

Original Research Article

<https://doi.org/10.20546/ijcmas.2020.902.156>

Partial Purification and Characterization of Proteases from the Visceral Waste of Indian Major Carp, *Labeo rohita* (Hamilton, 1822)

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ABSTRACT

Acidic and alkaline proteases from visceral waste of *Labeo rohita* (Hamilton, 1822) were isolated, partially purified by ammonium sulphate precipitation followed by dialysis, their kinetics and characteristics studied. The purification fold increased from 1.24 to 2.49 and 1.19 to 1.55 in acidic and alkaline protease respectively along the purification steps. The molecular weight was found in the range of 15-35 kDa and 25-63 kDa respectively in acidic and alkaline proteases. The pH and temperature optima for acidic and alkaline proteases were 3 and 10, at 40°C and 60°C respectively. The Protease activity was decreased by 40% and 60%, when incubated at 90°C for 30 min. Both the proteases showed a decreased activity of more than 50% after incubation with NaCl concentration of 0.5%. Degree of hydrolysis (DH) of the proteases on muscle protein increased with increase of enzyme concentrations. Both soybean trypsin inhibitor and EDTA exhibited high percentage of inhibition, when proteases were incubated with 50 mM of both the inhibitors. The study showed that proteases from Rohu visceral waste of could find use in applications where maximum activity at moderate temperature and low NaCl concentration is desired.

Keywords

Labeo rohita,
Acidic proteases,
Alkaline proteases,
Enzyme purification
fold, Enzyme
kinetics

Article Info

Accepted:
08 January 2020
Available Online:
10 February 2020

Introduction

A huge quantity of visceral wastes is generated in the retail fish markets due to pre-processing. Such biological wastes, if not utilized otherwise, would pose a problem of their disposal and subsequent environmental pollution. Fish processing wastes is about 30% of the of the whole fish and comprised of head, scales, skins and viscera (Klomklao *et*

al., 2006) and is considered as an excellent source of protein and bio-active peptides (Arnesen and Gildberg, 2007). Although there is scope to recover proteins and enzymes from the visceral wastes of fish, but huge quantity of such waste is discarded without any such attempt (Bhaskar and Mahendrakar, 2007). According to Bezerra *et al.*, (2005), the fish visceral waste usually accounts for 5% of the total mass and includes stomach, pyloric

caeca, intestines, liver, pancreas and so on and other organs like spleen and gonads. The digestive enzymes from the fish visceral waste are highly active over a wide range of pH and temperature conditions, and thereby represent an important valued by-product of fishing industry (Castillo-Yanez *et al.*, 2004). Amongst the hydrolytic enzymes, proteases represent an important class of industrial enzymes; have been employed in different applications, mostly in food, detergent, textile, leather and pharmaceuticals as well as in waste management and bioremediation process (Anwar and Saleemuddin, 1998; Gupta *et al.*, 2002). However, proteases require their purification and characterization before any application. Proteases contribute about 60% of the world's total enzyme production and used worldwide (Gupta *et al.*, 2002). Presently, most of the proteolytic enzymes are extracted from bacteria, and relatively few attempts have been made on the application of fish proteases as industrial processing aids. Usually, the fishery by-products are typically used as feeds and fertilizers. Recently, interest has grown to search high-value functional biomolecules from the fishery wastes, notably enzymes. Nevertheless, several researchers investigated proteases from the visceral wastes from marine fish (Klomklao *et al.*, 2009; Nasri *et al.*, 2011). But characterization of fish proteases especially from the visceral wastes of freshwater fish is seldom reported.

Based on the above rationale, the present study was carried out to characterize partially purified acidic and alkaline proteases from the fish visceral wastes for determining their application in food processing operations as well as to reduce waste disposal problems. Rohu (*Labeo rohita*), predominantly a column feeder and feeds mainly on filamentous algae, decomposed vegetation and mud was selected for study as it is most commonly consumed freshwater fish in India amongst the carps.

Materials and Methods

Rohu viscera

Viscera of Rohu was collected from the local markets at Agartala city, packed in polyethylene bags and transported with ice. In the laboratory, viscera was washed with chilled water in order to remove the adherent blood, slimes and dirt, kept in plastic bags and stored at -20°C until used for enzyme extraction.

Preparation of crude acidic and alkaline protease

Method suggested by Vannabun *et al.*, (2014) was followed for preparation of crude acidic and alkaline proteases. Initially the visceral mass was thawed and homogenization was done for 2 min with different extraction buffers, such as citrate buffer (10mM Citrate/HCl pH 3.0) for acid protease and tris buffer (10mM Tris-HCl pH 8.0, 10mM CaCl₂) for alkaline protease, in the ratio of 1:5(w/v). The homogenate was centrifuged at 10,000 x g for 10min at 4°C. After homogenization, the pellet was discarded to collect the supernatant which was used as 'crude enzyme extract'.

Enzyme purification

Crude enzyme extract was subjected to two-step (NH₄)₂SO₄ precipitation. As per preliminary assay, (NH₄)₂SO₄ concentration of 40-60% gave the highest purification fold and specific activity. The crude enzyme was precipitated with 40-60% saturation of ammonium sulphate and then allowed to settle for 24h at 4°C. The supernatant was discarded and the precipitate was dissolved in 0.02 M acetate buffer, pH 3.0 and 0.02 M Tris-HCl buffer, pH 8.0 for acidic and alkaline proteases respectively, by centrifugation at 10,000xg for 30 min at 4°C.

The enzyme thus obtained was dialyzed against the same buffer for 24 h at 4°C with intermittent change of buffer after 12 h. After dialysis, the crude enzyme was referred as 'partially purified proteases'.

Determination of molecular weight

The molecular weight (MW) of partially purified enzyme was carried out by SDS-PAGE, following the method suggested by Laemmli (1970). A sample buffer was prepared by mixing 2.5 ml 0.5 M Tris-HCl (pH 6.8), 4 ml 10% SDS, 2 ml glycerol, 1 ml 1% b-mercaptoethanol, 0.03 ml 0.002% bromophenol blue and the final volume was made to 10 ml. Protein solutions were mixed at a 1:2 (v/v) ratio and boiled for 10 min. Samples (10 µl) were loaded on the gel made of 4% stacking and 12.5 % separating gels and fractionated for 90 min at a constant current of 400 mA. After electrophoresis, the gels were stained with 0.05g Coomassie brilliant blue R-250 in 15% methanol and 5% acetic acid, and destained with destaining solutions [solution-1 (50% methanol and 7.5% acetic acid) and solution-2 (5% methanol and 7.5% acetic acid)]. The molecular weight was estimated using protein standard (10-245kDa) (HiMedia, India).

Protein content

The protein content was estimated following Lowry's method (Lowry *et al.*, 1951) by measuring sample absorbance at 280 and 260 nm, using bovine serum albumin as standard.

Assay of protease activity

The acidic protease activity was determined as suggested by Natalia *et al.*, (2004) using 2% bovine haemoglobin solution containing 0.04M HCl (acid denatured) as substrate at pH 3.0 and 37°C, whereas, method of Rawdkuen *et al.*, (2010) was followed to

determine alkaline protease activity using casein as a substrate. The absorbance read at 280 nm and converted into µmoles of tyrosine liberated using solutions of 25-250 µg/ml concentration of tyrosine for calibration curve. Enzymatic activity was expressed as one unit equivalent to the amount of enzyme capable of hydrolyzing bovine haemoglobin to liberate 1 µmole tyrosine under standard assay conditions. Total activity and specific activity was expressed as units of enzymatic activity per ml protein (U/ml) and per mg protein (U/mg) respectively.

