

Review Article

<https://doi.org/10.20546/ijcmas.2019.801.322>

Genetic Improvement of Berseem (*Trifolium alexandrinum*) in India: Current Status and Prospects

Tejveer Singh^{1*}, A. Radhakrishna¹, D. Seva Nayak¹ and D.R. Malaviya²

¹ICAR- Indian Grassland and Fodder Research Institute, Jhansi-284 003, India

²ICAR- Indian Institute of Sugarcane Research, Lucknow - 226 002, India

*Corresponding author

ABSTRACT

Berseem, a nitrogen-fixing, annual, multicut forage crop cultivated around 2 million hectares areas of northern, central and eastern parts of India. Berseem has variability for pollination behavior however variation for morphological and agronomic traits are scarce, perhaps, because of initially introduction of crop with narrow genetic base. Genetic variability have been fortified through introduction of exotic materials, intra-interspecific hybridization, induction of polyploidy and mutation. ICAR-Indian Grassland and Fodder Research Institute maintain >900 accessions of *Trifolium* spp. Different genetic improvement programmes in India resulted with development of >15 cultivars apart from unique agro-morphological variants. Affinity of Berseem was tested with other species and suitable donors for introgression of genes especially for biotic stress were identified. By adopting embryo rescue technique, wide crosses of Berseem with *T. constantinopolitanum*, *T. apertum*, *T. resupitanum* and *T. vavilovii* successfully developed and genes for biotic stresses and agro-morphological traits were incorporated. Longer duration, an important agronomic trait in Indian condition, has been induced through induction of mutation by physical mutagens. Induction of autopolyploidy by using colchicine treatment made major breakthrough in berseem breeding in India by the development of high biomass producing cultivars. The future breeding strategies contemplate to intensification of gene pool through exotics from the centre of origin, increase of variability, development of genomic resources, development of inbreds, remodeling of breeding procedure as substantial points.

Keywords

Cross incompatibility, Genomic resources, Gene pool, Polyploidy, Wide hybridization

Article Info

Accepted:

26 December 2018

Available Online:

10 January 2019

Introduction

The genus *Trifolium* from the tribe Trifolieae of the family Leguminosae (Fabaceae) is important for its agricultural value. A few of the 237 species of this large genus have actually been cultivated to date (Zohary and Heller, 1984), out of which 25 species are

agriculturally important as cultivated and pasture crops (Lange and Schifino-Wittmann, 2000). Berseem or Egyptian clover (*Trifolium alexandrinum* 2n=2x=16) is commonly cultivated as winter annuals in the tropical and subtropical regions. Berseem, introduced in India from Egypt in 1904, started cultivation as a rotational crops at government cattle

farm, since 1910 its cultivation was taken up by cultivators (Das Gupta, 1943). Berseem has been established as one of the best *Rabi* (winter season) fodder crop in entire North West Zone, Hill Zone and part of Central and Eastern Zone of the country (Mehta and Swaminathan, 1965; Singh, 1988), occupy more than two million hectare (Pandey *et al.*, 2011). Berseem are popular due to its multicut (4-8 cuts) nature, providing fodder for a long duration (November to May), very high quantum of green fodder (85 t/ha) and better quality of fodder (20% crude protein), high digestibility (up to 65%) and palatability.

Pollination behavior

Understanding the natural mating behavior (self- or cross-pollination) is important for designing a suitable breeding strategy for genetic improvement of crop. The Berseem crop is a dilemma with regard to its self and cross pollination.

In Indian conditions, a number of reports on pollination in Berseem suggest that this crop is not self-sterile but tripping is essential for a good seed-set (Chowdhury *et al.*, 1966, Roy *et al.*, 2005). The crop is predominantly self-pollinated and shows wide diversity for self fertility and population with self compatible and self pollinating, self compatible requiring tripping, self incompatible with broad genetic base and self incompatible with narrow genetic base have been identified (Dixit *et al.*, 1988). Roy *et al.*, (2005) indicated considerable variation between different populations of Berseem for self-compatibility, together with a requirement of tripping for pollination and seed set, even in self-compatible lines. Extant of natural cross pollination was reported up to 4.73% by Beri *et al.*, (1985a) and seed setting were higher under un-caged condition against caged condition due to tripping mechanism done by honey bees (Beri *et al.*, 1985b)

Germplasm management

In India, National Bureau of Plant Genetic Resources (NBPGR), New Delhi is the nodal organization for exchange, quarantine, collection, conservation, evaluation and the systematic documentation of plant genetic resources. It has introduced >500 accessions of *Trifolium* spp from different countries and maintained in long term storage conditions. The Indian Grassland and Fodder Research Institute (IGFRI), Jhansi is a National Active Germplasm Site for the systematic management and utilization of germplasm wealth of forage crops including agro-forestry trees maintains >900 accessions of *Trifolium* spp. For effective utilization and maintenance of conserved germplasm, IGFRI has catalogue the information on different qualitative, quantitative and origin place of all 594 accessions. To assist the utilization of Berseem germplasm by curators/scientists throughout the international plant genetic resources network, IGFRI has developed descriptor list of Berseem (Roy *et al.*, 2009). Many of the Berseem germplasm having unique characteristics have been generated (Singh *et al.*, 2017) and registered at NBPGR, New Delhi (Table 1).

