The characters were compared with description in standard literature for confirmation of identification (Barnett, 1960, Dube, 1983).

Pathogenicity test of isolated fungi

Four pathogens isolated from the collected seed samples were selected for testing their pathogenicity under greenhouse condition.

Apparently healthy seeds of pea var. Mayurbhanj local free from any blemishes or disease symptoms were surface sterilized. Spore suspension of the test fungi were prepared having 10⁸ spores per ml. Surface sterilised seeds were soaked in equal volume of spore suspension separately for fifteen minutes. Inoculated seeds were sown in sterilized sand taken in sterilized plastic pots of 5 kg capacity at the rate of ten seeds per pot.

Observation was recorded on seedling emergence and occurrence of seed rot and seedling blight diseases. The plants were grown up to 30 days and then gently uprooted to observe symptoms on roots. The trial was replicated thrice.
Results and Discussion

Health status of seed samples collected from different places of Odisha

Thirty six Garden pea seed samples of eight varieties were collected from eight districts of Odisha. The colour of seed varied from yellow to different shades of green. Seed germination of samples was 60-65% (Table-1) which was below IMSCS (i.e. 75%). Visible symptoms of seed infection were observed as different types of seed discolouration (Fig-1a, b, c) and percentage of discolouration was calculated. The seed discolouration was observed as appearance of dark brown, light brown, black and white patches on seed coat. Seed discolouration varied from 11.0% to 15.0%. The lowest was in variety Jawahar pea of Koraput district and the maximum was in VL-matter of Bargarh district and local variety of Mayurbhanj district.

The results indicated that seed with higher seed discoloration had lower germination. Samples with higher seed discoloration also contained higher total fungal count with seed. Lowest seed discoloration was observed in variety Kasishakti of Khurdha and local variety of Kandhmal (9% each) which had highest germination (66 and 67%) and lowest total fungal count (40% and 41.7%) respectively. Highest seed discoloration was in variety VL-matter of Bargarh (16.5%) with lowest germination (60%) and highest total fungal count (40.1% to 46.5%).

Association of fungi like species of Aspergillus, Pencillium and Fusarium with pea seeds have been reported earlier by Begum et al., (2008) and Narayan et al., (2013), and association of Alternaria of sps. have been reported by Javied and Anjum (2006), Marcinkowska (2008) and Wilmanet al.,(2014).

Pathogenicity of isolated fungi

Pathogenicity of isolated fungi Aspergillusflavus, A. niger, Pencillium sp. and Fusarium sp. were proved under green house condition (Table-2,). Through seed inoculation all the fungi infected the seed and caused seed rot, seedling blight, root rot symptoms and reduced germination. Germination in uninoculated control seeds was 90%. It was reduced to 40% which was 55.6% less than that in control. Minimum of 24.4% reduction in germination was due to Pencillium sp. Seedling blight symptoms appeared 10 days after sowing.

There was black discoloration on stem at soil level and on cotyledons. Maximum seedling blight was due to Aspergillusflavus (14%) followed by A. niger (10.5%) and Fusarium sp. (9.2%). Blighted seedling often showed black rotting of root tips. Root rot was maximum by Aspergillusniger and Fusarium sp. (20.2 and 20.0%, respectively). Pencillium sp. didnot cause any seedling blight or root rot. Seedlings in control pots were normal and healthy.
### Table 1: Health status of seed samples collected from different places of Odisha

