

Original Research Article

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To Find out the Toxicity of Insecticides, Bio-Pesticides and Plant Product against *Helicoverpa armigera* (Hubner) under Laboratory Conditions

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ABSTRACT

The laboratory experiment was conducted at RARI, Durgapura during rabi 2017 to find out the toxicity of insecticides, bio-pesticides & plant product against *H. armigera* were reared on gram leaves and pods as well as on artificial diets. The nucleus culture of *H. armigera* was maintained in the laboratory under controlled conditions (26 ± 2°C and 80 ± 5 percent RH). The 12 days old larvae of *H. armigera* use as a test insect by residue film method for bioassay. Various concentration of treatments were prepare and mortality data obtained were corrected using Abbot's formula (Abbott's 1925) The treatments were divided into three categories viz., insecticides, bio- pesticides and plant products. The result shows that, the LC₅₀ value of Quinalphos was (0.054005%) shows its superiority over the acephate with LC₅₀ of 0.227715 per cent against *H. armigera*. Among the bio-pesticides the LC₅₀ value of spinosad (1.0256087%) which was lower than the b.t.k. (3.86555%) and diflubenzuron (5.37584%), so it was most effective as compared to other bio-pesticides. Among various plant products the LC₅₀ value of neem oil (1.5738827%) and eucalyptus oil (3.2800034%) thus; neem oil shows its superiority over other plant products. So order of toxicity of different insecticides, bio-pesticides and plant products as under: Insecticides: Quinalphos > Acephate, Bio-pesticides: Spinosad > B.t.k. > Difiubnzuron, Plant product: Neem oil > Karanj oil > Mahua oil > Eucalyptus oil.

Keywords

Helicoverpa armigera, Toxicity, Gram pod borer, Bioassay

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Introduction

Chickpea (*Cicer arietinum*) is one of the most important pulse crops of India. This is widely cultivated Rabi crop in India and Rajasthan. Among the biotic constraints in its production, the losses caused by insect the pod borer, *Helicoverpa armigera* (Hubner) is the main constraint during the flowering and pod stage. The yield loss in chickpea due to pod borer was 10-60 % and it is 50 per cent under

favourable weather conditions (Bhatt and Patel, 2001). Due to ease of availability and ease of application farmer respond to chemical method for controlling the insect pest to reduce pod borer infestation. However, The use of conventional insecticides causes sudden decrease in natural enemies also. Frequent and high doses of insecticides posed the problems of resistance and resurgence of the pest. Keeping these in view, the present study was undertaken to study the effectiveness of eco-

safe and to test the relative efficacy of some insecticides, bio-pesticides and botanical with conventional insecticides under laboratory conditions.

Materials and Methods

The laboratory experiment was conducted at RARI, Durgapura during rabi 2017 with the object of finding out the toxicity of insecticides, bio-pesticides and plant products against *H. armigera*. The nucleus culture of *H. armigera* was maintained in the laboratory under controlled condition ($26 \pm 2^{\circ}\text{C}$ and 80 ± 5 percent RH) from one to three pairs of *H. armigera* moths which were collected from light trap during night. Adults of *H. armigera* (male and female) were confined in matting cages ($50 \times 25 \times 25 \text{ cm}^3$). Cotton plug dipped in 10% sucrose solution was provided in the cage. After copulation, the gravid female was transferred to glass jar lined with blotting paper. A cotton wool plug soaked with 10% sucrose solution was also placed. The glass jar was kept in a tray filled with water to save them from ant, spider etc. Eggs laid on the blotting paper were collected date wise and kept in jar and plastic container for hatching. The newly hatched larvae were transferred with a camel hair brush to new set of plastic container ($6 \text{ cm} \times 6 \text{ cm}$) having leaves and pods of gram. The larvae were transferred to cleaned new containers every day and used containers were washed with detergent and sterilized by rinsing with 1 percent formaldehyde solution. The larvae attaining pupal stage were transferred date wise to enamel trays containing 4 cm thick layer of moist sand soil. The trays were examined after 5 days to collect the pupae. The normal healthy pupae collected from the trays were kept date wise in sterilize battery jars. The mouth of the jars was covered with muslin cloth and tightened with rubber bands. The adults so emerged were kept in separate jars in the same manner for mass multiplication up to five generation.