Total activity

The total enzymatic activity was estimated using the following equation.

$$\text{Total Activity } \left(\frac{U}{mL}\right) = \frac{\mu\text{mole of tyrosine equivalent released} \times \text{Total Volume of reaction assay (mL)}}{\text{Volume of enzyme used (mL)} \times \text{Time of assay (min)} \times \text{Volume used in Cuvette (mL)}}$$

Specific activity of enzyme

The specific activity of both the enzymes was determined using the equation as suggested by El-beltagy *et al.*, (2005).

$$\text{Specific activity } \left(\frac{U}{mg}\right) = \frac{\text{Total activity } \left(\frac{U}{mL}\right)}{\text{Protein content } \left(\frac{mg}{mL}\right)}$$

Enzyme purification fold

The level of purification was evaluated by determining the purification fold following the equation given by El-beltagy *et al.*, (2005).

$$\text{Purification fold} = \frac{\text{Specific Activity}}{\text{Specific Activity of crude extract}}$$

Optimum pH and pH stability

The optimum pH for enzymatic activity was determined following the method of Vannabun *et al.*, (2014), by assaying protease

activity at different pH conditions using 100mM buffer solutions ranging from pH 1.0 to 12.0 (Glycine- HCl buffer for pH (1.0-3.0); sodium acetate buffer for pH (4.0-6.0); Tris-HCl buffer for pH (7.0–9.0); and Glycine–NaOH buffer for (9.0-12.0), at the optimum temperature for activity previously determined.

The effect of pH on enzyme stability was determined by the method of Vannabun *et al.*, (2014). The enzyme was incubated at various pH (1.0–12.0) using different buffers of 100mM Glycine–HCl (1.0–3.0), Na-acetate (4.0–6.0), Tris–HCl (7.0–9.0) and Glycine–NaOH (10.0–12.0) for 30 min along with the blanks prepared simultaneously. The residual enzymatic activity after incubation was evaluated and compared with the condition that showed the highest value (100% activity).

Optimum temperature and temperature stability

Protease activity at different temperatures (30–90°C) was performed by using different buffers like Glycine–HCl (pH 3.0) and Tris–HCl (pH 8.0) for acidic and alkaline protease activity respectively according to the method given by Vannabun *et al.*, (2014). To determine thermal stability of proteases, enzyme extract was incubated for various time durations like 1,3,5,10,15,20,30,40,50, and 60 min at 90°C and the remaining enzymatic activity was determined. The control was not pre-incubated and considered as 100% activity.

Effect of NaCl concentration on enzyme activity

The reaction mixture was made with different conc. of NaCl (0–2.5%, w/v) and enzyme was incubated followed by determination of residual activity. The control was made without NaCl and its activity was considered as 100% activity (Vannabun *et al.*, 2014).

Effect of isolated enzymes on proteins hydrolysis

Extracted acidic and alkaline proteases were used to hydrolyze the ground fish muscle protein to determine the degree of hydrolysis of enzyme on the fish muscle. The ground muscle (2g) was incubated with enzyme at different concentrations (10-50 mL) for 30 minutes at 60°C. The reaction was stopped by adding 5 mL of 20% TCA followed by centrifugation at 3300 rpm for 10 minutes to collect the 10% TCA soluble material as the supernatant. The protein content of the supernatant was estimated by Biuret method. The degree of hydrolysis was determined by the method of Hoyle and Merritt (1994)

$$\%DH = \frac{10\% \text{ TCA soluble protein in the sample}}{\text{Total protein content of sample}} \times 100$$

Enzyme inhibitors and activators

Enzyme inhibitors such as soybean trypsin inhibitor (STPI) and ethylene diamine tetraacetic acid (EDTA) were used to determine their effects on the enzymatic activity.

Pre-incubation of the substrates with those at different conc. like 10, 20, 30, 40 and 50 mM were done for 10 min at the optimum temperature of enzyme followed by determination of enzyme activity. The results were expressed as a relative percentage of the activity without modifiers.

Statistical analysis

Analysis of variance (ANOVA) followed by Duncan's multiple range test was carried out to determine differences between means. Statistical analysis was performed using the Statistical Package for Social Sciences (SPSS for Windows version 16.0, SPSS, Inc., Chicago, IL).

Results and Discussion

Partial purification of proteases

The protein content, total activity, specific activity and purification fold for acidic and alkaline proteases of visceral waste of Rohu is presented in Table 1. The average protein content was found to be 6.31 mg/ml and 7.79 mg/ml in acidic and alkaline crude proteases respectively. After ammonium sulphate fractionation (40-60%), the protein content decreased in the crude proteases and the values reached to 3.72 mg/ml and 4.15 mg/ml in acidic and alkaline crude proteases respectively. The saturated ammonium sulphate solution selectively precipitates proteins from the crude enzyme extract by the salting-in and salting-out mechanism to form a partially purified enzyme extract (Klomkiao *et al.*, 2006). This may be due to the impurities present in the crude sample which are removed after ammonium sulphate precipitation. Dialysis, a step in the purification of proteases, exhibited a further decline of protein content in the $(\text{NH}_4)_2\text{SO}_4$ precipitated proteases. In acidic and alkaline proteases the average protein content decreased to 1.68 mg/ml and 2.96 mg/ml respectively. Such decrease of protein content after dialysis may be due to further removal of other proteins, not removed by ammonium sulphate fractionation.

Molecular weight of proteases

The electrophoretic pattern showed several clear bands indicating the presence of different proteases of varying molecular mass in case of both alkaline and acidic protease samples (Fig. 1). In case of crude and partially purified acidic and alkaline proteases 3-4 bands were observed ranging from 15-35 kDa and 25-63 kDa respectively. Several authors reported the molecular weights of visceral alkaline and acidic proteases in the

range of 17-90 kDa. Molecular weight of acidic protease from *Tilapia nilotica* after gel filtration on Sephadex G-100 was reported as 31.0 kDa (El-Beltagy *et al.*, 2004). Whereas, Lopez-Liorca (1990) and Liu *et al.*, (2008) reported the molecular weight of visceral acidic protease of fish as around 32 kDa and 28.5 kDa respectively. Molecular weight of fish visceral alkaline proteases have been reported as 23.5 kDa (Bezerra *et al.*, 2005), 23-28 kDa (Balti *et al.*, 2009), 23 kDa (El-Beltagy *et al.*, 2004), 24-30 kDa (Sekizaki *et al.*, 2000). The present study revealed that the molecular weights of alkaline proteases are higher compared to acidic proteases. Presence of several bands in the electrophoretic separation of digestive proteases was explained as due to constituent enzymes like trypsin, chymotrypsin, collagenase, gastrin, pepsin, elastase, carboxypeptidase and carboxyl esterase (Barkia *et al.*, 2010), and also due mostly to the different molecular weights of individual enzyme. The present study justified the observations reported by earlier researchers in the context of molecular weight distribution of digestive proteases (Younes *et al.*, 2014; Sila *et al.*, 2012).

Assay of proteolytic activity

The average total activity of crude acidic and alkaline Rohu viscera waste was determined to be 18.33 U/ml and 34.11 U/ml respectively. Total proteolytic activity reduced after $(\text{NH}_4)_2\text{SO}_4$ fractionation (ASF) and further reduction took place after dialysis. In case of alkaline proteases the recovery percentage was found to be 63.67 and 58.95 after ASF and dialysis respectively (Table 1). Similar was observed in case of acidic proteases, wherein, recovery of total activity after ASF and dialysis was found to be 72.76 and 66.28 respectively (Table 1). Purification might have removed other catheptic enzymes which were probably present in the visceral waste, and resulted decrease of the total

activity. Such decrease of protease activity after purification was also reported (Subash *et al.*, 2011; Kim *et al.*, 2012).