Breeding approaches

Berseem is an introduced crop in India and one of the most important drawbacks in genetic improvement of Berseem is lack of genetic variability (Verma and Mishra, 1995; Roy *et al.*, 2004; Malaviya *et al.*, 2005; Malaviya *et al.*, 2007). Variability in the existing gene pool of Berseem has been induced in through mutation, polyploidization and interspecific hybridization. Different genetic improve programmes by utilizing breeding approaches like selection, polyploidy and mutation leads to the development of >15 varieties for different berseem growing regions of India (Table 2). High biomass

production potential alongwith extended growth period and resistance to biotic stresses specially root rot and stem rot are the main target traits has to be improved genetically.

Inter-specific hybridization

Initially, the aim of interspecific hybridization was to clarify the closest relatives of *T. alexandrinum*. *T. alexandrinum* ($2n = 16$) was successfully hybridized with *T. berytheum* ($2n = 16$) and *T. salmoneum* ($2n = 16$) and found the most probable parent. Recently, efforts has been put into using this approach with the aim of improving *T. alexandrinum*'s resistance to biotic and abiotic stresses, tolerance to soil alkalinity and length of the vegetative period. Genes for wide scale adaptability and disease resistance widely distributed in several wild species of *Trifolium* (Table 3) could not be incorporated into the present day cultivars because of interspecific incompatibility barrier which are common among other *Trifolium* species also. Embryo culture has been effectively used in developing interspecific hybrids of Berseem with *Trifolium apertum* (Malaviya *et al.*, 2004), *T. constantinopolitanum* (Roy *et al.*, 2004), *T. resupinatum* (Kaushal *et al.*, 2005) and *T. vesiculosum* (Kaur *et al.*, 2017). Progenies of interspecific hybrids showed introgression of various desirable traits, including late flowering and resistance to root rot and stem rot diseases.

Ploidy manipulation

A major breakthrough in Berseem breeding in India was achieved through induction of polyploidy. The work on polyploidization of Berseem genome was started with the aim to induce grater leaf and stem size (Mehta and Swaminathan, 1957; Sikka *et al.*, 1959). Autotetraploid induced by using colchicine treatment, and selection at tetraploid level resulted the development of first Berseem variety 'Pusa Giant' with more fodder

production and good regeneration capacity, uniform and higher yield throughout the season than diploid varieties released for general cultivation in India (Mehta and Swaminathan, 1965). Another big achievement in polyploidy breeding was achieved at IGFRI, Jhansi by developing an autotetraploid variety namey 'Bundel Berseem-3' through colchiploidy followed by recurrent single plant selection followed with mass selection. It is released for north east zone, Bihar Orissa, WB and eastern UP.

Mutation breeding

Major constraints in genetic improvement of Berseem are narrow genetic base of the crop coupled with cross incompatibility barriers (Malaviya *et al.*). Efforts have been made to generate variation in the existing gene pool through mutation by using physical or chemical mutagens (Sindhu and Mahindiratta 1976: Jatasra *et al.*, 1980; Shukla and Tripathi, 1984). Major success was achieved by induction of longer duration mutant in Mescavi variety through gamma ray treatment (Sohoo *et al.*, 1985). These longer duration mutant in the form of BL-22 a variety released in 1988 for temperate and north west zone; BL-180 released in 2006 for cultivation in north-west zone of India. Longer duration (flowering in May-June) is the important agronomic trait in Indian condition. Incorporation of this trait in Berseem variety for additional cut of green fodder during the scarcity period of summer months may be achieved.

Biotic and abiotic stress tolerance

Berseem cultivars are susceptible to diseases like root rot (*Rhizoctonia solani* and *Fusarium semitactum*), stem rot (*Scferotinia trifoliorum*), leaf blight (*Epicocum* sp.) powdery mildew (*Oidium* sp.) and downy mildew (*Perenospora trifolif*) (Bhaskar *et al.*, 2002).

Table.1 Novel genetic stock of Berseem (*Trifolium alexandrinum*) registered with NBPGR, New Delhi

s.no.	Trait	Ploidy level	Year	INGR number
1	Purple leaf and flower	Diploid (2n=2X=16)	2005	NGR05017
2	Pentafoliate Berseem Penta 1	Diploid (2n=2X=16)	2009	INGR 09045
3	Pentafoliate	Tetraploid (2n=4X=32)	2012	INGR 12010
4	Black seeded	Diploid (2n=2X=16)	2012	INGR 12009
5	Self incompatible	Tetraploid (2n=4X=32)	2012	INGR 12011
6	Self compatible	Diploid (2n=2X=16)	2012	INGR 12012
7	Blackseeded pentafoliate	Diploid (2n=2X=16)	2016	INGR 15026

