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Effect of Pre-harvest Spray of Nutrients and Growth Regulators on Quality Parameters of Sapota under Hill Zone of Karnataka, India

Vani Kumbar¹*, B.S. Shivakumar¹, M. Ganapathi², Y. Kantharaj³ and H.S. Chaitanya⁴

¹Department of Fruit Science, ²Department of Crop Physiology, ³Department of Post-Harvest Technology, COH, Mudigere, India

⁴Horticulture, Krishi Vigyan Kendra, Brahmavar, Udapi University of Agricultural and Horticultural Sciences, Shivamogga

*Corresponding author

ABSTRACT

Keywords

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An experiment was carried out at Department of Fruit science, College of Horticulture-Mudigere, University of Agricultural and Horticultural Sciences, Shivamogga during 2017-18. The objective of this study was to determine the effect of pre-harvest spray of nutrients and growth regulators on quality parameters of sapota under hill zone of Karnataka. This experiment was laid out in randomized complete block design with fifteen treatments and three replications. The chemicals were sprayed on selected sapota trees at two intervals i.e. 1st spray at 40 days before the harvest and 2nd spray at 20 days after the first spray and the harvested fruits were stored in the laboratory at room temperature. The results revealed that pre-harvest application of CaNO₃ (1%) or CaNO₃ (1.5%) were found effective for increasing total soluble solids, total sugar, reducing sugar, non-reducing sugar, ascorbic acid with minimum acidity of sapota fruits.

Introduction

Sapota (*Manilkara achras* Mill.) is tropical fruit, belongs to the family sapotaceae. It is popularly known by several names such as chiku in India, sapodilla, zapota or sapodilla plum in different regions of the world. It is native to Mexico and tropical America. India is considered to be the largest producer of sapota in the world and it is mainly cultivated for its delicious fruits. In India, the major sapota growing states are Maharashtra,

Gujarat, Karnataka, Tamil Nadu, Andra Pradesh, West Bengal, Uttar Pradesh, Punjab and Haryana. Among all the states, Karnataka is the leading sapota producing state and contributes to about 26.5 per cent of the total production in the country (NHB 2016-17). Sapota is a climacteric fruit and exhibits a sudden rise in respiration after harvest (Chundawat, 1998). The shelf life of the sapota fruit deteriorates as soon as the climacteric peak is reached.

Plant growth regulators as pre-harvest treatments have proved to be a useful tool in delaying fruit ripening. Auxins, gibberellins and cytokinins are independently or in combination are known to resist the fruit senescence by interfering the action of ethylene production during ripening (Sacher, 1973). Among various plant nutrients, calcium has its desirable effect in delaying ripening and senescence, increase in firmness, vitamin C and phenolic contents. Potassium has also received considerable attention due to enhanced production of protein, thus it improves the efficiency of nitrogenous fertilizer and increases the ability of plants to withstand stress condition. Boron is another chemical nutrient which plays major role in cell division, cell wall strengthening and development, fruit and seed development. Hence adequate boron nutrition is critical for getting high yield and quality of sapota. Therefore the present investigation was carried out to assess the response of preharvest spray of chemicals and plant growth regulators on quality attributes of sapota.

Materials and Methods

The present investigation was carried out at sapota orchard, College of Horticulture Mudigere, Chikamagalur during 2017-18. The experiment was laid out in randomized complete block design with three replications and fifteen treatments. The uniform sized sapota trees were marked and sprayed with different nutrients and plant growth regulators viz., T₁- control, T₂-GA₃ 200ppm, T₃- GA₃ 300ppm, T₄- Kinetin 50ppm, T₅- Kinetin 100ppm, T₆- 2,4-D 10ppm, T₇- 2,4-D 20ppm, T₈- Boron 0.2%, T₉- Boron 0.4%, T₁₀- CaCl₂ 1%, T₁₁- CaCl₂ 1.5%, T₁₂- Ca(NO₃)₂ 1%, T₁₃-Ca(NO₃)₂ 1.5%, T₁₄- KCl₂ 0.5% and T₁₅- KCl₂ 1% at two intervals i.e 40 days before harvest and 20 days after 1st spray. The fruits were harvested when colour of the fruit turned to light brown i.e potato colour. The harvested

fruits were brought to the laboratory and two kilograms of sapota fruits for each treatment were kept for observation at room temperature in the laboratory. The fruits were assessed at 3rd, 6th, 9th and 12th day of storage for TSS, acidity, total sugar, reducing sugar, non-reducing sugar and ascorbic acid.