The average specific activity after dialysis was found to be 6.79 and 7.23 in case of alkaline and acidic proteases respectively. Such increase of specific activity along the purification steps may be explained as the removal of interfering proteins during $(\text{NH}_4)_2\text{SO}_4$ fractionation and further during dialysis, resulting enhanced activity. Increase of specific activity with the progress of the purification was also reported by Liu *et al.*, (2008), Bezerra *et al.*, (2005) and El-Beltagy *et al.*, (2004).

This study also revealed that the specific activity of post dialysis acidic proteases was more than the alkaline proteases, although, the later showed more total activity and also pre-dialysis specific activity compared to the acidic one. Since the Rohu being lacking of true stomach, might be the reason for low amount of acidic proteases secretion in the gut content of viscera, as stomach constitutes an important source of digestive proteolytic enzymes (Simpson, 2000).

The specific activity of the enzyme determines the purification fold. In the two-step purification system, the purification fold experienced an increase from step two to step three in case of both acidic and alkaline proteases. In case of alkaline proteases the result showed that increase in purification fold was from 1.19 to 1.55, whereas, it was 1.24 to 2.49 in case of acidic proteases. Increase of purification fold following dialysis has also been reported by El-Beltagy *et al.*, (2004) and Liu *et al.*, (2008). Purity of trypsin-like enzyme from anchovy digestive tract was increased by 2.7-fold following ammonium sulphate precipitation (20-70%) (Martinez *et al.*, 1988).

Optimum pH and pH stability

Partially purified acid and alkaline proteases was found to be active over a range of pH 1.0–12.0 using casein and acid-denatured bovine haemoglobin as substrates for alkaline and acidic proteases respectively. The acidic protease exhibited high activity in the pH range from 2-4 with an estimated maximum at 3.0 and then decreased significantly ($p < 0.05$) with increasing pH (Fig. 2a). Relative activity of about more than 50% was lost over pH 4. In other study, the optimum pH for hydrolysis of acid denatured bovine haemoglobin by partially purified acidic protease from *Tilapia nilotica* was found to be 2.5 (El-Beltagy *et al.*, 2004). Our results corroborate well with the observation of Bougatef *et al.*, (2009), who reported pH optima for acidic proteases in the range of 2-4. The alkaline protease exhibited maximum activity at pH 10, and then decreased significantly at higher pH levels (Fig. 2c). Optimum pH for maximum activity of alkaline protease was reported in the range of 8-10 (Nasri *et al.*, 2011). Determination of pH optima of an enzyme is very essential as this is considered to be an important indicator for its potential application in different purposes.

Both the acidic and alkaline proteases were highly stable over a wide pH range, maintaining more than 90% of its original activity between pH 1.0-5.0 and pH 8.0-12.0 in respect of acid and alkaline proteases respectively after 30 minutes incubation at 37°C (Fig. 2b, d). The pH stability of proteases depends on the differences in molecular properties, which includes bonding and stability of the structure; conformation of enzyme in different anatomical locations amongst various species (Klomklao *et al.*, 2007). Similar findings regarding pH stability of acidic protease from fish has also been reported by Castillo-Yanez *et al.*, (2004) for Monterey sardine. The pH stability of alkaline

proteases in the range of 6-12, has been reported by several authors (Sila *et al.*, 2012; Younes *et al.*, 2014; Klomklao *et al.*, 2011). Acidic protease activity showed a decrease of about 15-20% at pH over 6.0 whereas; a similar decrease was shown by alkaline protease at pH below 7.0.

Optimum temperature and thermo stability

In this study, the optimum activity of acidic protease was found at 40°C (Fig. 3a) which is similar to the earlier reports from other fish, viz., pepsins from *Sardinelle* by Ben Kahled *et al.*, (2008) and smooth hound by Bougateg *et al.*, (2009). The optimal temperature of alkaline protease activity was found as 60°C (Fig. 3c) and the similar result was reported by Klomklao *et al.*, (2011) and Cao *et al.*, (2000) for trypsin from the pyloric caeca of Chinook salmon (*Oncorhynchus tshawytscha*) and Japanese seabass (*Lateolabrax japonicas*) respectively.

The result showed that the enzyme activity of proteases increased to a certain point followed by a decrease with increase in temperature forming a bell shaped curve. At temperature above optimum the native conformation of protein is changed due to breakdown of weak intramolecular bonds responsible of stabilization of three dimensional structure of the enzyme active site (Klomklao *et al.*, 2011). As opined by Klomklao *et al.*, (2006), environmental and genetic factors among the different species might be responsible for the native conformations of enzymes.

The study also revealed that acidic and alkaline proteases' activity decreased by 40 and 60% respectively when the incubation condition was 90°C for 30 min (Fig. 3b & d). This may be explained as the inactivation of enzymatic activity following stretching out of

the enzyme's native conformation during thermal treatment (Klomklao *et al.*, 2011). Vannabun *et al.*, (2014) also reported similar findings while characterizing visceral acidic and alkaline proteases of farmed giant cat fish. As proposed by Sabtecha *et al.*, (2014), stability of a fish enzyme in different temperature is influenced by their habitat, environment and genetic characters.

Effect of NaCl on enzyme activity

Fig. 4 illustrates the effect of NaCl on enzyme activity of proteases. Relative enzyme activity showed a decrease of more than 50% for both acidic and alkaline proteases at NaCl concentration of 0.5%. Further increase of NaCl concentration slightly decreased protease activity.

A 10% decrease in the relative activity of acidic protease from *Sardinelle* at 20% NaCl concentration was reported by Ben Kahled *et al.*, (2008). This indicates that the activity of proteases of freshwater fish differs from the marine fish, and this is mainly due to salt concentration of the habitat.

Klomklao *et al.*, (2011) investigated trypsin activity from hybrid catfish and found that the enzyme activity significantly decreased gradually with the increasing concentration of NaCl. This loss of enzyme activity might be due to the denaturation of enzyme (Ben Kahled *et al.*, 2008) resulting the "salting out" effect.

The ionic strength is increased with the increasing salt concentration. In the high ionic strength, the enzyme activity is reduced due to superior hydrophobic-hydrophobic interaction between proteins of the enzyme and also enhanced affinity of ionic salts for water thereby resulting precipitation of enzyme (Klomklao *et al.*, 2009).

Table.1 Purification of acidic & alkaline proteases from visceral waste of rohu

	Purification steps	Protein Content (mg/mL)	Total Activity (U/mL)	Specific Activity (U/mg)	Recovery (%)	Purification Fold
Acidic Protease	Crude	6.31±0.08	18.33±0.06	2.90±0.05	100	1
	Ammonium Sulfate Fractionation (40-60%)	3.72±0.04	13.37±0.02	3.59±0.01	72.76	1.24
	Dialysis	1.68±0.05	12.15±0.28	7.23±0.15	66.28	2.49
Alkaline Protease	Crude	7.79±0.06	34.11±0.11	4.38±0.01	100	1
	Ammonium Sulfate Fractionation (40-60%)	4.15±0.03	21.72±0.76	5.23±0.04	63.67	1.19
	Dialysis	2.96±0.03	20.11±0.61	6.79±0.07	58.95	1.55

*Values given in the table are means ± SD, n=3.

