Antibacterial Activity of Silver Nanoparticles

Gihan A. EL Batouti*

Department of Microbiology and Immunology, Faculty of Pharmacy, Pharos University in Alexandria, Egypt

*Corresponding author

ABSTRACT

Bacterial infections are a major cause of chronic infections and mortality. Drug resistant microorganisms pose a serious public health problem and strategies for controlling bacterial activity are needed. Nanoparticles have emerged as a novel alternative to overcome bacterial multidrug resistance due to misuse and abuse of antibiotics. Their use as antimicrobial agents could overcome mechanisms of bacterial resistance as the micbicidal nature of nanoparticles result from direct contact with the bacterial cell wall, inhibiting cellular adhesion and attachment, without the need to penetrate into the cell, interfering in bacterial physiology, quorum sensing, and avoiding biofilm development. The physicochemical properties of nanoparticles are significant elements that regulate their antibacterial actions. Moreover, environmental conditions, the bacterial strain, and the exposure time are other major factors that influence their effects. This review focuses on the antibacterial mechanism and effect of silver nanoparticles, as well as the potential use as an alternative antimicrobial agent.

Keywords
Antibacterial activity, Antibiotics, Silver nanoparticles

Introduction

The properties of metal nanoparticles (NPs) have been widely studied for their antimicrobial activity. The most tested metallic NP’s are silver, copper, gold, aluminum, titanium, iron, and zinc (Seil et al., 2012; Aderibigbe, 2017).

They have shown broad spectrum antimicrobial activity against Gram positive and negative bacteria, mycobacteria and fungi. However, their antibacterial activity varies among the different types of NPs well as the different microorganisms (Lesniak et al., 2013).

NPs rely on different antibacterial mechanisms when compared to antibiotics; including their size, high surface area, unusual crystal morphologies (edges and corners) and reactive sites (Slavin et al., 2017).

The low surface to volume ratio of NPs can increase their antibacterial activity allowing greater interaction of the nanomaterial with the surrounding environment (Wang et al., 2017).
Antibacterial mechanisms of silver nanoparticles

Amongst the metal NPs, silver nanoparticles (Ag-NPs) have been widely used as an effective antimicrobial agent against bacteria (Alshareef et al., 2017, Adur et al., 2018), fungi (Dojčilović et al., 2017, Kalaivani et al., 2018) and viruses (Etemadzade et al., 2016, Tamilselvan et al., 2017). Silver (Ag) and its compounds have long been used for the disinfection of medical devices and water purification. In medicine, Ag compounds are commonly applied to treat burns, wounds, and a variety of infectious diseases (Avalos et al., 2014).

The use of Ag was nearly abandoned with the discovery penicillin and later on other antibiotics. However, with the emergence of antibiotic-resistant strains, it has regained interest once again (Gaynes et al., 2017). Ag was found to be an efficient bactericidal, antibacterial agent against various pathogens in vitro and in vivo (de Simone et al., 2014).

Moreover, bacteria are less prone to develop resistance against Ag than against conventional antibiotics (Beyth et al., 2015). Classical antimicrobial agents must have the ability to reach vital molecular target sites involved in bacterial metabolism such as cell wall synthesis. In addition, it should avoid ejection by efflux pumps and molecule modification by enzymes. Ag-NPs eliminate bacteria either through their microbicidal effects such as the release of free metal ions, DNA interactions, free radical generation, or by bacterial growth inhibition coupled with the host’s immune response. (Hemeg, 2017, Al Matar et al., 2017, Slavin et al., 2017, Bassegoda et al., 2018, Katva et al., 2018)

Cell membrane damage

The contact of Ag-NPs onto the cell membrane of Gram negative bacteria results in the formation of “pits” or “pores” and thus the alteration of the cell membrane permeability, it’s rupture and hence depletion of intracellular ATP. (Durán et al., 2016, Akter et al., 2018, Zheng et al., 2018)

Release of toxic ions

Silver ions (Ag+) can react with different groups of proteins in bacteria. Ag -NPs display their antibacterial activity by releasing Ag+ and also by penetrating cells interfering in their metabolic systems. Ag+ ions can also damage DNA by inhibiting its replication. The concentrations required for their bactericidal activity is low (in the range 10^-9 mol/l) (Qing et al., 2018).

