Carcass Characteristics and Physico-chemical Properties of Broiler Meat as Influenced by Triphala in Commercial Broiler Chicken

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Abstract

The biological experiment was carried out to determine the carcass characteristics and physico-chemical properties of broiler chicken meat as influenced by Triphala supplementation in broilers. The study was carried out with three hundred commercial sexed, one day-old, broiler chicks that were randomly grouped into six treatments with five replicates for each treatment containing ten chicks per replicate. The chicks were reared from day-old to thirty five days of age in deep litter system under standard management conditions throughout the experimental period. The treatment groups were fed with basal diet without supplementation of antibiotic growth promoter/Triphala (T₁) or with supplementation of oxytetracycline at 0.004 per cent (T₂), or with supplementation of Triphala at 0.025 per cent (T₃), 0.05 per cent (T₄), 0.075 per cent (T₅) or 0.10 per cent (T₆). At the end of the experiment (35th day), one male and one female bird per replicate, totally ten birds per treatment group were randomly selected. Blood samples were collected for immunological and serum lipid profile study. The experiment revealed that there were no significant difference in carcass characteristics viz. eviscerated yield, yield of gizzard, heart and abdominal fat, except liver weight, in which it was found to be significantly (P<0.01) highest in group supplemented with Triphala at 0.10 per cent (T₆) and lowest in control (T₁). There existed no significant differences among treatment groups in physico-chemical properties of breast meat of broiler chicken due to supplementation of graded levels of Triphala for a period of five weeks.

Keywords
Physico-chemical properties
Broiler meat
Triphala

Introduction

In India, broiler industry is one of the fastest growing sectors and has attained fourth place in broiler meat production at present in the world. The country has a higher demand of animal protein due to its high population. Chicken meat has become the most common source of animal protein, because the chicken meat is cheaper and has no religious taboo for consumption. The quality of broiler meat is becoming a major issue for the poultry industry. The continued exploration of genetic potential and expansion of knowledge on poultry nutrition made the maximum market weight achievable in five weeks of age. It has all been known that Triphala (in Sanskrit, “tri” means three, and “phala” means fruits),
is an important rasayana drug in the traditional Indian System of Medicine, the Ayurveda (Baliga et al., 2012). It is a mixture of the dried powders of the three fruits such as Emblica officinalis (Amla / Nellikkaai), Terminalia chebula (Haritakil Kadukkaai) and Terminalia bellerica (Bibhitaki / Thaandrikkaai) and in which dried pericarps of the three myrobalans are included at 1:1:1 proportion. Triphala is rich in various phytochemicals including the tannins, phenolics, saponins, flavonoids, gallic acid, chebulagic acid, chebulinic acid, vitamin C etc. Triphala preparation is used in human ayurvedic treatments to cure gastro-intestinal disorders and it is considered as the “Cure for all diseases”. Considering upon the possible beneficial effects of Triphala through dietary supplementation in broiler chicken, this research trial is carried out to assess the carcass characteristics and physico-chemical properties of broiler chicken meat as influenced by Triphala in commercial broiler chicken.

Materials and Methods

Biological Experiment

The biological experiment was carried out with three hundred, sex separated and commercial (Vencobb 400) broiler chicken belonging to single hatch. The whole experimental period was divided into three phases viz., pre-starter (1 to 12 days), starter (13 to 24 days) and finisher phase (25 to 35 days). All the chicks were reared from 0 days to five weeks of age under deep litter in an open sided housing system under standard management conditions. The experimental chicks were wing banded, weighed and randomly allotted into six treatment groups. Each treatment had five replicates with ten chicks per replicate comprising fifty chicks per treatment. The experimental broiler chicks were vaccinated intra-ocularly with New Castle disease vaccine (B1 strain), Infectious Bursal disease vaccine (Intermediate Georgia live strain) and New Castle disease vaccine (‘La Sota’ strain) at 7th, 14th and 21st day of age, respectively.

Experimental diet

A basal diet was formulated according to the Vencobb standard, without supplementation of antibiotic growth promoter or Triphala. The experimental diets were prepared by supplementing the basal diet with antibiotic growth promoter (Oxytetracycline @ 0.004 per cent) or Triphala at 0.025, 0.050, 0.075 or 0.10 per cent levels. The control group (T1) was fed with basal diet without any supplementation and the treatment groups were fed with antibiotic growth promoter (T2) and Triphala at 0.025, 0.05, 0.075 and 0.10 percent (T3, T4, T5 and T6) in the basal diet, respectively.

