

## Original Research Article

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## Multiple Heliconia Genotypes under 50 Per Cent Shadehouse Conditions: Physiological and Biochemical Responses

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### ABSTRACT

The present investigation was carried out at College of Horticulture, Anantharajupeta during 2018-19. The experiment was laid out in Randomized Block Design, with three replications and with 8 genotypes. The treated rhizomes were planted under 50 per cent shadehouse condition. All the physiological attributes and anthocyanin content varied significantly among multiple heliconia genotypes studied under shadehouse conditions. Among different genotypes tried, leaf area (814.57 cm<sup>2</sup>), SPAD (SCMR) value for chlorophyll content (65.62), stomatal conductance (0.38 mol m<sup>-2</sup> s<sup>-1</sup>), rate of photosynthesis (9.23 μmol m<sup>-2</sup> s<sup>-1</sup>), transpiration rate (4.17 mmol m<sup>-2</sup> s<sup>-1</sup>) and anthocyanin content in flowers (3.64 mg 100 g<sup>-1</sup> tissue) recorded were highest in genotype G<sub>6</sub>. Significantly more leaf intercellular CO<sub>2</sub> (317.38 μmol m<sup>-2</sup> s<sup>-1</sup>) and leaf temperature (34.30 °C), respectively was recorded in genotype G<sub>3</sub>. While the genotype G<sub>5</sub> had maximum number of stomata (45.80 mm<sup>2</sup>).

#### Keywords

Heliconia genotypes, Physiological, Biochemical attributes, Shadehouse condition, Performance to the region

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### Introduction

Heliconia, is a rhizomatous, herbaceous perennial plant and commonly known as 'Lobster-claws', 'Wild plantains' or 'False bird of paradise'. Heliconia (*Heliconia* spp.) belongs to family Heliconiaceae and is amongst the most attractive of all the exotic tropical flowering plants, comprises of single

genus, with about 250-300 species. Heliconias are native to Central and South America, the Caribbean Islands and some of the islands of South Pacific, possessing chromosome number 2n=24. It is distributed primarily from the North of Mexico to the South of Brazil. Some species of heliconia are utilized as ornamental plants, usually being grown both as landscaping plant and as cut flowers owing

to its colour and longer durability of its floral bracts. Wide variation in vegetative growth, size, shape and arrangement of bracts has been reported by different authors (Kokila *et al.*, 2016). Due to its unusual inflorescence, heliconia is categorized as 'Specialty Flower' (Malakar *et al.*, 2015).

Colourful bracts protect the small flowers and form the inflorescence, which are used as cut flowers and landscaping featured plants. Heliconia is mostly grown for beautifying the garden, presently growing as cut flower because of its brilliant color, exotic form, long, straight peduncles and excellent postharvest characteristics, tolerance to biotic and abiotic stresses and reasonable prices. These all features made it an outstanding flower for the florist. It likes warm and humid conditions and can grow well even under partial shade (Dalawai *et al.*, 2017).

Now-a-days, research in the field of crop improvement leads to the introduction of different varieties having different forms and colors by both government and private institutions. According to flowering habit, they are grouped in different groups *viz.*, erect growing ones and pendant or hanging type. Though many genotypes of heliconia can be grown in any agro-climatic region, all of them are not suited for cut flower purpose or for garden display or for exhibition purpose. So, there is a need to evaluate hybrids and varieties in any particular agro-climatic region. Anantharajupeta falling in Southern region of Andhra Pradesh is a potential region with rich crop diversity. It is endowed with mild tropical climate and red loamy soils. Several reports of good performance by modern cut flowers are available from the location (Sirisha, 2016). To meet the growing demand for cut flowers in the fast growing areas of Rayalaseema region of Andhra Pradesh, introduction and popularization of modern flowers is necessary.

Present day varieties are available in diverse forms, types and a wide spectrum of colors. At present, the farmers are not aware of the improved varieties and are still growing only local varieties which are not only less attractive in their shape, size and colour, but also exhibit very low productivity. In heliconia, sufficient number of varieties or genotypes is under cultivation but their performance varies from place to place. The quality of cut flower is primarily a varietal trait and is generally influenced by climatic conditions prevailed during the growing period at a particular place. It is very essential to study the performance of varieties in a particular place before recommending to the farmers. Though the crop has great significance in the market, there are some bottlenecks associated in its cultivation. Non availability of planting material, lack of improved varieties, high market fluctuations are some of the other problems which are often faced by the farmers. The results obtained from this study would be a base to develop a strong breeding program for heliconias and to identify the best genotypes suitable for the region. Keeping the above in view, an investigation was planned and carried out.