 Interruption of electron transport

The positive charge of Ag + from NPs is critical for its antibacterial activity, as it becomes attached to the negatively charged bacterial cell wall resulting in its rupture, leading to denaturation of proteins and finally cell death. In addition, Ag + can affect membrane-bound respiratory enzymes as well as efflux pumps of ions that also result in cell death. (Dakal et al., 2016, Slavin et al., 2017)

Generation of Reactive Oxygen Species

The oxidizing power of oxygen (O2) can be lethal for some bacteria. Moreover, the formation of hydrogen peroxide (H2O2) by the respiratory burst results in O2 consumption with the production of free radicals and thus the oxidation of DNA and peroxidation of cellular constituents such as proteins and lipids. Bacteria affected by reactive oxygen species will progressively lose their membrane integrity, rendering them unable to adhere to surfaces, to maintain appropriate communication with other bacteria, nor to efficiently express other cell functions (Durán et al., 2016, Dakal et al., 2016).
Table.1 Factors affecting the antibacterial mechanism of Ag -NPs

<table>
<thead>
<tr>
<th>No.</th>
<th>Factor</th>
<th>Effect of Ag -NPs</th>
<th>Study</th>
</tr>
</thead>
<tbody>
<tr>
<td>a.</td>
<td>Particle Size</td>
<td>Smaller Ag -NPs have larger specific surface areas, which result in a higher antimicrobial activity.</td>
<td>Poulose et al., 2014; Wang et al., 2017</td>
</tr>
<tr>
<td>b.</td>
<td>Particle Shape</td>
<td>Ag -NPs with different shapes can cause varying degrees of bacterial cell damage.</td>
<td>Van Dong et al., 2012</td>
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<td>c.</td>
<td>Zeta Potential</td>
<td>Ag -NPs which have a positive surface charge, are prone to being adsorbed on the bacterial surface, in contrast to their negatively charged counterparts.</td>
<td>Halder et al., 2015</td>
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<tr>
<td>d.</td>
<td>Doping Modification</td>
<td>Prevent the aggregation of Ag -NPs and allow their dispersal in aqueous environments or other hydrophilic media.</td>
<td>Hartmann et al., 2015</td>
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<tr>
<td>e.</td>
<td>Roughness</td>
<td>As the roughness of Ag-NPs increases, the size and the surface area-to-mass ratio promotes the adsorption of bacterial proteins, which is followed by a reduction in bacterial adhesion.</td>
<td>Wang et al., 2017</td>
</tr>
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<td>f.</td>
<td>Environmental Conditions</td>
<td>Different environmental conditions cause significant differences in antimicrobial activity such as temperature and pH.</td>
<td>Wang et al., 2017</td>
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Table.2 Antibacterial Activity of Ag -NPs against Common Pathogenic Bacteria

<table>
<thead>
<tr>
<th>Pathogenic Bacteria</th>
<th>Study</th>
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<tbody>
<tr>
<td>Acinetobacter baumannii</td>
<td>Lysakowska et al., 2015; Wan et al., 2016; Singh et al., 2018; Chen et al., 2019;</td>
</tr>
<tr>
<td>Escherichia coli</td>
<td>Wang et al., 2014, Dhas et al., 2014; Paredes et al., 2014; Shalaby et al., 2015; Vu et al., 2018</td>
</tr>
<tr>
<td>Enterococcus faecalis</td>
<td>Wu D et al., 2014; Alabdulmohsen and Saad, 2017; Chandra et al., 2017; Halkai et al., 2018</td>
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<td>Klebsiella pneumonia</td>
<td>Kumar et al., 2016; Chhibber et al., 2017; Hosny et al., 2017; Acharya et al., 2018</td>
</tr>
<tr>
<td>Pseudomonas aeruginosa</td>
<td>Das et al., 2016; Yuan et al., 2017; Salomoni et al., 2017; Yan et al., 2018; Punjabi et al., 2018; Liao et al., 2019</td>
</tr>
<tr>
<td>Staphylococcus aureus</td>
<td>Paredes et al., 2014; Shalaby et al., 2015; Adibhesami et al., 2017; Kang et al., 2019</td>
</tr>
<tr>
<td>Mycobacterium tuberculosis</td>
<td>Sarkar et al., 2015; Punjabi et al., 2018</td>
</tr>
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</table>

