Original Research Article

Study of Potassium Silicate and Silica Solubilizing Bacteria and its Impact on Yield and Quality of Sugarcane under Water Stress Condition

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A B S T R A C T

The present investigation was carried out at Sugarcane Research Station, Cuddalore on CoC25 sugarcane variety during the period of 2016-2019. This study was conducted to determine the effect of silicon nutrition on physiology, yield and quality of sugarcane under drought condition. Result showed that soil application of silica solubilizer @ 12.5 kg + 50 kg FYM/ha + sett treatment with 0.5% K2SiO3 + 2.5% urea and potash foliar spray on 15 days interval from 60 to 150 DAP showed maximum germination percentage, tiller population, leaf area, millable canes, relative water content, silica solubilizing microbial count, total chlorophyll content, nitrate reductase activity, catalase, proline, plant height, number of nodes, internode length, silicon content, carbon content, potassium content, single cane weight, commercial cane sugar per cent, cane yield and sugar yield under drought condition in both plant and ratoon crop.

Keywords
Silicon, Physiology, Sugarcane, Quality, Yield

Introduction

Sugarcane is an important cash crop & plays an important role in the country's Agricultural economy. But day by day due to various reasons sugarcane productivity is going down, on the other side, production cost is increasing heavily due to increase in fertilizer cost, Labour cost, electricity charges, etc. Deteriorating soil health is also one of the concerns of low cane yields. Sugarcane is silicon (Si) accumulating crop and responds
very well for silicon fertilization. Deficiency of silicon to sugarcane is also one of the prime reasons for low cane yields & low sugar recovery. Silicon (Si) is abundantly available nutrient in the Earth's crust & is second to oxygen. Its content in soils varies greatly and ranges from less than 01% to 45% by dry weight. Silicon is supposed to be beneficial element for the plants & not as an essential element. However Si has enhanced the growth, development & yield of many crops. The content of Si in many plants is more than N, P & K supplied through fertilizers. Sugarcane being Si accumulating crop and it absorbs silicon more than N, P & K. Normally 12 month old sugarcane crop of 100 MT/ha absorbs about 400 Kg of Si; 205 Kg of N; 55 kg of P and 275 Kg of K per ha. It has been observed that one of the reasons for low yields of sugarcane is lack of Si fertilization to the crop. The productivity of sugarcane depends on mainly soil fertility, cane variety, Number of tillers, cane weight, water management, use of organic & chemical fertilizers in balance proportion, management of pests & diseases, time of harvesting etc. It is an essential element in high yielding plant species like rice, sugarcane, cereals, legumes, vegetables, tree crops and some leafy ornamentals and many plants species. However it may be observed in limiting levels under some growing conditions. Without Silicon these plants may suffer in a similar way to any plant suffer from a major or micro-nutrient deficiency but the signs are not as evident. Silicon deficiency reduced photosynthesis, low Brix, increased disease attack and insect, increased sunburn and wilting, enhanced postharvest fall are all signs of stress.

Silicon observed to be beneficial to plants where high levels of some nutrients content e.g. manganese and iron may be present in the soil. Silicon will alleviate the reinforcement of these elements in plant tissue preventing tissue damage. Researchers have shown that Silicon have a positive effect on yield quality and quantity. This might be due to its role in ethylene inhibition, which reduces the speed of aging and death of harvested plant parts. Silicon also have a good effect on plants chlorophyll content and help plants to maintain over a longer period with better shelf life and appearance. Potassium silicate foliar application has many benefits in improving leaf erectness, and improving photosynthesis efficiency also reducing capability to lodging in grasses. In addition, it offers benefits in many agricultural applications e.g. increases growth and yield, improves strength, minimize climate stress and provides impedence to mineral stress. The great attention in this study is to show the best method and rate of adding liquid potassium silicate using foliar application and observe its effect on some growth parameters, some macro and micro nutrients content and the fodder yield of maize plants.