### **Materials and Methods**

The present investigation was carried out during the year 2018-2019 at College of Horticulture, Dr. Y.S.R Horticultural University, Anantharajupeta, Y.S.R Kadapa Dist. of Andhra Pradesh. The trial was planned with seven genotypes collected from Horticultural Research Station, Pandirimamidi, East Godavari Dist. (Dr. Y.S.R Horticultural University, Andhra Pradesh) and one genotype is collected from local farmer at Dumpalagattu (village), Khazipet (mandal), Y.S.R Kadapa Dist. Andhra Pradesh. The experiment was laid out in Randomized Block Design, with three

replications and with 8 genotypes viz., G<sub>1</sub>- *Heliconia* cv. Golden Torch, G<sub>2</sub>- *Heliconia psittacorum rubra*, G<sub>3</sub>- *Heliconia densiflora*, G<sub>4</sub>- *Heliconia* cv. Orange By Gyro, G<sub>5</sub> - *Heliconia* cv. Alan Carle, G<sub>6</sub> - *Heliconia rostrata*, G<sub>7</sub> - *Heliconia* cv. Eden Pink and G<sub>8</sub> - Local check.

Ploughing and digging of the land was done and brought to fine tilth. The plots were prepared under shadehouse condition with 50 per cent shade. All the stubbles of previous crop and weeds were removed from the field and burnt. The required numbers of plots (24) were prepared of size (4.00 m x 3.20 m). The soil of experimental block was red loamy with pH 7.40 and E.C. 0.27 dSm<sup>-1</sup>. Rhizomes were treated with ridomil MZ 1.5 g + dimethoate 2 ml + dhanuvit 0.5 ml litre<sup>-1</sup> water before sowing. The healthy rhizomes were planted into the pits at a depth of 10-15 cm, at a spacing of 1.00 m x 0.80 m during first week of June, 2018. Rhizome planting was done in the morning hours and light irrigation was given immediately after planting. Organic manure in the form of well rotten farm yard manure was applied @ 1.5-2.0 kg per planting pit prior to rhizome sowing and mixed well. Nitrogen 20g plant<sup>-1</sup>, phosphorus 20g plant<sup>-1</sup> and potassium 20g plant<sup>-1</sup> (Dr. Y.S.R.H.U, Andhra Pradesh) were applied monthly prior to blooming. After the commencement of flowering, each clump is applied with nitrogen 10g plant<sup>-1</sup>, phosphorus 10g plant<sup>-1</sup> and potassium 20g plant<sup>-1</sup> at monthly intervals. *Heliconia* genotypes were foliar feeded with potassium nitrate (13:0:45) @ 7 g l<sup>-1</sup> at 6, 8 and 10 months after rhizome sowing. Depending upon the weather, soil and crop growth condition, watering was given to *heliconia* genotypes through drip irrigation at weekly thrice. Necessary plant protection measures were followed to prevent insect pest incidence. Five plants were selected in each plot at random, tagged and labeled properly for recording observations. The experiment

was carried out for the period from June, 2018 to May, 2019. The data recorded on various parameters were statistically analyzed by the procedure outlined by (Panse and Sukhatme, 1978).

## Results and Discussion

### Leaf area (cm<sup>2</sup>)

Eight *heliconia* genotypes showed statistically significant difference for leaf area (Table 1). Leaf area recorded was maximum in G<sub>6</sub> (814.57 cm<sup>2</sup>) which was on par with G<sub>7</sub> (700.85 cm<sup>2</sup>) and G<sub>5</sub> (606.37 cm<sup>2</sup>). While the minimum leaf area was recorded in G<sub>2</sub> (67.06 cm<sup>2</sup>). The variation in the leaf area in different varieties of *heliconia* may be due to additive gene effect, genetic makeup of the cultivars and the growing conditions. Development of optimal leaf area is important to photosynthesis and dry matter yield. The variation in leaf area might be due to increase in length of leaf, petiole, width of leaf and number of leaves among the different genotypes of *heliconia* (Dalawai *et al.*, 2017). The parallel results were reported with the studies of Sheela *et al.*, (2007) where maximum leaf area was found in *Heliconia bourgenia* x *Heliconia collinsiana* cv. 'Pedro Ortiz'.