Figure.1 Electrophoretic pattern of Rohu viscera waste (C=standard protein marker, A=crude enzyme extract, and B=purified enzyme extract)

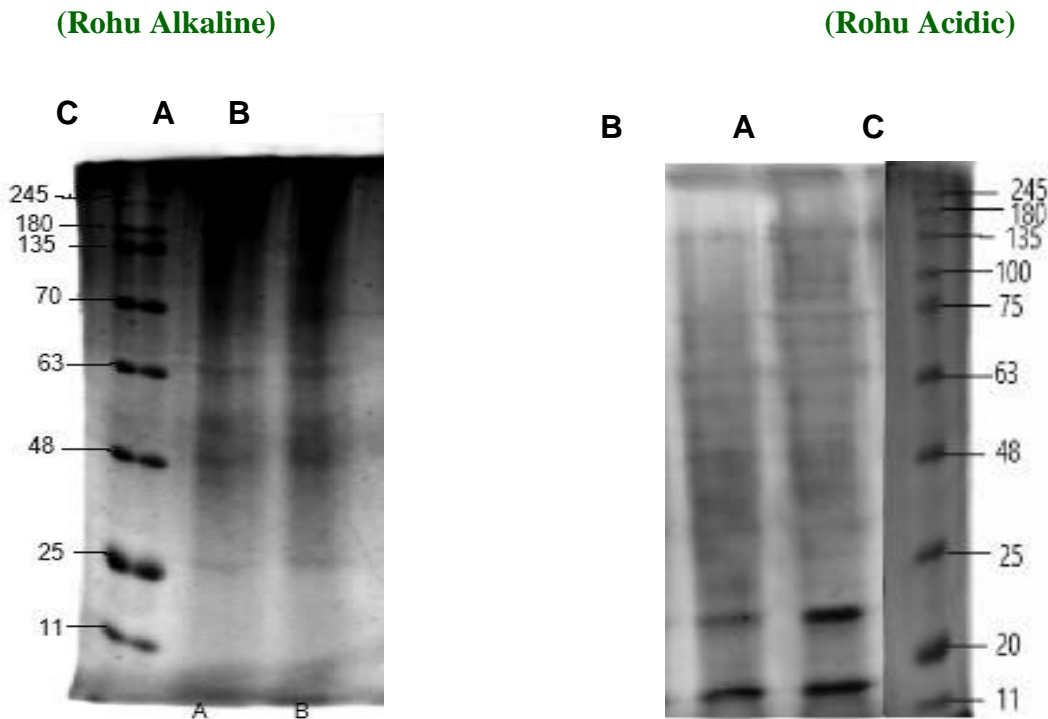
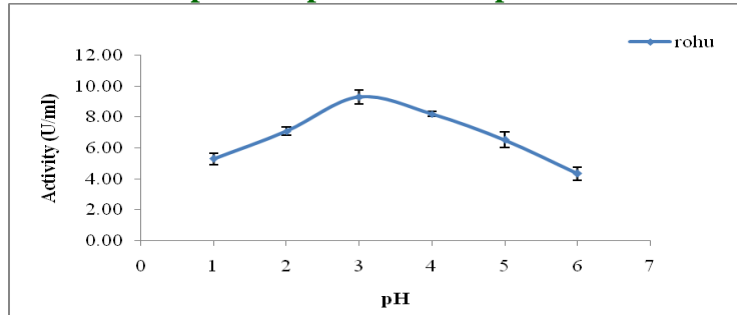
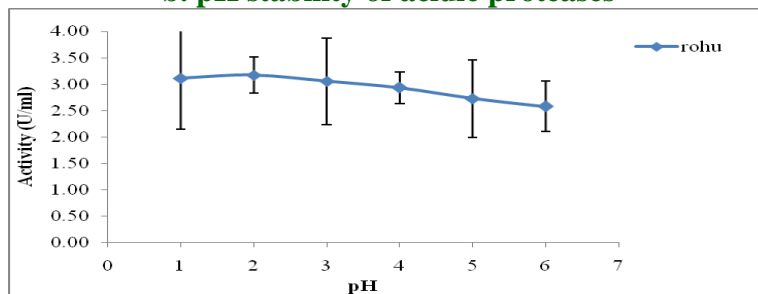


Figure.2 Optimum pH and pH stability for maximum activity of proteases from Rohu viscera waste (a-optimum pH for acidic proteases, b-pH stability of acidic proteases, c-optimum pH for alkaline proteases, d-pH stability of alkaline proteases)

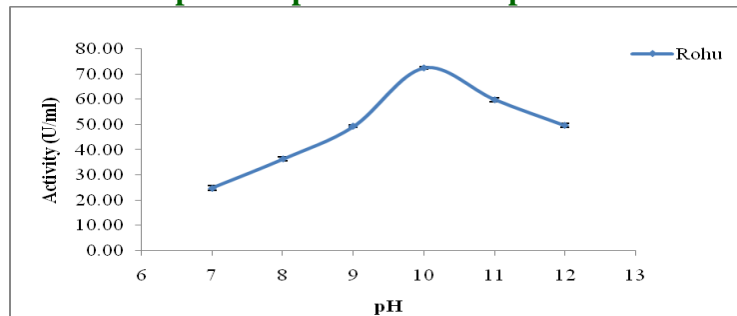
a. Optimum pH for acidic proteases



b. pH stability of acidic proteases



c. Optimum pH for alkaline proteases



d. pH stability of alkaline proteases

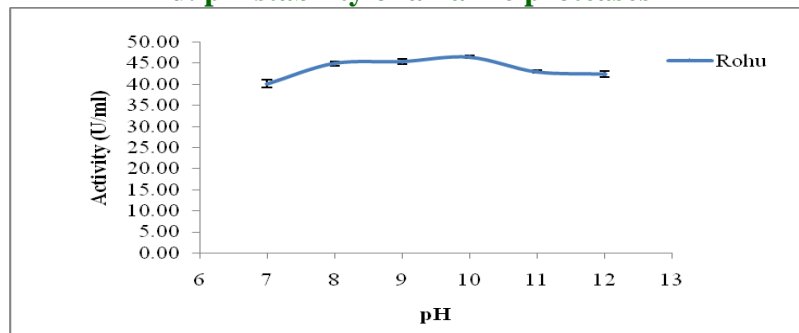
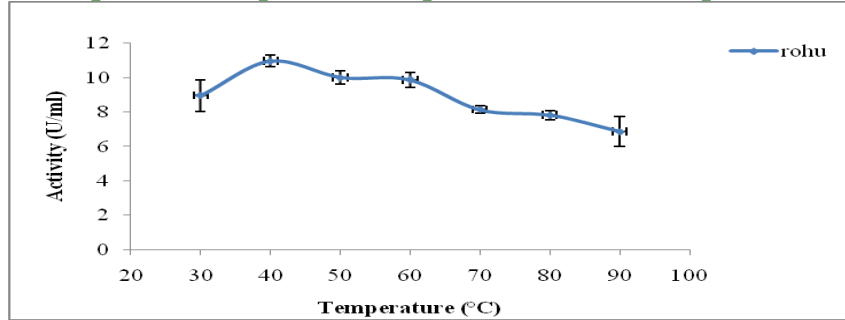
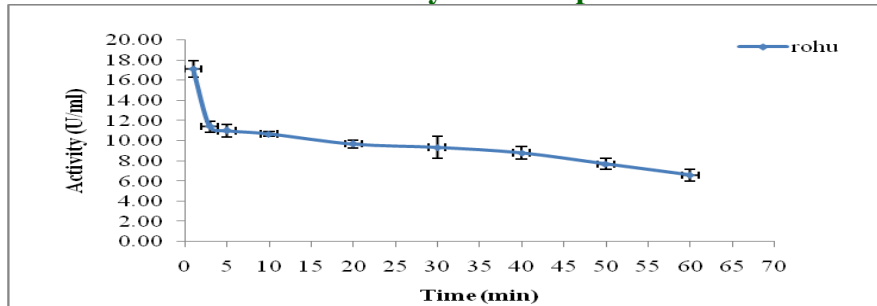