Chen et al.,(2010) found that applying 1.5 mM silicon to drought-stressed rice increased transpiration rate from 19% in a drought-susceptible line and 53% in a drought-resistant line. According to Shen et al., (2010), 1.7 mM silicon increased leaf relative water content from 62.3% to 80.7%, and transpiration by 29% in hydroponic soybean seedlings subjected to polyethylene glycol stress. According to Sonobe et al., (2010), 1.78 mM Si(SiO₂) in a 15% PEG 6000 (v/v) solution (to create -0.6 MPa) at 23 days increased root water uptake and root water content, even with decreased osmotic potential of roots. Ahmed et al., (2011) conclude that “silicon application may be useful to improve the drought tolerance of sorghum through the enhancement of water uptake ability.

To ascertain the interaction effect of basal silicon incorporation, setts dipping in silicon solution and foliar spraying of urea and potash on cane productivity under drought situations.
The application rate of silicon for alleviation of detrimental effects of drought on sugarcane.

**Materials and Methods**

Field experiments were conducted during the early season of 2016 to 2019 at Sugarcane Research Station, Cuddalore to evaluate the interaction effect of silicon incorporation as basal dress, setts dipping in silicon solution and foliar spraying of urea and potash on cane productivity under drought situations. The clone CoC 25 was selected as test variety the soil of the experimental site was sandy loam in texture with low available nitrogen of 73kg/ac, higher available phosphorus of 120kg/ac, available micro nutrients of Fe of 18.35ppm, Mn of 10.46ppm, Zn of 0.78ppm, Cu of 1.72 ppm present in the soil, pH of 7.6 and the EC of 0.12dSm⁻¹. In water sample pH 6.3, EC of 3.8 dSm⁻¹, HCO₃⁻ of 13.4meq/l, Cl of 24.3meq/l, SO₄²⁻ of 0.3meq/l, Ca of 6.0meq/l, Mg of 7.0meq/l, Na of 20.0meq/l, K of 5.0 meq/l, SAR of 7.84, Mg:Ca of 1.17. After the imposition of stress evading treatments the setts were transplanted to the main field. Regarding NPK fertilization, while the entire quantity of phosphorus was applied as basal, the doses of phosphorus and potassium were applied in 3 equal splits on 30 and 60 days after planting. T₁-Control (imposing of stress for 9 times i.e., once in 15 days interval from 60 to 150 DAP), T₂- T₁ + 2.5% urea and potash foliar spray on 15 days interval from 60 to 150 DAP, T₃-T₁ + Sett treatment of 0.5mM CaSiO₃ alone, T₄-T₁ + Sett treatment of 0.5% K₂SiO₃ alone, T₅-T₁ + Soil application of silica solublizer @ 12.5 kg+50kg FYM/ha, T₆- T₁ + Soil application of silica solublizer @ 12.5 kg+50kg FYM/ha + Sett treatment of 0.5mM CaSiO₃ alone + 2.5% urea and potash foliar spray on 15 days interval from 60 to 150 DAP, T₇-T₁ + Soil application of silica solublizer @ 12.5 kg+50kg FYM/ha + Sett treatment of 0.5mM CaSiO₃ alone + 2.5% urea and potash foliar spray on 15 days interval from 60 to 150 DAP. (Note: Water stress was imposed by withholding irrigation at critical period of 60-150DAP for scheduling of irrigations once in 15 days)

All the crop management practices were adopted for all the treatment plots. The data on germination, tiller population, leaf area, millable canes, relative water content, silica solubilizing microbial count, total chlorophyll content, nitrate reductase activity, catalase, proline, plant height, number of nodes, internode length, single cane weight, commercial cane sugar per cent, cane yield and sugar yield were documented and presented in Tables 1, 2, 3 and 4.

**Sett treatment with silica solubilizer**

The biofertilizer, namely silica solubilizer was obtained from the Department of Microbiology, Agricultural College and Research Institute, Madurai. Soil application was performed with silica solubilizer @ 12.5 kg/ha along with 50kg of FYM /ha.