Similarly, variation in leaf production with different length and breadth might also be due to genetic character of the genotypes. The results of present study confirmed that there is large variation in leaf area within *heliconia* genotypes and are in line with the conclusions of previous *heliconia* researchers (Aundhakar, 2008; Kadam *et al.*, 2011). Leaf length and leaf area was significantly different among different cultivars might be due to the genetic variation and difference in adaptation to the agro-climatic condition (Balaram *et al.*, 2009; Pragya *et al.*, 2010; Saleem *et al.*, 2013). Maximum leaf area may lead to more dry

matter accumulation, which resulted in the accumulation of maximum photosynthates that contributed to produce bigger sized flower or more number of flowers.

### **Chlorophyll content (SCMR values) in leaves**

Chlorophyll content in leaves was significantly influenced by multiple heliconia genotypes under shadehouse conditions (Table 1). Among the genotypes, the SCMR value recorded was highest (65.62) in G<sub>2</sub> which was on par with G<sub>7</sub> (64.95) and G<sub>5</sub> (61.30), respectively. Lowest SCMR value of 20.62 was recorded in G<sub>3</sub>. The variation in chlorophyll content of leaf among the genotypes could be attributed to the genetic constitution and varietal character. Increased foliar N concentration under shade grown plants has been associated with increased leaf chlorophyll concentration, a plant response intended to increase light capture under shaded conditions (de Groot *et al.*, 2002). Wang *et al.*, (2005) demonstrated that the chlorophyll concentration is directly related to the degree of leaf greenness, which represents another important parameter in determining the quality and marketability of the plants at the end of production. Lykas and Katsoulas (2012) also reported that use of shading nets to favour chlorophyll production in compact gardenia (*Gardenia jasminoides* Ellis.) potted plant. The shade levels of 50 per cent were optimal as a result recorded better chlorophyll content. The shade nets provide the optimal environment for growth of cut greens in subtropical condition both in winter and summer (Gaurav *et al.*, 2015). Chlorophyll content in leaf enhanced photosynthetic activity, which produce carbohydrates. Carbohydrates serve as energy source for growing bud, flower opening and longevity. The ultimate effect of all these factors resulted into strong and long flower stalks, large sized buds or flower (Tarannum and Naik, 2014).

### **Leaf intercellular CO<sub>2</sub> ( $\mu\text{mol m}^{-2} \text{s}^{-1}$ )**

The data corresponding to this trait is presented in Table 2 and significant response was observed among the genotypes. Among multiple genotypes studied for evaluation, the genotype G<sub>3</sub> recorded highest leaf intercellular CO<sub>2</sub> ( $317.38 \mu\text{mol m}^{-2} \text{s}^{-1}$ ) which was on par with G<sub>4</sub> ( $315.55 \mu\text{mol m}^{-2} \text{s}^{-1}$ ), G<sub>6</sub> ( $315.20 \mu\text{mol m}^{-2} \text{s}^{-1}$ ), G<sub>1</sub> ( $312.20 \mu\text{mol m}^{-2} \text{s}^{-1}$ ), G<sub>7</sub> ( $307.87 \mu\text{mol m}^{-2} \text{s}^{-1}$ ) and G<sub>8</sub> ( $300.00 \mu\text{mol m}^{-2} \text{s}^{-1}$ ). The lowest value for leaf intercellular CO<sub>2</sub> was recorded in G<sub>2</sub> ( $266.50 \mu\text{mol m}^{-2} \text{s}^{-1}$ ). The above findings obtained might be due to congenial micro climatic conditions prevail under 50 per cent shade level for the heliconia genotype. This statement find support from Li *et al.*, (2009) they reported that increase in shading increased the leaf intercellular CO<sub>2</sub>. Among 2 ornamental passion flower hybrids studied, Abreu *et al.*, (2014) recorded higher values for leaf intercellular CO<sub>2</sub> in *Passiflora* 'Aninha' ( $0.52 \pm 0.29 \text{ mmol (H}_2\text{O) m}^{-2} \text{s}^{-1}$ ), *Passiflora* 'Priscilla' ( $0.74 \pm 0.81 \text{ mmol (H}_2\text{O) m}^{-2} \text{s}^{-1}$ ) and in *Passiflora palmeri* var. *sublanceolata* ( $0.74 \pm 0.01 \text{ mmol (H}_2\text{O) m}^{-2} \text{s}^{-1}$ ) among 13 ornamental passion flowers (Pires *et al.*, 2011).