Mass rearing of *H. armigera* on semi synthetic diet

The larvae of *H. armigera* were reared to find out toxicity of insecticides, bio-pesticides and plant product. Rearing was carried out on gram leaves and pods as well as on the artificial diet. The diet formulation use is as per the Anonymous (1995) for rearing the culture. The ingredients used for preparation of semi-synthetic diet have been presented in the table 1.

Diet preparation procedure

Water (390 ml) was mixed with fraction A of the diet in the blender which was run for two minutes. Fraction B was boiled in the remaining 390 ml water. Fraction "A" and B were mixed and blender is run again for 1 minute. Finally, fraction C was added to the mixture of A and B in water and the blender run again for 1 minute. Formaldehyde solution was added in the end. The diet was poured as per requirement either on the nylon mesh for rearing up to 5-7 days old larvae or in tray cells for rearing the larvae above 5-8 days or poured into sterilized petri plates and allowed to solidify. The diet so prepared could be stored in the refrigerators up to 2 weeks.

Mass rearing procedure on semi-synthetic diet

For starting culture on artificial diet, newly hatched larvae were transferred from laboratory culture. Initially, they were transferred to a wide mouth glass jar containing fresh gram leaves and pods. Mouth of jars was covered with a piece of transparent plastic sheet perforated finely with pins. The jar was covered with black cloth to allow larvae to settle on the leaf/pods. Next day fresh pods were placed below the older ones. Most of the larvae developed into third instar within six days. The third instar larvae were

transferred to the cage containing plastic dishes with the artificial diet and then covered by black cloth, the larvae approached the food through the wire screen support and fed from below. The excrement dropped through the screen. Regular cleaning was carried out and fresh supply of food served as and when needed. The fully mature larvae entered the soil for pupation. The soil was sieved to separate pupation carefully.

Laboratory evaluation

For this purpose various insecticides (Quinalphos, acephate), biopesticides (*Bacillus thuringiensis* var. Kurstakii (B.t.k), spinosad and diflubenzuron (dimlin) and plant products (neem oil, karanj oil, mahua oil, eucalyptus oil) were taken.

Bioassay technique

The bioassay was carried out using 12 days old larvae of *H. armigera* as a test insect by residue film method. Various concentration of above treatments was prepared in distilled water and a thin film was prepared on the upper and lower surface of the petriplate. Then the 12 day old larvae of *H. armigera* were

released for 24 hours. After 24 hours the mortality was noted. The moribund larvae were also considered as dead. Control was also run simultaneously. The mortality data so obtained were corrected using Abbot's formula (Abbott's, 1925)

$$P = \frac{T - C}{100 - C} \times 100$$

Where,
 P = Corrected per cent mortality
 T = Observed per cent mortality in treatment
 C = per cent mortality in control

The corrected per cent mortality data thus obtained from different concentration of each treatment was subjected to Probit analysis (Finney, 1971) for computing LC₅₀.

Results and Discussion

The toxicity of different insecticides, bio-pesticides and plant products against larvae of *Helicoverpa armigera* (Hubner) under laboratory conditions. The treatments were divided into three categories viz., insecticides, bio-pesticides and plant products.

Table.1 composition of semi-synthetic diet for rearing the larvae of *H. armigera*

	Item/Ingredients	Quantity
A	Chickpea (kabuli gram)flour	105 g
A	Methyl para-hydroxybenzoate	2 g
A	Sorbic acid	1 g
A	Streptomycin sulphate	0.25 g
A	10 % formaldehyde solution	2 ml
B	Agar-agar	12.75
C	Yeast tablets	25 (Tablets)
C	Ascorbic acid	3.25 g
C	Multivitaplex	2 capsules
C	Vitamin E	2 g
C	Distilled water	780 ml
C	Miscellaneous	---

Table.2 Relative toxicity of different insecticides, bio-pesticides and plant products against third instar (12 days old) larvae of *H. armigera*