**Silica solubilising bacterial count**

Ashby’s agar medium was used to enumerate the population of silica solubilizing bacteria by following spread plate procedure. The silica solubilizing potential of the bacteria picked out from the total silica solubilizing bacterial count plate

**Scanned electronic image protocol for silicon content in sugarcane leaves**

Leaves of each sugarcane plant were sectioned and immersed in karnovsky fixative solution in Eppendorf tubes for 24 hours. Immediately after fixation, the solutions were transferred to a cryoprotective solution (30% glycerol) for
30 minutes. The samples were then frozen in liquid nitrogen and fragmented with a scalpel on a cooled metal surface. The fragments were subsequently placed in petri dishes containing distilled water, dried on paper towels and then fixed with double-sided carbon tape onto a stub specimen holder (brass disc 12-13mm (0.5 in diameter) wrapped in aluminum foil. Some fragments were fixed with the abaxial leaf surface facing up, while others were fixed with the abaxial side down. The stubs containing the specimens were placed in a desiccator’s contain silica gel, and after three days, the stub-mounted samples were placed in a Balzers SCD 050 evaporator for plating (covering of the samples with gold). Same analysis was performed using a LEO EVO 40XVP scanning electron microscope. Several images at different magnifications were generated and digitally recorded. Photo paint software in the Corel Draw 12 package will be used for image preparation. The documented data on various observations were analyzed statistically (Fig. 1–4).

**Results and Discussion**

Soil application of silica solublizer @ 12.5 kg+50kg FYM/ha + sett treatment with 0.5% \( K_2SiO_3 \) + 2.5% urea and potash foliar spray on 15 days interval from 60 to 150 DA Precorded higher germination of 83.05%. Abro et al., (2009) also documented similar report in wheat crop. This treatment also significantly registered the maximum mean tiller population of 1,48,354/ha and 1,47,852/ha in plant and ratoon crops respectively and it is on par with soil application of silica solublizer @ 12.5 kg+50kg FYM/ha + sett treatment with 0.5mM CaSiO \(_3\) + 2.5% urea and potash foliar spray on 15 days interval from 60 to 150 DAP with plant height of 427cm and 431.56cm in plant and ratoon crops respectively. It was comparable with the control recorded the minimum mean plant height of 380cm and 360cm in plant and ratoon crops respectively. This study also found that drought stress postponed the plant growth due to reduction of photosynthesis effects. It was because of stomatal closure and reduction of water entrance into the plant ultimately caused reduction in plant height. With the increase in silicon level dry weight, height of the plant and number of tillers were increased in the rice plants (Gerami et al., 2013).

The soil application of silica solublizer @ 12.5 kg+50kg FYM/ha + sett treatment with 0.5% \( K_2SiO_3 \) + 2.5% urea and potash foliar spray on 15 days interval from 60 to 150 DAP significantly registered the maximum mean number of buds of 23.00 and 25.23 in plant and ratoon crops respectively and it is on par with soil application of silica solublizer @ 12.5 kg+50kg FYM/ha + sett treatment with 0.5mM CaSiO \(_3\) + 2.5% urea and potash foliar spray on 15 days interval from 60 to 150 DAP with 22.02 and 23.67 in plant and ratoon crops respectively.

Among the evaluated treatments the soil application of silica solublizer @ 12.5 kg+50kg FYM/ha + sett treatment with 0.5% \( K_2SiO_3 \) + 2.5% urea and potash foliar spray on 15 days interval from 60 to 150 DAP significantly registered the maximum mean plant height of 435cm and 453.23cm in plant and ratoon crops respectively and it is on par with soil application of silica solublizer @ 12.5 kg+50kg FYM/ha + sett treatment with 0.5mM CaSiO \(_3\) + 2.5% urea and potash foliar spray on 15 days interval from 60 to 150 DAP with plant height of 427cm and 431.56cm in plant and ratoon crops respectively.
respectively. The control recorded the minimum mean number of buds of 17.67 and 16.98 in plant and ratoon crops respectively.

The soil application of silica solublizer @ 12.5 kg+50kg FYM/ha + sett treatment with 0.5% K$_2$SiO$_3$ + 2.5% urea and potash foliar spray on 15 days interval from 60 to 150 DAP significantly registered the maximum mean internode length of 13.03cm and 13.42cm in plant and ratoon crops respectively. The control recorded the minimum mean internode length of 11.13cm and 10.56cm in plant and ratoon crops respectively.