Figure.3 Optimum temperature and thermostability for maximum activity of proteases from Rohu viscera waste (a-optimum temp. requirement for acidic proteases, b-thermostability of acidic proteases, c-optimum temp. requirement for alkaline proteases, and d-thermostability of alkaline proteases)

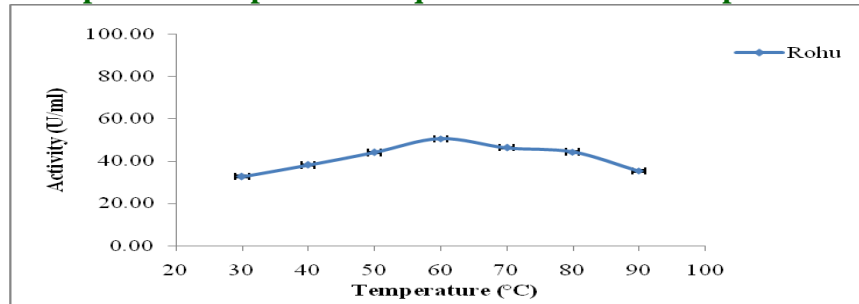
a. Optimum temperature requirement for acidic proteases



b. Thermostability of acidic proteases



c. Optimum temperature requirement for alkaline proteases



d. Thermostability of alkaline proteases

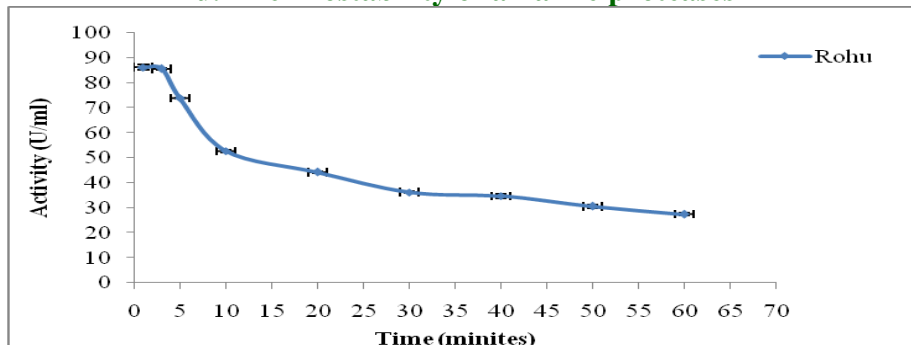
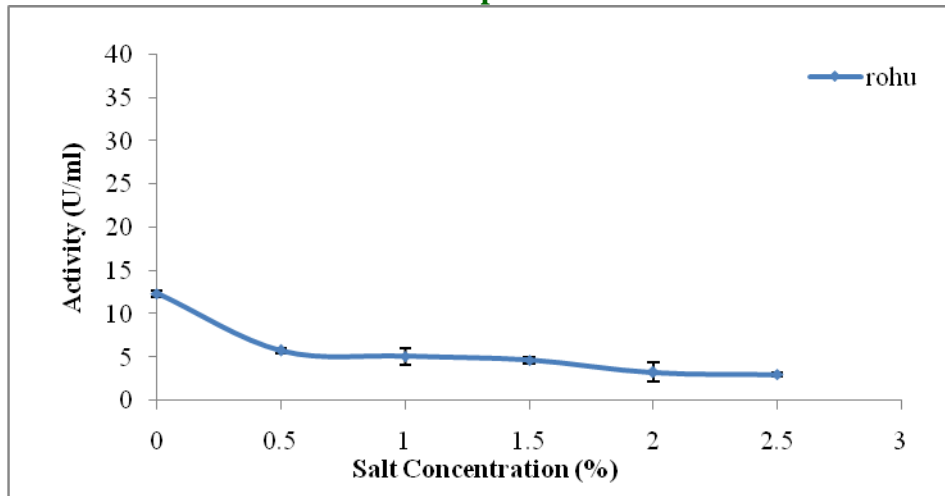


Figure.4 Effect of NaCl concentration on the activity of proteases from Rohu viscera waste (a= acidic proteases, and b= alkaline proteases)

a. Acidic proteases



b. Alkaline proteases

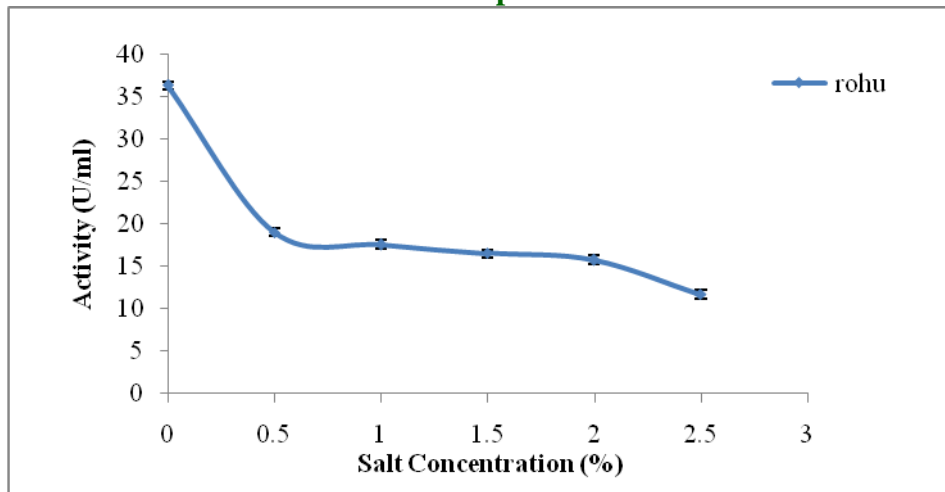
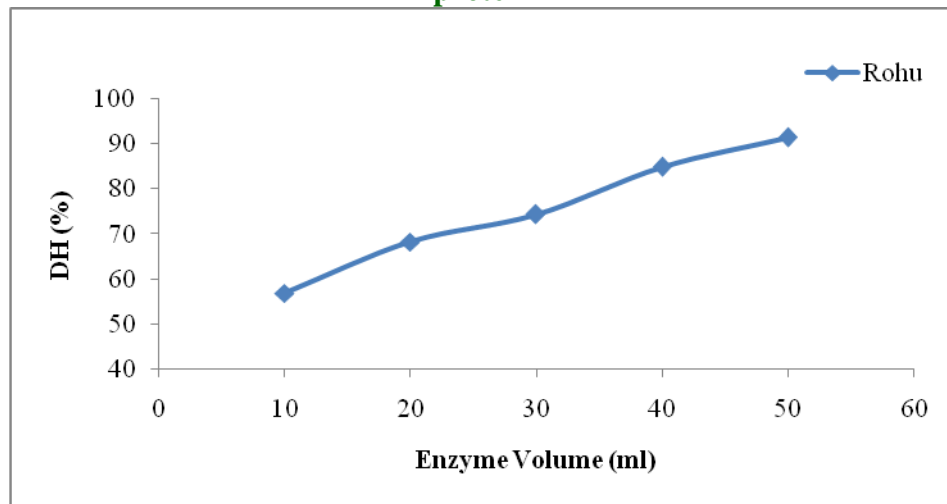