Results and Discussion

Total soluble solids (Table 1) increased initially up to 9^{th} day of storage period in all the treatments and later on declined up to end of storage. The maximum TSS (19.67, 21.33, 24.97 and 23.81 o Brix) was recorded by treatment T_{12} (CaNO₃ @ 1 %) which was on par with treatment T_{13} (CaNO₃ @ 1.5%) i.e. 19.63, 20.97, 24.93 and 23.67 o Brix, respectively at 3^{rd} , 6^{th} , 9^{th} and 12^{th} day of storage period. At 9^{th} day onwards control fruits showed more shriveling and loss of appearance hence fruits were discarded.

The increase in TSS during initial storage period might be due to hydrolysis of starch into sugar on complete hydrolysis of starch no further increases occur and subsequently decline in TSS is predictable. Similar findings have been reported by Jawandha *et al.*, (2007) and Yadav *et al.*, (2009) in ber, Rajput *et al.*, (2008) in guava, Ramezanian *et al.*, (2010) in pomegranate, Bhalerao *et al.*, (2010) in sapota, Karemera and Habimana (2014) in mango and Kirmani *et al.*, (2015) in Plum.

Data on titratable acidity showed nonsignificant differences among all the treatments of sapota fruit under ambient condition (Table 1). Nevertheless, acidity showed decrease during the storage period, the decline in acidity might be due to the conversion of acid into sugar. The similar result on acidity was also reported by Gupta et al., (1987) in ber, Jayachandran et al., (2005) in guava and Bhalerao et al., (2010) in sapota. The present investigation exhibited a continuous decline in the ascorbic acid content of fruit with the increased storage duration. Data (Table 2) on ascorbic acid revealed that under ambient condition, the highest ascorbic acid (23.86, 19.11, 15.38 and 9.64 mg /100g pulp) at 3^{rd} , 6^{th} , 9^{th} and 12^{th} day of storage period, respectively was recorded in treatment T_{12} (CaNO₃ @ 1 %) which, remained at par with treatment T_{13}

(CaNO₃ @ 1.5%) i.e. 23.85, 19.07, 13.50 and 9.61 mg /100 g pulp at 3^{rd} , 6^{th} , 9^{th} and 12^{th} day of storage period, respectively.

The gradual reduction in ascorbic acid content during entire storage period might be due to its degradation through enzymatic oxidation of L-ascorbic acid to dehydro ascorbic acid during metabolic processes.

Table.1 Effect of pre-harvest foliar application of nutrients and plant growth regulators on Total soluble solids and Titratable acidity of sapota at different days after harvest

Treatments	То	tal soluble	solids (°B	rix)	Titratable acidity (%)				
	3 rd day	6 th day	9 th day	12 th day	3 rd day	6 th day	9 th day	12 th day	
T ₁ - Control	16.57	18.17	22.10		0.24	0.19	0.14		
T ₂ - GA ₃ at 200 ppm	18.27	19.93	23.47	21.24	0.19	0.16	0.11	0.08	
T ₃ - GA ₃ at 300 ppm	17.93	19.70	23.30	20.32	0.14	0.16	0.11	0.08	
T ₄ - Kinetin at 50 ppm	17.73	19.43	22.73	20.23	0.21	0.19	0.13	0.06	
T ₅ - Kinetin at 100	16.83	19.40	22.40	19.25	0.21	0.19	0.13	0.08	
ppm									
T ₆ - 2,4-D at 10 ppm	18.63	20.03	24.03		0.19	0.14	0.11		
T ₇ - 2,4-D at 20 ppm	18.53	20.00	23.87		0.19	0.16	0.11		
T₈- Boron at 0.2%	19.13	20.63	24.27		0.16	0.13	0.08		
T ₉ - Boron at 0.4%	18.83	20.40	24.26		0.16	0.13	0.08		
T ₁₀ - CaCl ₂ at 1%	19.47	20.67	24.77	23.53	0.14	0.11	0.08	0.05	
T ₁₁ - CaCl ₂ at 1.5%	19.37	20.65	24.30	22.87	0.16	0.11	0.08	0.05	
T ₁₂ - CaNO ₃ at 1%	19.67	21.33	24.97	23.81	0.13	0.11	0.08	0.05	
T ₁₃ - CaNO ₃ at 1.5%	19.63	20.97	24.93	23.67	0.14	0.11	0.08	0.05	
T ₁₄ - KCl ₂ at 0.5%	18.77	20.30	23.93		0.16	0.13	0.11		
T ₁₅ - KCl ₂ at 1%	18.70	20.17	23.97		0.19	0.14	0.12		
S.Em ±	0.57	0.43	0.09	0.30	0.00	0.01	0.01	0.00	
C.D at 1%	2.23	1.69	0.34	1.19	NS	NS	NS	NS	