The results obtained from mean comparisons for silicon contents were affected by all different rates of potassium silicate which showed that silicon application caused a significant increase in silicon concentrations. Similar results were observed by Morsy et al., (2013) and Jafarei et al., (2015) who reported similar results in wheat and bean plants and stated that with the usage of silicon, amount of silicon absorption increases in plant.

Silicon content in leaves of the plants was found to be high due to sett treatment of potassium silicate. Similar results were noticed by Salim et al., (2013) who observed that potassium silicate increased mineral nutrients (N, P, K, Ca, Mg, Zn, Mn and Fe) in wheat leaves. This might be attributed to the promotion effect of Si in translocation of those nutrients including Cu, Mo and B to the growing parts of the plant and which could enhance the growth of sugarcane as found by Huang et al., (2011). The control recorded the minimum Si, C and K of 0.2%, 51.7% and 0.5% in plant crop respectively.

Among the evaluated treatments the soil application of silica solublizer @ 12.5 kg+50kg FYM/ha + sett treatment with 0.5% K$_2$SiO$_3$ + 2.5% urea and potash foliar spray on 15 days interval from 60 to 150 DAP significantly registered the maximum mean single cane weight of 1656g and 1633g in plant and ratoon crops respectively which might be attributed to the effect of soil application of silica solublizer and accumulation of photosynthates was comparable with soil application of silica solublizer @ 12.5 kg+50kg FYM/ha + sett treatment with 0.5mM CaSiO$_3$ + 2.5% urea and potash foliar spray on 15 days interval from 60 to 150 DAP with 1517g and 1595g in plant and ratoon crops respectively.

The control recorded the minimum mean single cane weight of 1167g and 1056g in plant and ratoon crops respectively.

**Physiological indices**

**Leaf Area (cm) (Table 1)**

Among the treatments evaluated the soil application of silica solublizer @ 12.5 kg+50kg FYM/ha + sett treatment with 0.5% K$_2$SiO$_3$ + 2.5% urea and potash foliar spray on 15 days interval from 60 to 150 DAP significantly registered the maximum mean leaf area of 392.12cm$^2$ and 372.32cm$^2$ in plant and ratoon crops respectively.

Leaf area is the important in yield deciding indices since sugarcane with higher leaf area
under ample sunlight resulted high photosynthetic efficiency under ample sunlight, compared to other crops. In line with this higher ethylene evolution led to higher leaf area which resulted in greater light interception and photosynthesis (Mir et al., 2010).

The soil application of silica solublizer @ 12.5 kg+50kg FYM/ha stands next with the 379.38cm$^2$ and 369.34cm$^2$ in plant and ratoon crops respectively. The control recorded the minimum mean leaf area of 215.06cm$^2$ and 115.06cm$^2$ in plant and ratoon crops respectively.

Relative water content (%) (Table 1)

Drought stress leads to closure of stomata and it subsequently decreased the photosynthetic rate. The RWC of leaves was used to assess cellular damage as indicated by the extent of dehydration. In this study, drought stress caused a decrease in RWC content, but Si application can alleviate water stress by decreasing the transpiration. Among the treatments the soil application of silica solublizer @ 12.5 kg+50kg FYM/ha + sett treatment with 0.5% K$_2$SiO$_3$ + 2.5% urea and potash foliar spray on 15 days interval from 60 to 150 DAP significantly registered the maximum mean relative water content of 89.64%and 86.62% in plant and ratoon crops respectively.

This result is in agreement with those of Lobato et al., (2009), who showed that an increase in the Si concentration promoted an increase in the water retention of pepper leaf tissue.

The control recorded the minimum mean relative water content of 72.85% and 78% in plant and ratoon crops respectively.