### **Leaf temperature ( $^{\circ}\text{C}$ )**

A critical examination of the data depicted in Table 1 revealed that leaf temperature varied significantly among eight heliconia genotypes grown under shadehouse conditions. Significantly maximum value for the above attribute was observed in G<sub>3</sub> which recorded maximum leaf temperature of  $34.30^{\circ}\text{C}$  which was on par with G<sub>7</sub> ( $33.97^{\circ}\text{C}$ ), G<sub>2</sub> ( $33.40^{\circ}\text{C}$ ), G<sub>4</sub> ( $33.28^{\circ}\text{C}$ ), G<sub>5</sub> ( $33.07^{\circ}\text{C}$ ) and G<sub>6</sub> ( $32.55^{\circ}\text{C}$ ). The lowest leaf temperature was recorded in G<sub>8</sub> ( $30.84^{\circ}\text{C}$ ). Thick leaves store more heat than do thin leaves and, consequently, have typically higher leaf temperatures (Lewis and Nobel, 1977). The results suggest, therefore, that different plant genera may depend on

different processes/traits to effectively regulate the temperature of their leaves and this is also dependent on substrate moisture availability (Gaurav *et al.*, 2015). However, temperature of thin leaves was additionally dependent on leaf color and, in pubescent leaves, the length of leaf hairs (with lighter leaf color and longer hair length being associated with lower temperatures). Conversely, in succulent leaves, temperature was mostly controlled by leaf thickness, with other simultaneously measured factors (such as leaf hairiness and leaf color) not being significant (Gaurav *et al.*, 2015). In contrast, high shade levels reduced leaf temperature (Juan, 2013).

### Number of stomata (mm<sup>2</sup>)

The data representing to number of stomata of eight genotypes is furnished in Table 2 and none of the genotypes tried responded significantly to the attribute. However the genotype G<sub>5</sub> had highest number of stomata (45.80 mm<sup>2</sup>). The minimum number of stomata was recorded in G<sub>7</sub> (29.60 mm<sup>2</sup>). It can be linked to the fact that highly pubescent leaves can have a higher number of stomata

per leaf area than do glabrous and less pubescent leaves (Skelton *et al.*, 2012). Similar to our results, higher stomatal density has been observed in rose plants (Fanourakis *et al.*, 2013) and in chrysanthemum (Aliniaiefard and Meeteren, 2016).

### Stomatal conductance (mol m<sup>-2</sup> s<sup>-1</sup>)

Analysis of data with respect to stomatal conductance is presented in Table 2. The above parameter shows significant response among 8 genotypes tried under shadehouse conditions. The genotype G<sub>6</sub> recorded the maximum stomatal conductance (0.38 mol m<sup>-2</sup> s<sup>-1</sup>) which was on par with G<sub>7</sub> (0.21 mol m<sup>-2</sup> s<sup>-1</sup>). Higher stomatal conductance in G<sub>6</sub> genotype found during current study indicates that this may tend to diffuse more CO<sub>2</sub> to chloroplast and thus have greater photosynthetic activity and produce more biomass. Stomatal conductance is a numerical measure of the rate of passage of either water vapour or carbon dioxide through the stomata. Stomatal conductance plays an important role in the plant-atmosphere water exchange and, hence, it is a key parameter in many ecological models (Chen *et al.*, 1999).

**Table.1** Physiological traits of different heliconia genotypes under shadehouse conditions

Genotypes		Leaf area plant <sup>-1</sup> (cm <sup>2</sup> )	Chlorophyll content (SCMR)	Leaf intercellular CO <sub>2</sub> (μmol m <sup>-2</sup> s <sup>-1</sup> )	Leaf temperature (°C)
G <sub>1</sub>	<i>Heliconia</i> cv. Golden Torch	505.37	49.08	312.20	32.29
G <sub>2</sub>	<i>Heliconia psittacorum rubra</i>	67.06	34.33	266.50	33.40
G <sub>3</sub>	<i>Heliconia densiflora</i>	87.40	20.62	317.38	34.30
G <sub>4</sub>	<i>Heliconia</i> cv. Orange By Gyro	288.61	41.43	315.55	33.28
G <sub>5</sub>	<i>Heliconia</i> cv. Alan Carle	606.37	61.30	275.32	33.07
G <sub>6</sub>	<i>Heliconia rostrata</i>	814.57	65.62	315.2	32.55
G <sub>7</sub>	<i>Heliconia</i> cv. Eden Pink	700.85	64.95	307.87	33.97
G <sub>8</sub>	Local Check	243.37	44.10	300.00	30.84
SEM ±		76.49	5.22	8.20	0.57
CD (P= 0.05)		234.26	16.01	25.11	1.77