Figure.5 Effect of crude proteases from Rohu viscera waste on hydrolysis of muscle protein

a. Effect of crude proteases (Acidic) from Rohu viscera waste on hydrolysis of muscle protein



b. Effect of crude proteases (Alkaline) from Rohu viscera waste on hydrolysis of muscle protein

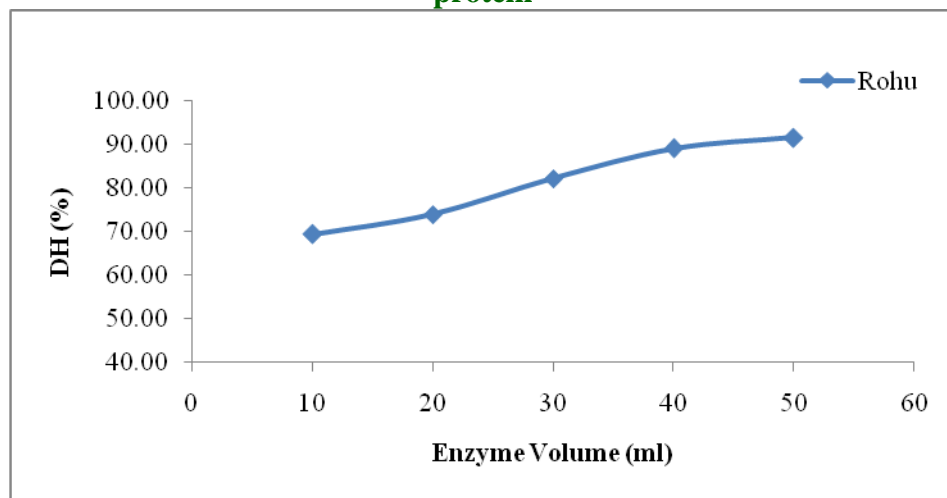
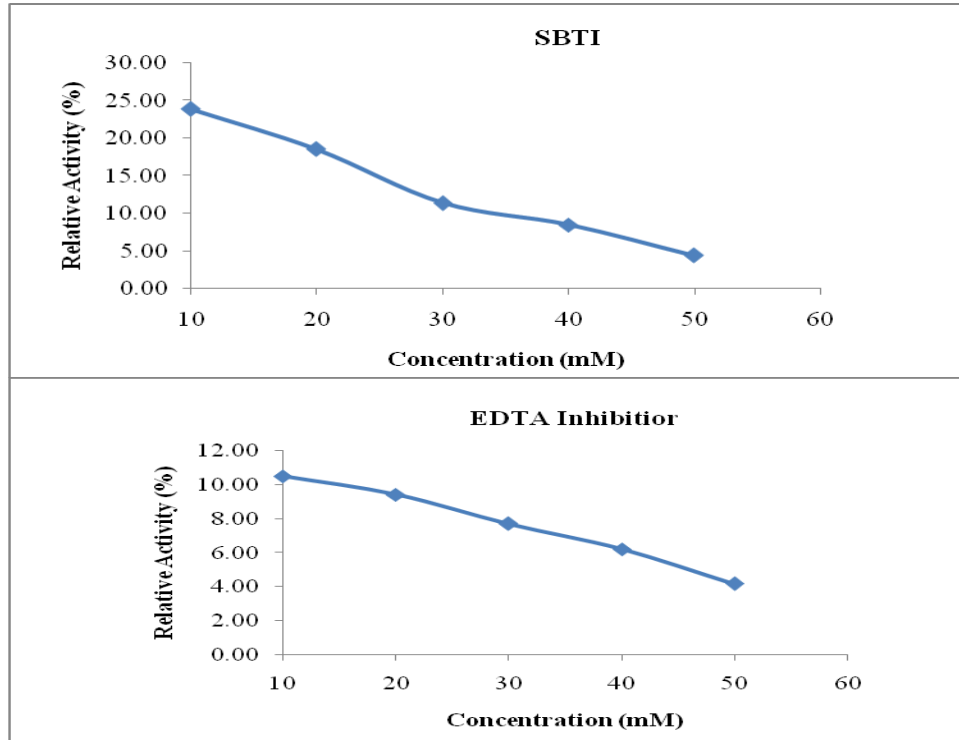


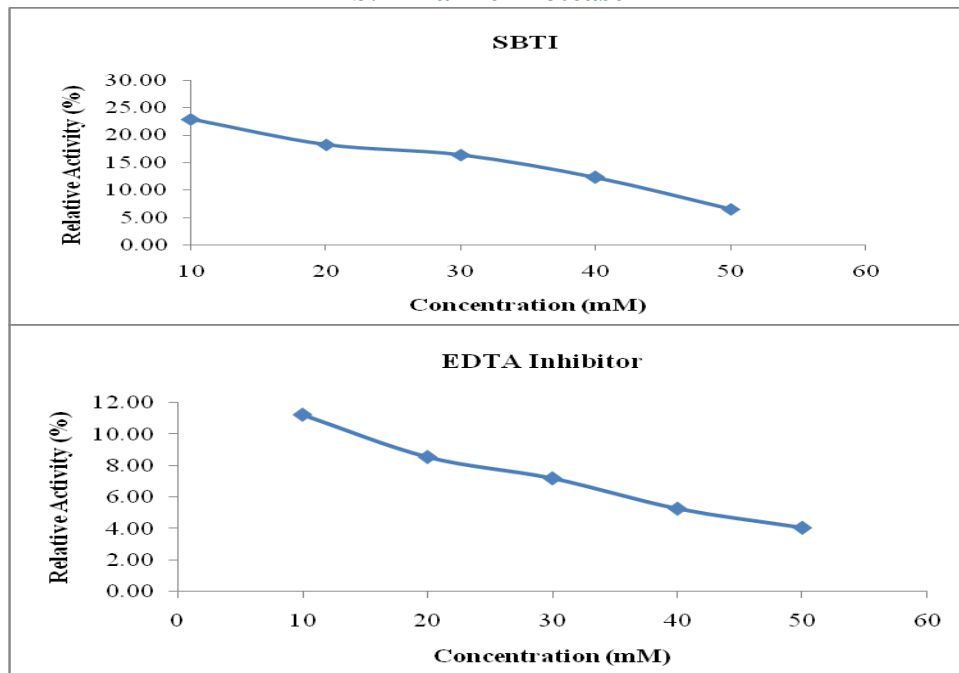
Figure.6 Influence of some inhibitors on the activity of proteases from Rohu viscera waste (a= acidic proteases, and b= alkaline proteases)

Influence of inhibitors on the activity of proteases from Rohu visceral waste

a. Acidic Protease



b. Alkaline Protease



Effect of isolated enzymes on proteins hydrolysis

As degree of hydrolysis (DH) is the indicative of the extent of peptide bonds broken down (Adler-Nissen, 1979), its determination is crucial since several characteristics of protein hydrolysates is DH dependent. Using ground muscle meat of fish as substrate, hydrolysis was conducted at temperature 37°C and optimum pH for both the enzymes. The degree of hydrolysis (DH) as a function of the enzyme concentration is given in Fig. 5. The result of this study signifies that higher amount of proteases in enzyme fraction cleaved more peptide bonds and similar observation was also reported by Klompong *et al.*, (2008).