Factors affecting the antibacterial mechanism of Ag -NPs

Several factors may play a role regarding the antibacterial activity of Ag- NPs against commonly encountered pathogenic bacteria (Table 1) Peptidoglycan plays an important role in hindering the activity of the Ag- NPs, suggesting that a reduction of peptidoglycans in Gram-negative bacteria may increase their
susceptibility when exposed to Ag-NPs. On the other hand, the thick peptidoglycan layer of *Staphylococcus aureus* (S.aureus) compared to *Escherichia coli* (E. coli), prevents the penetration of Ag-NPs into the bacteria. (Wang *et al.*, 2017) In addition, the bactericidal activity of Ag-NPs with smaller dimensions (<30 nm) were found to be more effective against *Klebsiella pneumonia* (K. pneumoniae) and *S.aureus*. (Van Dong *et al.*, 2012; Wang *et al.*, 2017) Hence, the antimicrobial effect of nanoparticles can be related to the interaction with the bacterial surfaces when dealing with Gram-positive strains, whereas for Gram-negative the penetration of the particles can be expected to take place depending on size, charge and other features of the material (Halder *et al.*, 2015; Wang *et al.*, 2017).

**Antibacterial effect of Ag-NPs**

Several studies have demonstrated the antibacterial activity of Ag-NPs against the most commonly encountered pathogenic bacteria. (Table 2)

**Synergistic application of Ag-NPs with antibiotics**

It has been reported that as compared to the application of Ag-NPs alone, the combination of antibiotics with Ag-NPs complexes will release Ag+ at a higher rate.

Moreover, it has been proposed that the combination of antibiotics with Ag-NPs renders the active groups of antibiotics such as hydroxyl and amine groups to result in the conjugation of both molecules, thus increasing in the effective concentration of antibiotics at a specific site and thus producing a synergistic effect that may overcome antibacterial resistance (Dixit *et al.*, 2017, Kumar *et al.*, 2016, Rajora *et al.*, 2016, Kumar *et al.*, 2018).

Wan *et al.*, 2016, reported the synergistic effects of combining Ag-NPs and Polymyxin B or Rifampicin against drug-resistant *Acinetobacter baumannii* isolated from clinical patients. Kanamycin and Rifampycin were combined with Ag-NPs at a low concentration (5 μg/disk), and were tested against five foodborne pathogens; namely *Bacillus cereus*, *Listeria monocytogenes*, *S.aureus*, *E.coli* and *Salmonella typhimurium*. When both antibiotics and Ag-NPs were mixed, they displayed strong antibacterial activity against all pathogens, with zones of inhibition ranging in diameter from 10.62 to 14.33 mm in contrary to no inhibitory effect when applied alone at the same concentration (Patra *et al.*, 2017).

In another study, Gentamicin and Chloramphenicol were combined alone and together with Ag-NPs and were tested against *Enterococcus faecalis*, where the antimicrobial effect of Ag-NPs increased in proportion to the increase in its concentration (Katva *et al.*, 2018). The synergistic effects of with Amakacin and Ampicillin with enhanced anti-biofilm activity was recorded against multidrug resistant *Pseudomonas aeruginosa*, *E. coli*, *K. pneumoniae* isolated from burn wounds in a study conducted by Nasser, 2018.

Although numerous studies reported the effective antimicrobial activity of Ag-NPs alone and in combination with different antibiotics, further studies are required to determine the minimal inhibitory concentration (MIC) of Ag, the emergence of resistant strains, the lethal effect on biofilm, and its side effects on humans. (Baptista *et al.*, 2018)

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