Treatment	Heterogeneity	Regression equation	LC ₅₀ (%)	Fiducial limit	Relative toxicity	Remark
Insecticides						
Quinalphos	2.5141	Y =2.0358X +- 2.598	0.054005	0.06572 0.04437	4.21	Within insecticides
Acephate	5.5996	Y =2.0061X + - 3.741	0.227715	0.28288 0.18330	1.00	
Biopesticides						
Spinosad	2.1917927	Y =0.9962X +0.008	1.025608	1.51018 0.69652	5.24	Within bio-pesticides
Bt (HALT)	3.7633203	Y =3.5208X +- 14.672	3.865553	4.34655 3.43778	1.39	
Diflubenzuron (Dimlin)	0.7228064	Y = 1.4840X +- 3.504	5.375841	8.21406 3.51831	1.00	
Plant products						
Neem oil	0.6766981	Y =1.6338X +- 3.491	1.573882	2.02716 1.22195	2.08	Within plant product
Karanj oil	1.7403561	Y =1.7596X +-4.374	2.125132	2.65716 1.69962	1.54	
Mahua oil	1.63615455	Y =2.0370X +- 6.075	2.734821	3.42410 2.18429	1.20	
Eucalyptus oil	6.9783746	Y =1.9235X +- 5.610	3.280003	4.13731 2.60033 3.43778	1.00	

The toxicity of different treatments was worked out by conducting bioassay against 3rd instar larvae of *H. armigera* in laboratory by residue film method. The results obtained are presented in Table 2. The LC₅₀ value of Quinalphos, acephate, diflubenzuron, spinosad, *b.t.k.*, neem oil, karanj oil, mahua oil and eucalyptus oil were 0.054005, 0.227715, 5.37584, 1.025608, 3.86555, 1.573882, 2.125132, 2.73482 and 3.28000 per cent, respectively. Quinalphos with LC₅₀ (0.054005%) shows its superiority over the acephate with LC₅₀ of 0.227715 per cent against *H.armigera*. Among the bio-pesticides the LC₅₀ value of spinosad (1.0256087%) which was lower than the *b.t.k.* (3.86555%) and diflubenzuron (5.37584%), so it was most effective as compared to other biopesticides. Among various plant products the LC₅₀ value of neem oil (1.5738827%) and eucalyptus oil (3.2800034%) thus; neem oil shows its superiority over other plant products. So order of toxicity of different insecticides, bio-pesticides and plant products as under:

Insecticides: Quinalphos > Acephate
Bio-pesticides: Spinosad > *B.t.k.* > Difiubnzuron
Plant product: Neem oil > Karanj oil > Mahua oil > Eucalyptus oil

The toxicity, different treatments were divided into three categories viz., insecticides, bio-pesticides and plant products. The toxicity of different treatments was worked out in laboratory by bioassay method against third instar larvae of *H. armigera*. On the basis of LC₅₀ values against third instar larvae, the insecticides in descending order of toxicity were arranged as: insecticides: Quinalphos (0.054005%) > acephate (0.227715%), bio-pesticides: spinosad (1.573882%), >*b.t.k.* (3.86555%) > difubenzuron (5.37584%) and Plant products: neem oil (1.573882) > karanj oil (2.125132%) > mahua oil (2.7348%) > eucalyptus oil (3.28000%) (Table 2). In the

present investigation it was observed that Quinalphos was more toxic than acephate against third instar larvae of *H.armigera*. these finding are in conformity with earlier work of justin *et al.*,(1989). In present study the LC₅₀ value of *B.t.k.* was 3.86555 per cent under laboratory condition, it get support from the work of Reddy *et al.*, (1997) who reported the effectiveness of *B.t.k.* against *H.armigera* and found that the medium lethal concentration LC₅₀ value for *B.t.k.* against third instar larvae of *H. armigera* gave 90 per cent mortality. In present investigation on the basis of the LC₅₀ value of different plant products, the eucalyptus oil was found to be less toxic as compared to other plant products. Jain and Gupta (1995) reported the effectiveness of insecticides, bio-pesticides and various plant products against *H.armigera*. The order of toxicity of different insecticides, bio-pesticides and plant products against larval instar of *H.armigera* was found as judo (LC₅₀ 0.001965-0.003376 per cent) > Dipel (LC₅₀0.002498-0.0041113 per cent) > decamethrin (LC₅₀ 0.003064-0.004625 per cent) > NPV (LC₅₀ 0.1078-0.2269) > neemax (LC₅₀ 2.1032- 3.4745) > green commandos (LC₅₀ 2.4945- 3.7489 per cent)

These finding are also in conformity with the finding of Kohja and Gupta (1992) who reported the LC₅₀ value of azadit 0.04518-0.05277 per cent against *H. armigera*. Bajpai and Sehgal (1998) observed 50 per cent mortality of neonate (24 hrs. old) larvae of *H.armigera* when semi- synthetic diets were treated with neem, karanj and tobacco formulations under laboratory conditions.

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