Carcass characteristics

At the end of the experiment (35th day), one male and one female bird per replicate, totally ten birds per treatment group were randomly selected. The birds were slaughtered as per the method of Arumugam and Panda (1970). The eviscerated carcass weight, weight of gizzard, liver, heart and abdominal fat were recorded and expressed as percentage of pre-slaughter live weight.

Physico-chemical properties of meat

The breast meat samples collected during the slaughter studies were analyzed for physical and chemical properties viz., pH, water-holding capacity, extract release volume and their chemical properties viz., thiobarbituric acid and tyrosine values. The physical properties of the breast meat were analyzed on the day of slaughter (0th day). The meat was stored at -19 °C for 45 days and the
chemical properties were assessed on the breast meat at 45th day post-slaughter.

**Meat pH**

The fresh meat sample (5g) collected immediately after slaughter was minced and taken in a beaker containing 45ml of double glass distilled water (pH – 6.7). The pH of meat was estimated using Oaklon multi parameter PCSTM35.

**Water holding capacity**

The estimation of water holding capacity was carried out by adopting the filter paper press method recommended by Grau and Hamm (1957) with modification. Accurately 300mg meat was kept in-between a folded Whatman No.41 filter paper. The folded filter paper with meat was then kept in-between two glass slides. The muscle tissue was subjected to a downward force by placing a 100g weight on the top of the upper glass slide for 3 minutes. The area of the two resultant wet impressions was expressed in square centimeter.

**Extract release volume**

Accurately 20 g meat sample was homogenized with 100ml distilled water for 2 minutes. The homogenate was directly poured into a funnel lined with Whatman filter paper No. 1, folded thrice to make eight sections. The homogenate was allowed to seep between the folds and the extract was collected in 100 ml graduated cylinder for 15 minutes (Food Safety and Standards Authority of India, 2012).

**Thiobarbituric acid value**

The TBA value was estimated by using solvent trichloroacetic acid for extraction as described by Witte et al., (1970).

**Tyrosine value**

The tyrosine value was estimated from the muscle sample, as per the techniques of Strange et al., (1977). The data collected on various response criteria were subjected to statistical analysis in Completely Randomized Design (CRD) as per the methods suggested by Snedecor and Cochran (1989).

**Results and Discussion**

**Carcass characteristics and Physico-chemical properties of broiler meat**

The mean (± S.E.) carcass characteristics of broiler chicken (expressed as percentage of pre-slaughter live weight) at 35th day of age, as influenced by dietary supplementation of graded levels of Triphala is presented in Table 1. The analysis of variance of data on carcass characteristics revealed no significant difference among different treatment groups due to supplementation of Triphala, in percentage yield of eviscerated carcass, gizzard, heart and abdominal fat, except the percentage yield of liver.

The percentage yield of liver was significantly (P<0.01) highest in the group supplemented with 0.10 per cent dietary Triphala (T6) and was lowest in control group without antibiotic growth promoter or Triphala supplementation (T1). The other treatment groups supplemented either with antibiotic growth promoter (T2) or Triphala at 0.025, 0.05 and 0.075 per cent in diet (T3, T4 and T5) did not differ significantly either with T1 or T6.

The result of present study is in agreement with earlier findings of Bhattacharya et al., (2015) and Gaikwad et al., (2016) who found no significant differences in carcass dressing per cent among the treatment groups. Priya (2016) also found no significant differences in
eviscerated percentage of broilers due to supplementation of Triphala. On contrary, Shivakumar et al., (2005), Sanjyal and Sapkota (2011), and Mandal et al., (2017) obtained higher dressing percentage due to phytobiotic supplementation in the diet of broilers compared to the unsupplemented groups.