Silica solubilizing microbial count (Table 2)

The soil application of silica solublizer @ 12.5 kg+50kg FYM/ha + sett treatment with 0.5% K$_2$SiO$_3$ + 2.5% urea and potash foliar spray on 15 days interval from 60 to 150 DAP significantly registered the maximum mean microbial count of 27.89 cfu x 10$^5$/g and 28.76 cfu x 10$^5$/g in plant and ratoon crops respectively and it is on par with soil application of silica solublizer @ 12.5 kg+50kg FYM/ha and soil application of silica solublizer @ 12.5 kg+50kg FYM/ha + sett treatment with 0.5mM CaSiO$_3$ + 2.5% urea and potash foliar spray on 15 days interval from 60 to 150 DAP.

Soil contains a variety of micro-organisms but a few are capable of solubilizing silicates. Silicon plays two separate functions in root cell walls, strengthening the endodermal cell walls in the mature basal region and keeping the young expanding cell walls extensible in the apical region of the roots (Hattori et al., 2003). Hence its application seems to be quite beneficial to plants grown under drought conditions by encouraging the development of profuse root system and providing protection to roots against soil drying.

Moreover, the microbes like Bacillus Caldolyticus, Bacillus Mucilaginosus var Siliceous, Proteus Mirabilis, Pseudomonas and Penicillium were found to release silica from natural silicates.

The presence of Si in soil solution renders phosphorus and potassium availability to plants reversing its fixation as Si itself prefers for phosphorus fixation sites in the soil. The control recorded the minimum mean microbial count of 9.3 cfu x 10$^5$/g and 10.35 cfu x 10$^5$/g in plant and ratoon crops respectively.
Table 1: Effect of silicon nutrients on growth and physiological parameters of sugarcane plant and ratoon crop under drought condition

<table>
<thead>
<tr>
<th>Treatments</th>
<th>Germination %</th>
<th>Tiller population (/ha)</th>
<th>Millable cane population (/ha)</th>
<th>Leaf Area (cm²)</th>
<th>Relative water content (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>T₁</td>
<td>74.43</td>
<td>107784</td>
<td>92023</td>
<td>215.06</td>
<td>72.85</td>
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<tr>
<td>T₂</td>
<td>78.19</td>
<td>117230</td>
<td>100501</td>
<td>333.39</td>
<td>84.28</td>
</tr>
<tr>
<td>T₃</td>
<td>80.43</td>
<td>129037</td>
<td>107143</td>
<td>247.36</td>
<td>81.30</td>
</tr>
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<td>T₄</td>
<td>81.07</td>
<td>137941</td>
<td>110238</td>
<td>361.60</td>
<td>85.90</td>
</tr>
<tr>
<td>T₅</td>
<td>75.28</td>
<td>146785</td>
<td>109524</td>
<td>379.38</td>
<td>85.57</td>
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<tr>
<td>T₆</td>
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<td>147167</td>
<td>112345</td>
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<td>126428</td>
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<td>CD</td>
<td>NS</td>
<td>26954</td>
<td>31656</td>
<td>95.86</td>
<td>7.33</td>
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</tbody>
</table>

Table 2: Effect of silicon nutrients on growth and physiological parameters of sugarcane plant and ratoon crop under drought condition

<table>
<thead>
<tr>
<th>Treatments</th>
<th>Silica solublizer microbial count cfu/10⁵ g⁻¹</th>
<th>Total Chlorophyll mg g⁻¹</th>
<th>Nitrate Reductase activity mg of NO₂ g⁻¹ hr⁻¹</th>
<th>Catalase μg of H₂O₂ g⁻¹ of tissue</th>
<th>Proline μg g⁻¹ of tissue</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Plant crop I</td>
<td>Plant crop II</td>
<td>Plant crop</td>
<td>Ratoon crop</td>
<td>Plant crop</td>
</tr>
<tr>
<td>T₁</td>
<td>9.30</td>
<td>10.35</td>
<td>0.0192</td>
<td>0.0185</td>
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<tr>
<td>T₂</td>
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<td>16.80</td>
<td>0.0232</td>
<td>0.0214</td>
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</tr>
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<td>T₃</td>
<td>17.43</td>
<td>18.77</td>
<td>0.0240</td>
<td>0.0221</td>
<td>93.95</td>
</tr>
<tr>
<td>T₄</td>
<td>18.03</td>
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<td>0.0236</td>
<td>0.0243</td>
<td>101.09</td>
</tr>
<tr>
<td>T₅</td>
<td>27.23</td>
<td>25.23</td>
<td>0.0242</td>
<td>0.0253</td>
<td>113.25</td>
</tr>
<tr>
<td>T₆</td>
<td>25.67</td>
<td>25.85</td>
<td>0.0283</td>
<td>0.0321</td>
<td>124.87</td>
</tr>
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<td>T₇</td>
<td>27.89</td>
<td>28.76</td>
<td>0.0364</td>
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<td>136.35</td>
</tr>
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<td>CD</td>
<td>9.02</td>
<td>8.97</td>
<td>0.01</td>
<td>0.009</td>
<td>8.28</td>
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</table>
Table.3 Effect of silicon nutrients on growth and physiological parameters of sugarcane plant and ratoon crop at harvest stage under field condition