Note: -- Termination of shelf life

Table.2 Effect of pre-harvest foliar application of nutrients and plant growth regulators on Ascorbic acid and Total sugar content of sapota at different days after harvest

Treatments	Ascorbic acid (mg/100g pulp)				Total sugar (%)			
	3 rd day	6 th day	9 th day	12 th day	3 rd day	6 th day	9 th day	12 th
								day
T ₁ - Control	14.20	12.60	09.38		11.46	14.92	18.05	
T ₂ - GA ₃ at 200 ppm	19.03	16.53	10.83	9.48	15.11	16.97	21.84	19.76
T ₃ - GA ₃ at 300 ppm	19.02	16.51	10.76	9.44	13.87	16.63	21.58	19.45
T ₄ - Kinetin at 50 ppm	14.51	13.31	10.64	7.25	13.21	16.19	21.25	17.76
T ₅ - Kinetin at 100	14.26	12.77	10.56	6.14	12.89	15.79	20.91	16.93
ppm								
T ₆ - 2,4-D at 10 ppm	19.06	17.13	10.90		15.40	17.86	22.51	
T ₇ - 2,4-D at 20 ppm	19.04	16.81	10.88		14.95	17.23	22.39	
T ₈ - Boron at 0.2%	19.13	17.89	12.51		16.89	19.35	22.93	
T ₉ - Boron at 0.4%	19.12	17.74	11.55		16.55	18.55	22.87	
T ₁₀ - CaCl ₂ at 1%	23.78	18.83	12.87	9.57	18.34	19.93	23.31	22.40
T ₁₁ - CaCl ₂ at 1.5%	19.14	18.26	12.75	9.56	17.93	19.77	23.25	22.09
T ₁₂ - CaNO ₃ at 1%	23.86	19.11	15.38	9.64	20.06	21.16	23.65	22.42
T ₁₃ - CaNO ₃ at 1.5%	23.85	19.07	13.50	9.61	19.25	20.09	23.50	21.75
T ₁₄ - KCl ₂ at 0.5%	19.12	17.72	11.23		16.15	18.03	22.64	
T ₁₅ - KCl ₂ at 1%	19.06	17.59	11.11		15.49	18.01	22.57	
S.Em ±	0.06	0.13	0.17	0.24	0.64	0.57	0.64	0.50
C.D at 1%	0.22	0.51	0.67	0.93	2.52	2.27	2.51	1.95

Note: -- Termination of shelf life

Table.3 Effect of pre-harvest foliar application of nutrients and plant growth regulators on Reducing sugar and Non-reducing sugar content of sapota at different days after harvest

Tuo atm onta	Reducing sugar (%)				Non-reducing sugar (%)			
Treatments	3 rd day	6 th day	9 th day	12 th day	3 rd day	6 th day	9 th day	12 th
								day
T ₁ - Control	05.49	08.07	09.06		5.97	6.85	8.99	
T ₂ - GA ₃ at 200 ppm	07.98	09.78	12.58	11.51	7.13	7.19	9.26	8.25
T ₃ - GA ₃ at 300 ppm	07.60	09.37	12.25	11.25	6.27	7.26	9.33	8.20
T ₄ - Kinetin at 50 ppm	06.84	08.63	12.17	09.64	6.37	7.56	9.08	8.12
T ₅ - Kinetin at 100	06.77	08.60	11.56	09.57	6.12	7.19	9.35	7.36
ppm								
T ₆ - 2,4-D at 10 ppm	08.20	10.07	13.04		7.2	7.79	9.47	
T ₇ - 2,4-D at 20 ppm	08.18	10.01	12.60		6.77	7.22	9.79	
T ₈ - Boron at 0.2%	09.74	11.26	13.35		7.15	8.09	9.58	
T ₉ - Boron at 0.4%	09.56	11.18	13.16		6.99	7.37	9.71	
T ₁₀ - CaCl ₂ at 1%	10.72	11.41	13.46	12.47	7.62	8.52	9.85	9.93
T ₁₁ - CaCl ₂ at 1.5%	10.19	11.29	13.44	12.45	7.74	8.48	9.81	9.64
T ₁₂ - CaNO ₃ at 1%	11.73	12.51	13.72	12.90	8.33	8.65	9.93	9.52
T ₁₃ - CaNO ₃ at 1.5%	11.08	11.47	13.63	12.51	8.17	8.62	9.87	9.24
T ₁₄ - KCl ₂ at 0.5%	08.89	10.81	13.14		7.26	7.22	9.50	
T ₁₅ - KCl ₂ at 1%	08.80	10.38	13.12		6.69	7.63	9.45	
S.Em ±	0.52	0.46	0.44	0.36	0.08	0.03	0.03	0.03
C.D at 1%	2.05	1.81	1.73	1.41	0.33	0.10	0.11	0.10