Table 1. Mean (± S.E.) Carcass characteristics of broiler chicken (expressed as percentage of pre-slaughter live weight) as influenced by dietary supplementation of graded levels of Triphala

<table>
<thead>
<tr>
<th>Treatment groups</th>
<th>Eviscerated carcass</th>
<th>Gizzard</th>
<th>Liver</th>
<th>Heart</th>
<th>Abdominal Fat</th>
</tr>
</thead>
<tbody>
<tr>
<td>T1</td>
<td>72.61 ± 1.39</td>
<td>2.10</td>
<td>2.13B</td>
<td>0.79</td>
<td>1.28 ± 0.08</td>
</tr>
<tr>
<td>T2</td>
<td>74.01 ± 1.94</td>
<td>2.30</td>
<td>2.39AB</td>
<td>0.92</td>
<td>1.24 ± 0.14</td>
</tr>
<tr>
<td>T3</td>
<td>71.96 ± 0.44</td>
<td>2.33</td>
<td>2.39AB</td>
<td>0.86</td>
<td>1.13 ± 0.07</td>
</tr>
<tr>
<td>T4</td>
<td>71.86 ± 2.13</td>
<td>2.22</td>
<td>2.35B</td>
<td>0.86</td>
<td>1.15 ± 0.10</td>
</tr>
<tr>
<td>T5</td>
<td>72.14 ± 0.31</td>
<td>2.08</td>
<td>2.17AB</td>
<td>0.79</td>
<td>0.98 ± 0.07</td>
</tr>
<tr>
<td>T6</td>
<td>71.75 ± 0.57</td>
<td>2.10</td>
<td>2.52A</td>
<td>0.84</td>
<td>1.17 ± 0.13</td>
</tr>
</tbody>
</table>

Value given in each cell is the mean of 10 observations

A,B Means within a column with no common superscript differ significantly (P<0.01)

Table 2. Mean (± S.E.) Physico-chemical properties of breast meat of broiler chicken as influenced by dietary supplementation of graded levels of Triphala

<table>
<thead>
<tr>
<th>Treatment groups</th>
<th>pH</th>
<th>Water Holding Capacity (%)</th>
<th>Extract Release Volume (ml)</th>
<th>Thiobarbituric Acid Value (mg%)</th>
<th>Tyrosine Value (mg%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>T1</td>
<td>6.32 ± 0.03</td>
<td>59.00 ± 3.71</td>
<td>32.83 ± 0.87</td>
<td>0.75 ± 0.04</td>
<td>34.12 ± 2.43</td>
</tr>
<tr>
<td>T2</td>
<td>6.26 ± 0.03</td>
<td>60.00 ± 3.27</td>
<td>31.67 ± 0.96</td>
<td>0.83 ± 0.03</td>
<td>32.92 ± 1.90</td>
</tr>
<tr>
<td>T3</td>
<td>6.25 ± 0.04</td>
<td>60.00 ± 2.25</td>
<td>32.33 ± 0.49</td>
<td>0.77 ± 0.06</td>
<td>30.56 ± 1.99</td>
</tr>
<tr>
<td>T4</td>
<td>6.25 ± 0.04</td>
<td>58.00 ± 1.63</td>
<td>31.83 ± 0.31</td>
<td>0.76 ± 0.06</td>
<td>31.96 ± 1.25</td>
</tr>
<tr>
<td>T5</td>
<td>6.32 ± 0.05</td>
<td>61.67 ± 2.60</td>
<td>32.17 ± 0.60</td>
<td>0.81 ± 0.06</td>
<td>30.01 ± 1.58</td>
</tr>
<tr>
<td>T6</td>
<td>6.30 ± 0.02</td>
<td>61.33 ± 3.52</td>
<td>31.50 ± 0.96</td>
<td>0.87 ± 0.08</td>
<td>29.20 ± 1.11</td>
</tr>
</tbody>
</table>

Value given in each cell is the mean of 6 observations

The mean (± S.E.) physico-chemical properties of breast meat of broiler chicken as influenced by dietary supplementation of graded levels of Triphala is given in the Table 2. Statistical analysis revealed no significant differences among the various treatment groups on the physical properties viz. meat pH, water holding capacity, extract release volume and chemical properties viz. thiobarbituric acid value and tyrosine value. The result of present study indicates that supplementation of either antibiotic growth promoter or Triphala at the inclusion levels has no influence on physical or chemical properties of broiler meat.
Based on the salient research findings, it was concluded that carcass characteristics revealed no significant difference among different treatment groups due to supplementation of Triphala, in percentage yield of eviscerated carcass, gizzard, heart and abdominal fat, except the percentage yield of liver. However, the percentage yield of liver was significantly (P<0.01) highest in the group supplemented with 0.10 per cent dietary Triphala (T6) and was lowest in control group without antibiotic growth promoter or Triphala supplementation (T1). Supplementation of either antibiotic growth promoter or Triphala at the inclusion levels had no influence on physical or chemical properties of broiler meat.

References


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