<table>
<thead>
<tr>
<th>Treatments</th>
<th>Plant height (cm)</th>
<th>Number of buds</th>
<th>Internodal length (cm)</th>
<th>Macro nutrients and Si content</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Plant crop</td>
<td>Ratoon crop</td>
<td>Plant crop</td>
<td>Ratoon crop</td>
</tr>
<tr>
<td>T₁</td>
<td>380.00</td>
<td>360.23</td>
<td>17.67</td>
<td>16.98</td>
</tr>
<tr>
<td>T₂</td>
<td>396.66</td>
<td>403.45</td>
<td>21.00</td>
<td>20.34</td>
</tr>
<tr>
<td>T₃</td>
<td>400.00</td>
<td>421.25</td>
<td>18.33</td>
<td>19.29</td>
</tr>
<tr>
<td>T₄</td>
<td>388.33</td>
<td>396.29</td>
<td>18.67</td>
<td>18.78</td>
</tr>
<tr>
<td>T₅</td>
<td>403.33</td>
<td>423.08</td>
<td>20.00</td>
<td>22.54</td>
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<td>T₆</td>
<td>427.00</td>
<td>431.56</td>
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<td>23.67</td>
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<tr>
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<td>435.00</td>
<td>453.23</td>
<td>23.00</td>
<td>25.23</td>
</tr>
<tr>
<td>CD</td>
<td>24.5</td>
<td>21.3</td>
<td>3.26</td>
<td>3.54</td>
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</table>

Table.4 Effect of silicon nutrients on yield parameters of sugarcane plant and ratoon crop at harvest stage under field condition

<table>
<thead>
<tr>
<th>Treatments</th>
<th>Single cane weight (g)</th>
<th>Cane yield (t/ha)</th>
<th>CCS (%)</th>
<th>Sugar yield (t/ha)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Plant crop</td>
<td>Ratoon crop</td>
<td>Plant crop</td>
<td>Ratoon crop</td>
</tr>
<tr>
<td>T₁</td>
<td>1167</td>
<td>1056</td>
<td>92.61</td>
<td>72.42</td>
</tr>
<tr>
<td>T₂</td>
<td>1467</td>
<td>1324</td>
<td>117.89</td>
<td>114.30</td>
</tr>
<tr>
<td>T₃</td>
<td>1383</td>
<td>1564</td>
<td>118.62</td>
<td>126.40</td>
</tr>
<tr>
<td>T₄</td>
<td>1500</td>
<td>1532</td>
<td>117.90</td>
<td>117.34</td>
</tr>
<tr>
<td>T₅</td>
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<td>1517</td>
<td>1595</td>
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<td>T₇</td>
<td>1656</td>
<td>1633</td>
<td>128.52</td>
<td>127.37</td>
</tr>
<tr>
<td>CD</td>
<td>384.9</td>
<td>393.0</td>
<td>5.87</td>
<td>6.63</td>
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Fig.1 Effect of silicon nutrition on growth of sugarcane crop at germination stage under drought condition