Effect of inhibitors on the enzyme activity

A high inhibition percentages of total acidic protease activity were obtained when the acidic protease was incubated with 50 mM of both soybean trypsin inhibitor or ethylenediaminetetraacetic acid (90.9% and 68.8%, respectively), while they were 23.9% and 10.5% respectively, when 10 mM concentration was used (Fig. 6). Almost similar percent inhibition was obtained in case of alkaline protease. Our result is in agreement with the findings of Diaz-Lopez *et al.*, (1998) and El-Beltagy *et al.*, (2004, 2005) for the acidic protease. In a study with tilapia digestive proteases, a high inhibition of approx. 40% was reported using very low concentration of SBTI (Moyano Lopez *et al.*, 1999). Inhibition of Rohu alkaline proteases at 250 µM concentration of SBTI was reported to be 78.1% (Kumar *et al.*, 2007).

In conclusion, this study has revealed that considerable amounts of acidic and alkaline proteases are present in the visceral waste of Rohu fish, and those in purified form have potential for application as different food

processing aids, and on the other hand, would contribute to solve bio-waste disposal problem to a great extent. Both the proteases exhibited substantial activity in both acidic and alkaline conditions. As a requirement for their application purpose, the maximum activity of acidic and alkaline protease was found to be at 40°C and 60°C respectively. Nevertheless, the stability of these enzymes at elevated temperature and NaCl concentration was not found to be satisfactory. Based on the present study, the enzymes from Rohu visceral waste could find use in applications where maximum activity at moderate temperature and low NaCl concentration is desired.

References

- Adler-Nissen, J. 1979. Determination of the degree of hydrolysis of food protein hydrolysates by trinitrobenzene sulfonic acid. *Journal of agricultural and food chemistry*. 27(6): 1256-1262.
- Anwar, A., and Saleemuddin, M. 1998. Alkaline proteases: a review. *Bioresource technology*. 64(3): 175-183.
- Arnesen, J. A., and Gildberg, A. 2007. Extraction and characterisation of gelatine from Atlantic salmon (*Salmo salar*) skin. *Bioresource Technology*. 98(1): 53-57.
- Balti, R., Barkia, A., Bougatef, A., Ktari, N., and Nasri, M. 2009. A heat-stable trypsin from the hepatopancreas of the cuttlefish (*Sepia officinalis*): Purification and characterisation. *Food Chemistry*. 113(1): 146-154.
- Barkia, A., Bougatef, A., Nasri, R., Fetoui, E., Balti, R., and Nasri, M. (2010). Trypsin from the viscera of Bogue (*Boops boops*): isolation and characterisation. *Fish physiology and biochemistry*. 36(4): 893-902.
- Ben Khaled, H., Bougatef, A., Balti, R., Triki- Ellouz, Y., Souissi, N., and Nasri, M. 2008. Isolation and characterisation of trypsin from sardinelle (*Sardinella aurita*) viscera. *Journal of the Science of Food and*

- Agriculture. 88(15): 2654-2662.
- Bezerra, R. S., Lins, E. J., Alencar, R. B., Paiva, P. M., Chaves, M. E., Coelho, L. C., and Carvalho Jr, L. B. 2005. Alkaline proteinase from intestine of Nile tilapia (*Oreochromis niloticus*). *Process Biochemistry*. 40(5): 1829-1834.
- Bhaskar, N., & Mahendrakar, N. S. 2007. Chemical and microbiological changes in acid ensiled visceral waste of Indian major carp *Catla catla* (Hamilton) with emphasis on proteases. *Indian Journal of Fisheries*. 54(2): 217-225.
- Bougatef, A., Hajji, M., Balti, R., Lassoued, I., Triki-Ellouz, Y., and Nasri, M. 2009. Antioxidant and free radical-scavenging activities of smooth hound (*Mustelus mustelus*) muscle protein hydrolysates obtained by gastrointestinal proteases. *Food chemistry*. 114(4): 1198-1205.
- Cao, M. J., Osatomi, K., Suzuki, M., Hara, K., Tachibana, K., and Ishihara, T. 2000. Purification and characterization of two anionic trypsins from the hepatopancreas of carp. *Fisheries Science*, 66(6): 1172-1179.
- Castillo-Yanez, F. J., Pacheco-Aguilar, R., Garcia-Carreño, F. L., and de los Angeles Navarrete-Del, M. 2004. Characterization of acidic proteolytic enzymes from Monterey sardine (*Sardinops sagax caerulea*) viscera. *Food Chemistry*. 85(3): 343-350.
- Chakrabarti, R., and Sharma, J. G. 2005. Digestive physiology of fish larvae during ontogenic development: a brief overview. *The Indian Journal of Animal Sciences*. 75(11): 1337-1347.
- Diaz-López, M., Moyano-López, F. J., Alarcón-López, F. J., Garcia-Carreno, F. L., and del Toro, M. A. N. 1998. Characterization of fish acid proteases by substrate-gel electrophoresis. *Comparative Biochemistry and Physiology Part B: Biochemistry and Molecular Biology*. 121(4): 369-377.
- El-Beltagy, A. E., El-Adawy, T. A., Rahma, E. H., and El-Bedawey, A. A. 2004. Purification and characterization of an acidic protease from the viscera of boliti fish (*Tilapia nilotica*). *Food Chemistry*. 86(1): 33-39.
- El- Beltagy, A. E., El- Adawy, T. A., Rahma, E. H., and El- Bedawey, A. A. 2005. Purification and characterization of an alkaline protease from the viscera of boliti fish (*Tilapia nilotica*). *Journal of food biochemistry*. 29(5): 445-458.
- Grabner, M. 1985. An in vitro method for measuring protein digestibility of fish feed components. *Aquaculture*. 48(2): 97-110.
- Gupta, R., Beg, Q., and Lorenz, P. 2002. Bacterial alkaline proteases: molecular approaches and industrial applications. *Applied microbiology and biotechnology*. 59(1): 15-32.
- Hoyle, N. T., and Merritt, J. H. 1994. Quality of fish protein hydrolysates from herring (*Clupea harengus*). *Journal of food Science*. 59(1): 76-79.
- Jhingran, V. G. 1991. *Fish and Fisheries of India*. (3rd Ed.). Hindustan Publishing Corporation, New Delhi.
- Kim, S. K., Ngo, D. H., and Vo, T. S. 2012. Marine fish-derived bioactive peptides as potential antihypertensive agents. In Kim, S. K. (Eds.), *Advances in food and nutrition research*. Academic Press, United States. pp. 249-260.
- Klomklao, S., Benjakul, S., Kishimura, H., and Chaijan, M. 2011. 24 kDa Trypsin: A predominant protease purified from the viscera of hybrid catfish (*Clarias macrocephalus* × *Clarias gariepinus*). *Food chemistry*. 129(3): 739-746.
- Klomklao, S., Benjakul, S., Visessanguan, W., Kishimura, H., and Simpson, B. K. 2006. Purification and characterization of trypsin from the spleen of tongol tuna (*Thunnus tonggol*). *Journal of agricultural and food chemistry*. 54(15): 5617-5622.
- Klomklao, S., Benjakul, S., Visessanguan, W., Kishimura, H., and Simpson, B. K. 2007. Trypsin from the pyloric caeca of bluefish (*Pomatomus saltatrix*). *Comparative Biochemistry and Physiology Part B: Biochemistry and Molecular Biology*. 148(4): 382-389.