**Table.2** Physiological and biochemical attributes of different heliconia genotypes under shadehouse conditions

Genotypes		No. of stomata (mm <sup>2</sup> )	Stomatal conductance (mol m <sup>-2</sup> s <sup>-1</sup> )	Photosynthetic rate (µmol m <sup>-2</sup> s <sup>-1</sup> )	Transpiration rate (mmol m <sup>-2</sup> s <sup>-1</sup> )	Anthocyanin content (mg 100g <sup>-1</sup> tissue)
G <sub>1</sub>	<i>Heliconia</i> cv. Golden Torch	35.20	0.20	8.49	2.88	2.56
G <sub>2</sub>	<i>Heliconia psittacorum rubra</i>	30.00	0.12	2.00	0.55	2.64
G <sub>3</sub>	<i>Heliconia densiflora</i>	30.60	0.04	0.81	0.45	0.00
G <sub>4</sub>	<i>Heliconia</i> cv. Orange By Gyro	30.40	0.14	3.72	1.79	0.00
G <sub>5</sub>	<i>Heliconia</i> cv. Alan Carle	45.80	0.19	7.15	2.43	1.77
G <sub>6</sub>	<i>Heliconia rostrata</i>	38.80	0.38	9.23	4.17	3.64
G <sub>7</sub>	<i>Heliconia</i> cv. Eden Pink	29.60	0.21	5.39	3.10	2.69
G <sub>8</sub>	Local Check	31.26	0.18	4.26	2.69	0.00
SEM ±		1.93	0.05	1.38	0.34	0.21
CD (P= 0.05)		N.S	0.17	4.24	1.05	0.65

Increased stomatal conductance is an indicator of higher gas exchange capacity of the leaf (Rho *et al.*, 2012). These results are consistent with recent studies that suggest that a greater distribution of diffuse radiation photons improve leaf gas exchange in several protected crops (Li *et al.*, 2009). Our results find support from Abreu *et al.*, (2014) that they recorded maximum stomatal conductance in *Passiflora* ‘Aninha’ (0.32 ± 0.05 mol (H<sub>2</sub>O) m<sup>-2</sup> s<sup>-1</sup>) and in *Passiflora* ‘Priscilla’ (0.35 ± 0.18 mol (H<sub>2</sub>O) m<sup>-2</sup> s<sup>-1</sup>). Among 3 ornamental flowers, under 50 per cent shade net conditions, Pires *et al.*, (2011) found maximum stomatal conductance (0.30 ± 0.02 mol (H<sub>2</sub>O) m<sup>-2</sup> s<sup>-1</sup>) in *Passiflora palmeri* var. *sublanceolata*. Similar opinion was put forwarded by Koniarski and Matysiak (2013) in rhododendron cultivars.

#### Rate of photosynthesis (µmol m<sup>-2</sup> s<sup>-1</sup>)

The data in Table 2 conformed that photosynthetic rate was significantly

influenced by various genotypes evaluated. Significantly maximum photosynthetic rate of 9.23 µmol m<sup>-2</sup> s<sup>-1</sup> was recorded in G<sub>6</sub> which was on par with G<sub>1</sub> (8.49µmol m<sup>-2</sup> s<sup>-1</sup>), G<sub>5</sub> (7.15µmol m<sup>-2</sup> s<sup>-1</sup>) and G<sub>7</sub> (5.39µmol m<sup>-2</sup> s<sup>-1</sup>). The photosynthetic rate was lowest in G<sub>3</sub> (0.81 µmol m<sup>-2</sup> s<sup>-1</sup>). More number of suckers results in more production of leaves, the size of the leaf and number of leaves plant<sup>-1</sup> decides the efficiency of photosynthesis activity which contributed towards better growth and yield. In the present study, heliconia plants under low light intensity were taller, indicating that plants under low light intensity may allocate more biomass to the shoot for the growth of leaves and for the full absorption of limited energy to meet the demand for plant photosynthesis (Walters and Reich, 1999). The increased leaf area of heliconia species under shadehouse condition indicates that plants increase their photosynthetic surface to contribute to a more efficient absorption of light radiation. Fifty per cent shade levels recorded higher plant

height, number of leaves and petiole length. This phenomenon is consistent with our earlier results corresponding to biometric observations including leaf area. The superior performance level was because of higher leaf chlorophyll content and photosynthetic rate (Gaurav *et al.*, 2015). Under 50 per cent shade net conditions, among 3 ornamental flowers, higher photosynthetic rate ( $21.09 \pm 0.60 \text{ mol (CO}_2\text{) m}^{-2} \text{ s}^{-1}$ ) was recorded in *Passiflora morifolia* (Pires *et al.*, 2011). The maintenance of photosynthesis in shaded plants compared with exposed trees indicated that use of screen structures in semiarid environments could help reduce plant water stress and increase water use efficiency (Barradas *et al.*, 2005). The increase in diffuse light in greenhouses or tunnels has been noted in other studies, indicating that the polyethylene used promotes a greater transformation of direct light into diffuse light, which results beneficial for photosynthesis and productivity in horticultural crops (Li *et al.*, 2009).