Fig.2 Silica solubilizing potential of silica solubilizer in Ashby’s agar medium. The diameter of the clearing zone formed was 3.2 cm
**Fig. 3** Effect of silicon nutrition on growth of sugarcane crop on Silicon, Carbon and Potassium content through SEM image under drought condition

Soil application of silica solublizer@12.5kg along with 50kg FYM/ha + sett treatment of 0.5% K2SiO3 alone +2.5% urea and potash foliar spray on 15 days interval from 60 to 150 DAP
Fig.4 Effect of silicon nutrition on growth of sugarcane crop at harvest stage under drought condition

Total chlorophyll content (mg/g) (Table 2)

Among the treatments, the soil application of silica solublizer @ 12.5 kg+50kg FYM/ha + sett treatment with 0.5% K₂SiO₃ + 2.5% urea and potash foliar spray on 15 days interval from 60 to 150 DAP significantly registered the maximum mean total chlorophyll content of 0.0364mg/g and 0.0372mg/g in plant and ratoon crops respectively and it is on par with soil application of silica solublizer @ 12.5 kg+50kg FYM/ha + sett treatment with 0.5mM CaSiO₃ + 2.5% urea and potash foliar spray on 15 days interval from 60 to 150 DAP with 0.0283mg/g and 0.0321mg/g of chlorophyll in respective crops. Si could increase the photosynthesis of wheat plants under drought and this might be associated with the enhancement in activities of photosynthetic enzymes and chlorophyll content under stress conditions (Li et al., 2007) and sorghum (Hattori et al., 2005).

Further, Kaufman et al., (1979) found that Si as silica bodies deposited in leaf epidermis act as a “window” that enhances the light use efficiency by facilitating the transmission of light to the photosynthetic mesophyll tissues. Ahmed et al., (2011) also ascertained that increase in silicon leads to increase in leaf area index, specific leaf weight, chlorophyll content, leaf dry weight, root dry weight, total dry weight under stress in sorghum. The control recorded the minimum mean total...
chlorophyll content of 0.0192mg/g and 0.0185mg/g in plant and ratoon crops respectively.

**Catalase (µg of H₂O₂/g of tissue) (Table 2)**

Among the treatments, the soil application of silica solublizer @ 12.5 kg+50kg FYM/ha + sett treatment with 0.5% K₂SiO₃ + 2.5% urea and potash foliar spray on 15 days interval from 60 to 150 DAP significantly registered the minimum mean catalase content of 5.82 and 6.22µg of H₂O₂/g of tissue in both plant and ratoon crops respectively and it is on par with soil application of silica solublizer @ 12.5 kg+50kg FYM/ha + sett treatment with 0.5mM CaSiO₃ + 2.5% urea and potash foliar spray on 15 days interval from 60 to 150 DAP treatment. In metabolic processes plants produce H₂O₂ which causes damage to the cell oxidation function, while CAT can eliminate H₂O₂ and play a key role in the elimination of O₂. The combined action of CAT and SOD converts the toxic O₂ superoxide radical and H₂O₂ to water and molecular oxygen (O₂), thus averting the cellular damage under unfavourable conditions (Noctor *et al.*, 2000). The control recorded the highest catalase content of 7.41 and 9.12 µg of H₂O₂/g of tissue in plant and ratoon crops respectively.

**Nitrate Reductase activity (mg of NO₂/g/hr) (Table 2)**

Among the evaluated treatments the soil application of silica solublizer @ 12.5 kg+50kg FYM/ha + sett treatment with 0.5% K₂SiO₃ + 2.5% urea and potash foliar spray on 15 days interval from 60 to 150 DAP significantly registered the maximum mean nitrate reductase activity of 136.35 and 134.39 mg of NO₂/g/hr in plant and ratoon crops respectively. The nitrate reductase (NRase) activity was drastically reduced in sugarcane cultivars under drought stress conditions. The lower NRase activity could probably be due to reduced storage pool size of NO₃ and reduced NO₃ flux from roots to leaves. The decrease in NRase activity in leaves by external supply of PEG may be related to osmotic changes following PEG addition into the medium.

