The Effect of Thoracic Ganglion Extracts of the Endocrine Regulation of Larval Ecdysis in Macrobrachium ламерри

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A B S T R A C T

The effect of thoracic ganglion extracts from different stages of *M. ламерри* was observed on the endocrine regulation of larval ecdysis during the larval development of *Macrobrachium ламерри*. The frequency in molting was accelerated. Adult animals were found to have more potential or more number of accelerating morphogenetic hormones.

Decapod crustacean larvae undergo a series of molts culminating in metamorphosis. Very little is known of the development of any endocrine system in crustacean larvae and the extent to which their mechanisms are altered during molting and metamorphosis. Various physiological and biochemical activities are under hormonal control which originate from endocrine and neuroendocrine systems. Compared to adults, the crustacean larvae remained unexplored to neuroendocrinological manipulations. So this piece of work was undertaken to reveal the concealed unique roles displayed by the neuroendocrine center, thoracic ganglion in the endocrine regulation of larval ecdysis in *M. ламерри*.

Introduction

Molting is the dominant metabolic event in the life cycle of these decapods crustaceans. Somatic growth in crustacean is achieved only through this periodic shedding and reformation of the rigid integument. These decapod crustacean larvae undergo several molts during their larval life.

During molt cycle the crustaceans show many cyclic structural, biochemical and physiological changes resulting in ecdysis. The duration of a molt cycle in species specific and each phase of it in larvae can be measured in hours, days or weeks.
Materials and Methods

Berried forms of *M. lamerrii* were procured from Kham river, Aurangabad. Larvae were hatched out by 29-30th day of embryonic development. They were separated into two groups of 20 each, as control and experimental. Each group was maintained in aerated glass beakers containing one liter of dechlorinated water.

Thoracic ganglion were isolated from 20 intermolt adult (mature) *M. lamerrii*. Their extracts were prepared with a few ml of distilled water, centrifuged at 2000 rpm for 10 minutes. The supernatant of the extract was added into the experimental beakers. These extracts were added daily right from the day of their hatching up to their post larval stage (15 days old). During the tenure, water was changed daily and fed with freshwater algae. Molting records were noted twice daily and the dead animals if any were removed regularly. On 16th day they were fixed in 1% formalin. Length was measured from the tip of the rostrum to end of the telson exclusive of telson spines. The weight of the animals was also noted.

The experiment was also carried out with juvenile (immature) stage prawn’s (neuroendocrine center) thoracic ganglion extracts. The difference in the neuroendocrine center potentialities of adult (mature) and juvenile (immature) were noted.

Results and Discussion

The larvae of *M. lamerrii* molt very frequently upto metamorphosis. After the metamorphic molt the frequency in molting was decreased. The 6th molt was the metamorphic molt in these larvae. The intermolt duration in control animals was 40-46 hours, whereas in thoracic ganglion extract added larvae it was 20-24 hours (Table 1).

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<thead>
<tr>
<th>S. No.</th>
<th>Experimental condition</th>
<th>Intermolt period in hours</th>
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<tbody>
<tr>
<td>1.</td>
<td>Control</td>
<td>40-46</td>
</tr>
<tr>
<td>2.</td>
<td>Thoracic ganglion extract</td>
<td>20-24</td>
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</tbody>
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<table>
<thead>
<tr>
<th>S. No.</th>
<th>Experimental condition</th>
<th>Length (mm)</th>
<th>Weight (mg)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1.</td>
<td>Control</td>
<td>5.3</td>
<td>2.5 ± 0.11</td>
</tr>
<tr>
<td>2.</td>
<td>Thoracic ganglion extract</td>
<td>5.9</td>
<td>2.65 ± 0.035</td>
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<th>S. No.</th>
<th>Experimental condition</th>
<th>Length (mm)</th>
<th>Weight (mg)</th>
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<tbody>
<tr>
<td>1.</td>
<td>Control</td>
<td>6.4</td>
<td>2.79 ± 0.02</td>
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As indicated in Table 2 and 3 the thoracic ganglion extract (6.4 mm and 2.79 ± 0.02mm) added larvae, exhibited increment in their size and weight from that of control group (5.3 ± and 2.5 ± 0.11 mg).

The results of juvenile neuroendocrine extracts were similar to that of adults. But they were not so significant as that of adult neuroendocrine extracts (Table 2 and 3).

It is well known that molting in decapod crustaceans is under the endocrine and neuroendocrine regulations. There are many reports of the presence of a stimulating hormone in central nervous system (brain and thoracic ganglion of crustaceans. Our results are in sharp concurrence with these above investigators statements. The thoracic ganglion extract added larvae exhibited the acceleration in frequency of molting and growth (size and weight) indicating that it is a source of molt (growth) accelerating substances.

In Caridina radaohari, M. Kistnensis, Penaeus indicus there are reports of the more number of stainable Neurosecretory cell during intermolt. By premolt stage there was a rise in their number and immediately after post-molt (molt) there was a sudden depletion in the number of cells with stainable neurosecretory material in all three parts of central nervous system. This depletion was observed more in thoracic ganglion than in brain and eyestalk. Such a change was also observed by us during our study of “Neurosecretory system during development and metamorphosis” in these larvae. Before (5th day, III stage) and at metamorphic molting stage (10th day, IVth stage), the number of stainable neurosecretory cells were more in all the three neuroendocrine centres. The number of cells and stainable neurosecretory cells were more in thoracic ganglion than in brain and eyestalk. After this metamorphic molt a depletion was seen in number and staining intensity (15th day, post larval stage). This depletion was observed more in thoracic ganglion than in brain and eyestalk. And also the molt acceleration (growth) of thoracic ganglion extracts added animals was higher than that of other neuroendocrine extracts. From this we can draw a conclusion that thoracic ganglion is composed of more potential morphogenetic hormones or more number of morphogenetic hormones.

The results of thoracic ganglion extracts of juveniles were similar to that of the adults. The extract added larvae exhibited the acceleration of molting frequency (growth) thus indicating the presence of an acceleratory hormone or factors in thoracic ganglion.

The results of adult (mature) animals were more significant than that of the juveniles (immature). This is indicating that the neuroendocrine hormone or factors in matured animals are more potential than that of the juveniles (immature). Thus from this piece of work we can draw a conclusion that the thoracic ganglion is composed of stimulatory morphogenetic hormones.

Acknowledgements

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