### **Transpiration rate ( $\text{mmol m}^{-2} \text{ s}^{-1}$ )**

A close sight of the data revealed that the genotypes demonstrated highly significant differences for transpiration rate and data is represented in Table 2. Transpiration rate recorded was highest in G<sub>6</sub> ( $4.17 \text{ mmol m}^{-2} \text{ s}^{-1}$ ) which was found to be significantly superior to all other genotypes which was followed by genotype G<sub>7</sub> ( $3.10 \text{ mmol m}^{-2} \text{ s}^{-1}$ ). In chrysanthemum Aliniaiefard and Meeteren (2016) reported similar type of finding for rate of transpiration ( $4.95 \text{ mmol m}^{-2} \text{ s}^{-1}$ ). Pires *et al.*, (2011) also recorded maximum transpiration rate in *Passiflora palmeri* var. *sublanceolata* ( $5.09 \pm 0.20 \text{ mmol (H}_2\text{O) m}^{-2} \text{ s}^{-1}$ ) among 3 types of ornamental passion flowers. Our results coincide and get support from the above authors findings. The rate of transpiration was found to be inversely proportional to shade level. Transpiration rate

is directly dependent on temperature, light intensity, relative humidity and transmittance. Moderate shading levels resulted in reduced leaf temperature and leaf transpiration without reducing net photosynthesis. This reduced leaf transpiration was likely attributed to reduced evaporative demand and probably explains the increased soil water content and reduced plant water uptake under shaded conditions. The linear relationship between net photosynthesis and stomatal conductance suggests that stomatal control of photosynthesis was substantial under shade. The increased internal CO<sub>2</sub> concentration with increased shade level also suggests, however, that there were also non-stomatal factors such as mesophyll or biochemical factors, limiting net photosynthesis (Assmann, 1992).

### **Anthocyanin content in flowers ( $\text{mg } 10 \text{ g}^{-1}$ tissue)**

The data depicted for this attribute is made available in Table 2 and heliconia genotypes differed significantly for the attribute tested. Among the genotypes studied, the genotype G<sub>6</sub> produced the highest anthocyanin content ( $3.64 \text{ mg } 100 \text{ g}^{-1}$ ) which was found significantly superior over all other genotypes which was followed by G<sub>7</sub> ( $2.69 \text{ mg } 100 \text{ g}^{-1}$ ). The remaining genotypes *viz.*, G<sub>3</sub>, G<sub>4</sub> and G<sub>8</sub> did not show anthocyanin content as they do not come for blooming during the period of study. The presence of anthocyanin and carotenoid pigments colouration have been demonstrated in the inflorescence bracts of other ornamental rhizomatic plants like heliconia (Mangave, 2010) and bird-of-paradise (Pirone *et al.*, 2010). In addition, Tatsuzawa *et al.*, (2010) reported that both anthocyanins and carotenoids significantly influenced the flower colour in different cultivars of orchids that resulted in orange-red and red flowers. Our result showed that the increase in anthocyanin contents was affected the photosynthetic pigment accumulation in

the bracts during early pigmentation. These observations indicate that photosynthetic pigments were synthesized in the first stage of inflorescence development before replaced by the different phenolic compounds. The observation is in agreement with the finding of Slatnar *et al.*, (2013) in poinsettia inflorescence bracts. Shading treatments were significantly affected the pigmentation patterns and inflorescence development of heliconia. The results are corroborated with the findings in bougainvillea (Saifuddin *et al.*, 2010), petunia (Albert *et al.*, 2009) and daisy (Meng *et al.*, 2004).

Finally, it may be concluded that, *Heliconia rostrata* (G<sub>6</sub>) was proved to be the best among the other heliconia genotypes for enhanced growth, flowering, physiological, biochemical and yield parameters. Hence it may be suitable for commercial cultivation under shadehouse conditions in Rayalaseema region of Andhra Pradesh.

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