**Proline content (µg/g) (Table 2)**

Soil application of silica solublizer @ 12.5 kg+50kg FYM/ha + sett treatment with 0.5% K₂SiO₃ + 2.5% urea and potash foliar spray on 15 days interval from 60 to 150 DAP significantly registered the maximum mean proline content of 352.31 and 352.10 µg/g of tissue in plant and ratoon crops respectively. The control recorded the minimum mean proline content of 196.68 and 178.08 µg/g of tissue in plant and ratoon crops respectively. The nitrate reductase (NRase) activity was drastically reduced in sugarcane cultivars under drought stress conditions. The lower NRase activity could probably be due to reduced storage pool size of NO₃ and reduced NO₃ flux from roots to leaves. The decrease in NRase activity in leaves by external supply of PEG may be related to osmotic changes following PEG addition into the medium.

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ratoon crops respectively and it is on par with soil application of silica solublizer @ 12.5 kg+50kg FYM/ha + sett treatment with 0.5mM CaSiO$_3$ + 2.5% urea and potash foliar spray on 15 days interval from 60 to 150 DAP with 1,12,345/ha and 1,23,678/ha in respective crops.

The soil application of silica solublizer @ 12.5 kg+50kg FYM/ha + sett treatment with 0.5% K$_2$SiO$_3$ + 2.5% urea and potash foliar spray on 15 days interval from 60 to 150 DAP (T$_7$) significantly registered the maximum cane yield of 128.52t/ha and 127.37t/ha in plant and ratoon crops respectively. However, it was on par with the soil application of silica solublizer @ 12.5 kg+50kg FYM/ha with 0.5mM CaSiO$_3$ + 2.5% urea and potash foliar spray on 15 days interval from 60 to 150 DAP (T$_{7-}$) with 125.94t/ha and 123.32t/ha respectively in plant crop and in ratoon crops. The control recorded the minimum mean cane yield of 92.61t/ha and 72.42t/ha in plant and ratoon crops respectively.

These results agreement with Ren et al., (2002) findings who found that Si application increased yield of sugarcane due to increased utilization rate and absorbing ability of nutrients. The yield increment attributed to the direct beneficial effects of Si such as vegetative growth, increase of chlorophyll content, photosynthetic activity of plant, protective enzymes and water metabolism and to some indirect effects such as acquisition of macro- and micronutrients.

Soil application of silica solublizer @ 12.5 kg+50kg FYM/ha + sett treatment with 0.5% K$_2$SiO$_3$ + 2.5% urea and potash foliar spray on 15 days interval from 60 to 150 DAP significantly registered the maximum mean commercial cane sugar of 11.78% and 11.84% in plant and ratoon crops respectively. However, it was on par with the soil application of silica solublizer @ 12.5 kg+50kg FYM/ha + sett treatment with 0.5mM CaSiO$_3$ + 2.5% urea and potash foliar spray on 15 days interval from 60 to 150 DAP with 11.08% and 11.56% and 2.5% urea and potash foliar spray on 15 days interval from 60 to 150 DAP with 10.73% and 10.12% respectively in plant crop and in ratoon crops. The control recorded the minimum CCS% of 10.32% and 9.96% in plant and ratoon crops respectively.

Among the evaluated treatments soil application of silica solublizer @ 12.5 kg+50kg FYM/ha + sett treatment with 0.5% K$_2$SiO$_3$ + 2.5% urea and potash foliar spray on 15 days interval from 60 to 150 DAP significantly registered the maximum sugar yield of 15.26t/ha and 15.39t/ha in plant and ratoon crops respectively. However, it was on par with the soil application of silica solublizer @ 12.5 kg+50kg FYM/ha with 15.11t/ha and 15.18t/ha respectively in plant and ratoon crops. The control recorded the minimum mean sugar yield of 9.32t/ha and 8.26t/ha in respective crops.

Soil application of silica solublizer @ 12.5 kg + 50 kg FYM/ha + sett treatment with 0.5% K$_2$SiO$_3$ + 2.5% urea and potash foliar spray on 15 days interval from 60 to 150 DAP increase the growth and physiological parameters of sugarcane and increase the cane productivity in terms of cane and sugar yield under drought situations.

References


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