<table>
<thead>
<tr>
<th>Place of collection</th>
<th>Variety</th>
<th>No. of sample collected</th>
<th>Seed colour</th>
<th>Germination (%)</th>
<th>Seed discoloration</th>
<th>Fungi associated (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>A. f  A. n Pen. sp.  F. sp.  Alt. sp.  TFC</td>
</tr>
<tr>
<td>Keonjhar</td>
<td>Rachna</td>
<td>5</td>
<td>Bluish green</td>
<td>65</td>
<td>10.5</td>
<td>12.5  15.0  6.0  10.3  2  41.7</td>
</tr>
<tr>
<td>Mayurbhanj</td>
<td>Local</td>
<td>4</td>
<td>Bluish green</td>
<td>65</td>
<td>10.0</td>
<td>13.8  15.5  4.0  10.5  00  43.8</td>
</tr>
<tr>
<td>Kandhmal</td>
<td>Local</td>
<td>5</td>
<td>Green</td>
<td>67</td>
<td>9.0</td>
<td>13.5  14.2  5.0  9  00  41.7</td>
</tr>
<tr>
<td>Baragarh</td>
<td>Vl-matar</td>
<td>5</td>
<td>Yellow</td>
<td>60</td>
<td>16.5</td>
<td>12.8  13.5  6.0  9.5  00  45.8</td>
</tr>
<tr>
<td>Koraput</td>
<td>Jawahar pea</td>
<td>4</td>
<td>Green</td>
<td>65</td>
<td>10.0</td>
<td>10.5  19.0  5.5  9  2.0  46.0</td>
</tr>
<tr>
<td>Balangir</td>
<td>Local</td>
<td>4</td>
<td>Light green</td>
<td>61</td>
<td>15.0</td>
<td>12.2  15.0  5.2  9.5  00  41.9</td>
</tr>
<tr>
<td>Khurdha</td>
<td>Kasha shakti</td>
<td>4</td>
<td>Green</td>
<td>66</td>
<td>9.0</td>
<td>14.4  11.9  6.0  7.8  00  40.1</td>
</tr>
<tr>
<td>Sambalpur</td>
<td>GS-10</td>
<td>5</td>
<td>Green</td>
<td>64</td>
<td>10.5</td>
<td>11.2  13.3  5.5  8.5  2.0  40.5</td>
</tr>
</tbody>
</table>

* *A. f-Aspergillusflavus, A. n- Aspergillusniger, Pen. sp.-Penciliumsp, Alt. sp- Alternariasp, F. Sp – Fusariumsp, TFC-Total fungal count*

### Table 2: Pathogenicity test of isolated fungi of garden pea

<table>
<thead>
<tr>
<th>Fungi</th>
<th>Germination (%)</th>
<th>Seed rot (%)</th>
<th>Seedling blight (%)</th>
<th>Root rot (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Aspergillusflavus</td>
<td>50</td>
<td>50</td>
<td>14.0</td>
<td>14.3</td>
</tr>
<tr>
<td>Aspergillusniger</td>
<td>40</td>
<td>60</td>
<td>10.5</td>
<td>20.20</td>
</tr>
<tr>
<td>Penicillium sp.</td>
<td>68</td>
<td>32</td>
<td>00</td>
<td>00</td>
</tr>
<tr>
<td>Fusarium sp.</td>
<td>65</td>
<td>35</td>
<td>9.2</td>
<td>20.0</td>
</tr>
<tr>
<td>Un inoculated Control</td>
<td>90</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
</tbody>
</table>
**Fig. 1** (a, b, c) Different types of seed discolouration
The present findings are in conformity with results of earlier workers (Begum, 2004; El-Mohamedy and El-Baky, 2008; Sharma, 2011). Who have also reported seed and seedling diseases caused by these seed borne fungi.

Seed borne fungi caused reduction in germination. Many of them become active when seeds are sown, where they result in seed decay, root rot and post emergence seedling blight. Consequently, this results in poor plant stand in field which evident from the pathogenicity test. Quality of seed is the most important factor which ensures the productivity and quality of a crop. In vegetable crops like garden pea where seed is costly and its availability is limited, seed quality becomes more important for achieving optimum economic return. As quality seed is in short supply in the state the farmers are using their own seed saved from previous season crop. Hence, it is imperative to assess the quality of farmers saved seed and
suggested suitable measure for enhancing seed quality.

References


Javid A and Anjum T. 2006. Fungi associated, with seeds of some economically important crops in Pakistan. PJST, 1:68-78.


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**How to cite this article:**