Note: -- Termination of shelf life

Similar observations was also recorded by Jawandha *et al.*, (2007) and Yadav *et al.*, (2009) in ber, Rajput *et al.*, (2008) and Bisen *et al.*, (2014) in guava, Ramezanian *et al.*, (2009) in pomegranate, Lal *et al.*, (2011) in apricot and Kirmani *et al.*, (2015) in Plum.

Total sugar (Table 2) content of sapota fruit at different stages of ripening increased

significantly from mature to ripe stage with as slight decline at overripe stage. In ambient storage the total sugar increase up to 9^{th} day of storage period and then after it started declining. The maximum total sugar (20.06, 21.16, 23.65 and 22.42 %) was observed in treatment $T_{12}(\text{CaNO}_3 \ @ 1 \ \%)$ at 3^{rd} , 6^{th} , 9^{th} and 12^{th} day of storage period, respectively which, remained at par with treatment T_{13}

(CaNO₃ @ 1.5%) i.e. 19.25, 20.09, 23.5 and 21.75 % at 3^{rd} , 6^{th} , 9^{th} and 12^{th} day of storage period, respectively.

The increase in total sugar during initial storage period might be due to hydrolysis of starch into sugar as on complete hydrolysis of starch no further increase occurs and subsequently a decline in total sugar is predictable. The present investigation is in conformity with the results reported by, Rajkumar *et al.*, (2006) in papaya, Yadav *et al.*, (2009) in ber, Bhalerao *et al.*, (2010) in sapota, Lal *et al.*, (2011) in apricot, Alila and Achumi (2012) in litchi, Karemera and Habimana (2014) in mango, Bisen *et al.*, (2014) in guava and Kirmani *et al.*, (2015) in Plum.

Accumulation of reducing sugar was gradually increased with a slight decline at the end of storage period in ambient condition (Table 3). The maximum reducing sugar (11.73, 12.51, 13.72 and 12.90 %) was observed in treatment T₁₂ (CaNO₃ @ 1%) at 3rd, 6th, 9th and 12th day of storage period respectively, followed by treatment T_{13} (CaNO₃ @ 1.5%) i.e. 11.08, 11.47, 13.63 and 12.51 % at 3rd, 6th, 9th and 12th day of ambient storage period, respectively. The increase of reducing sugar content by calcium application might be due to the less utilization of sugar during respiration and conversion of starch in to sugar, while the subsequent decline was perhaps due to consumption of sugar for respiration during storage. The present investigation is in conformity with the results reported by Yadav et al., (2009) in ber, Bhalerao et al., (2010) in sapota, Alila and Achumi (2012) in litchi, Bisen et al., (2014) in guava and Kirmani et al., (2015) in Plum.

Significant difference was observed among all the treatments with respect to non-reducing sugar. Under ambient storage, highest non-reducing sugar (8.33, 8.65, 9.93 and 9.52 %)

at 3^{rd} , 6^{th} , 9^{th} and 12^{th} day of storage period, respectively was found in treatment T_{12} (CaNO₃ @ 1 %) which, remained at par with treatment T_{13} (CaNO₃ @ 1.5 %) i.e. 8.17, 8.62, 9.87 and 9.24 %, respectively (Table 3).

The increase in non-reducing sugar during storage was due to the conversion of starch into sugar, while, the subsequent decrease in sugar was might be due to consumption of sugar for respiration during storage period. These results are in accordance with the findings of Bhalerao *et al.*, (2010) in sapota, Alila and Achumi (2012) in litchi, Bisen *et al.*, (2014) in guava, Karemera and Habimana (2014) in mango and Kirmani *et al.*, (2015) in plum.

The present study clearly indicated that preharvest spraying of CaNO₃ at (1%) or CaNO₃ at (1.5%) is effective and found promising for maintaining quality attributes viz. TSS, acidity, total sugar, reducing sugar, nonreducing sugar and ascorbic acid under ambient storage.

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