- Klomklao, S., Benjakul, S., Visessanguan, W., Kishimura, H., Simpson, B. K., and Saeki, H. 2006. Trypsins from yellowfin tuna (*Thunnus albacores*) spleen: purification and characterization. *Comparative Biochemistry and Physiology Part B: Biochemistry and Molecular Biology*. 144(1): 47-56.
- Klomklao, S., Kishimura, H., Nonami, Y., and Benjakul, S. 2009. Biochemical properties of two isoforms of trypsin purified from the intestine of skipjack tuna (*Katsuwonus pelamis*). *Food Chemistry*. 115(1): 155-162.
- Klompong, V., Benjakul, S., Kantachote, D., Hayes, K. D., and Shahidi, F. 2008. Comparative study on antioxidative activity of yellow stripe trevally protein hydrolysate produced from Alcalase and Flavourzyme. *International journal of food science & technology*. 43(6): 1019-1026.
- Kumar, S., Garcia- Carreño, F. L., Chakrabarti, R., Toro, M. A. N., and Córdova- Murueta, J. H. 2007. Digestive proteases of three carps *Catla catla*, *Labeo rohita* and *Hypophthalmichthys molitrix*: partial characterization and protein hydrolysis efficiency. *Aquaculture Nutrition*. 13(5): 381-388.
- Laemmli, U. K. 1970. Cleavage of structural proteins during the assembly of the head of bacteriophage T4. *Nature*. 227(5259): 680.
- Liu, Z. Y., Wang, Z., Xu, S. Y., and Xu, L. N. 2007. Two trypsin isoforms from the intestine of the grass carp (*Ctenopharyngodon idellus*). *Journal of Comparative Physiology B*. 177(6): 655-666.
- López, F. M., Díaz, I. M., López, M. D., and López, F. A. 1999. Inhibition of digestive proteases by vegetable meals in three fish species; seabream (*Sparus aurata*), tilapia (*Oreochromis niloticus*) and African sole (*Solea senegalensis*). *Comparative Biochemistry and Physiology Part B: Biochemistry and Molecular Biology*. 122(3): 327-332.
- Lopez-Llorca, L. V. 1990. Purification and properties of extracellular proteases produced by the nematophagous fungus *Verticillium suchlasporium*. *Canadian Journal of Microbiology*. 36(8): 530-537.
- Lowry, O. H., Rosebrough, N. J., Farr, A. L., and Randall, R. J. 1951. Protein measurement with the Folin phenol reagent. *Journal of biological chemistry*. 193(1): 265-275.
- Maitra, S., and Ray, A. K. 2003. Inhibition of digestive enzymes in rohu, *Labeo rohita* (Hamilton), fingerlings by tannin: an in vitro study. *Aquaculture Research*. 34(1): 93-95.
- Martínez, A., Olsen, R. L., and Serra, J. L. 1988. Purification and characterization of two trypsin-like enzymes from the digestive tract of anchovy *Engraulis encrasicolus*. *Comparative biochemistry and physiology. B, Comparative biochemistry*. 91(4): 677-684.
- Moyano, F. J., Diaz, M., Alarcon, F. J., and Sarasquete, M. C. 1996. Characterization of digestive enzyme activity during larval development of gilthead seabream (*Sparus aurata*). *Fish Physiology and Biochemistry*. 15(2): 121-130.
- Nasri, R., Younes, I., Lassoued, I., Ghorbel, S., Ghorbel-Bellaaj, O., and Nasri, M. 2011. Digestive alkaline proteases from *Zosterisessor ophiocephalus*, *Raja clavata*, and *Scorpaena scrofa*: characteristics and application in chitin extraction. *Journal of amino acids*. 2011, 1-9.
- Natalia, Y., Hashim, R., Ali, A., and Chong, A. 2004. Characterization of digestive enzymes in a carnivorous ornamental fish, the Asian bony tongue *Scleropages formosus* (Osteoglossidae). *Aquaculture*. 233(1-4): 305-320.
- Raksakulthai, R., and Haard, N. F. 1999. Purification and characterization of aminopeptidase fractions from squid (*Illex illecebrosus*) hepatopancreas. *Journal of food biochemistry*. 23(2): 123-144.
- Rawdkuen, S., Chaiwut, P., Pintathong, P., and Benjakul, S. 2010. Three-phase partitioning of protease from *Calotropis procera* latex. *Biochemical Engineering Journal*. 50(3): 145-149.

- Sabtecha, B., Jayapriya, J., and Tamilselvi, A. 2014. Extraction and characterization of proteolytic enzymes from fish visceral waste: potential applications as destainer and dehairing agent. *International Journal of ChemTech Research*. 6(10): 4504-4510.
- Sekizaki, H., Itoh, K., Murakami, M., Toyota, E., and Tanizawa, K. 2000. Anionic trypsin from chum salmon: activity with p-amidinophenyl ester and comparison with bovine and *Streptomyces griseus* trypsins. *Comparative Biochemistry and Physiology Part B: Biochemistry and Molecular Biology*. 127(3): 337-346.
- Sila, A., Nasri, R., Jridi, M., Balti, R., Nasri, M., and Bougatef, A. 2012. Characterisation of trypsin purified from the viscera of Tunisian barbel (*Barbus callensis*) and its application for recovery of carotenoproteins from shrimp wastes. *Food chemistry*. 132(3): 1287-1295.
- Simpson, B.K. 2000. Digestive proteinases from marine animals. In: *Seafood Enzymes: Utilization and Influence on Postharvest Seafood Quality* (Eds.) B.K. Simpson and N.F. Haard. Marcel Dekker, New York. Pp. 531–540.
- Subash, A., Shobana, A., Shanmugakani, G., Swetha, V. P., and Josephine, J. L. 2011. Utilization of Visceral Waste of Ribbon Fish (*Trachipterus Trachipterus*) for the Extraction, Purification and Characterization of Protease. *Journal of Cell and Tissue Research*. 11(1): 2589.
- Temiz, H., Ustun, N. S., Turhan, S., and Aykut, U. 2013. Partial purification and characterization of alkaline proteases from the Black Sea anchovy (*Engraulis encrasicolus*) digestive tract. *African Journal of Biotechnology*. 12(1): 56-63.
- Vannabun, A., Ketnawa, S., Phongthai, S., Benjakul, S., and Rawdkuen, S. 2014. Characterization of acid and alkaline proteases from viscera of farmed giant catfish. *Food Bioscience*. 6, 9-16.
- Villalba-Villalba, A. G., Ramírez-Suárez, J. C., Valenzuela-Soto, E. M., Sanchez, G. G., Ruiz, G. C., and Pacheco-Aguilar, R. 2013. Trypsin from viscera of vermiculated sailfin catfish, *Pterygoplichthys disjunctivus*, Weber, 1991: Its purification and characterization. *Food chemistry*. 141(2): 940-945.
- Younes, I., Nasri, R., Bkhairia, I., Jellouli, K., and Nasri, M. 2015. New proteases extracted from red scorpionfish (*Scorpaena scrofa*) viscera: Characterization and application as a detergent additive and for shrimp waste deproteinization. *Food and Bioproducts Processing*. 94, 453-462.

How to cite this article:

Biswajit Mohanty, R. K. Majumdar, Upasana Mohanty and Naresh Kumar Mehta. 2020. Partial Purification and Characterization of Proteases from the Visceral Waste of Indian Major Carp, *Labeo rohita* (Hamilton, 1822). *Int.J.Curr.Microbiol.App.Sci*. 9(02): 1326-1342.
doi: <https://doi.org/10.20546/ijcmas.2020.902.156>