Table.2 Berseem varieties released/notified in India

S. No.	Variety	Breeding method	Year of release/ notification	Institution responsible for the development	Area of adaptation
1.	Mescavi	Selection	1975	CCS HAU, Hisar	Entire growing area
2.	Pusa Giant	Polyploidy breeding	1975	IARI	Entire growing area
3.	BL 1	Selection	1978	PAU, Ludhiana	Punjab, H.P., Jammu
4.	Wardan	Selection	1982	IGFRI, Jhansi	Entire growing area
5	Jawahar Berseem 1 (JB 1)	Selection	1981	JNKVV, Jabalpur	Central India, central and north western zones
6	BL-10	Mutation breeding	1985	PAU, Ludhiana	Punjab, Haryana, H.P., Jammu
7.	BL 22	Mutation breeding	1988	PAU, Ludhiana	Sub-temperate, hill regions of North India
8.	BL 2	Selection	1989	PAU, Ludhiana	Punjab, Haryana, H.P., Jammu, Western UP, Utrakhand
9.	UPB 10	Composite Selection	1993	GBPUAandT, Pantnagar	North-west India
10.	Bundel Berseem 2	Mass Selection	1997	IGFRI, Jhansi	Central, North-west zone
11.	Bundel Berseem 3	Polyploidy breeding	2001	IGFRI, Jhansi	North-east , Eastern region
12.	JB-5	Polyploidy breeding	2005	JNKVV, Jabalpur	Central, North-west zone
13.	BL 42	Mutation breeding	2003	PAU, Ludhiana	North-west India
14.	BL 180	Mutation breeding	2006	PAU, Ludhiana	North-west India
15	Hisar Berseem 1 (HFB 600)	Selection	2006	CCS HAU, Hisar	North-west India
16.	JBSC-1	Selection	2017	IGFRI, Jhansi	Central, North-west zone

Table.3 Desirable characters in Berseem ecotypes and wild *Trifolium* species

Species	Gene pool	Chromosome Number (2n)	Desirable characters	References
<i>T. alexandrinum</i> ecotype Mescavi	Primary	2n=16	Annual, multicut, highly productive, crude protein, high digestibility and palatability, basal branching	Malaviya et al., 2004
<i>T. alexandrinum</i> ecotype Fahli	Primary	2n=16	Annual, single cut, self compatible, stem branching	Singh et al., 2015
<i>T. alexandrinum</i> ecotype Saidi	Primary	2n=16	Annual, 2-3 cut, stem and basal branching	
<i>T. berytheum</i>	Secondary	2n=16	-	Putiyevksy and Katznelson, 1973
<i>T. salmoneum</i>	Secondary	2n=16	-	Putiyevksy and Katznelson, 1973
<i>T. apertum</i>	Secondary	2n=16	Annual, profuse basal branching, late flowering, resistance against root rot and stem rot, high protein content	Putiyevksy and Katznelson, 1973; Malaviya et al., 2004
<i>T. meironense</i>	Secondary	2n=16		Putiyevksy and Katznelson, 1973
<i>T. resupinatum</i>	Tertiary	2n=16	Root rot and stem rot resistance, soil alkalinity tolerance	Bhaskar et al., 2002; Kaushal et al., 2005
<i>T. constantinopolitanum</i>	Tertiary	2n=16	Profuse basal branching, resistance against root rot and stem rot	Roy et al., 2004
<i>T. vesiculosum</i>	Tertiary	2n=16	Lateness, disease resistance	Malaviya et al., 2004

Berseem cultivar ‘Bundel Berseem-3’ developed through polyploidy is moderately tolerant to the root rot and stem rot disease. Berseem is highly sensitive to drought conditions as it decreased plant fresh and dry matter yield (Sevanayak *et al.*, 2017).

Berseem cultivars and inbreds derived from interspecific hybrids were evaluated under drought stress condition and drought tolerance lines identified (Shipra *et al.*, 2010, Sevanayak *et al.*, 2017). Further, different species of *Trifolium* from secondary and tertiary gene pool are known to be resistance against various biotic and abiotic stresses (Table 3) and could be utilized for introgression of desirable genes by utilizing advanced molecular techniques.

Biotechnological approach

Biotechnological approaches offering alternative and effective tools for genetic improvement of crop plants. Utilization of biotechnological approaches in genetic improvement by genetic transformation and other means requires efficient method for plant regeneration via tissue culture using different parts of plant. Protocol for in vitro plant regeneration from meristematic tissue and the establishment of regenerable callus culture have been developed in Berseem and related species viz., *Trifolium glomeratum*, *T. apertum*, *T. resupinatum* (Kaushal *et al.*, 2004, Kaushal *et al.*, 2006). Embryo rescue technique has been effectively utilized to overcome the problems of post fertilization

barriers in interspecific crosses of Berseem with *Trifolium apertum*, *T. constantinopolitanum*, *T. resupinatum* and *T. vesiculosum* (Malaviya *et al.*, 2004; Kaushal *et al.*, 2005; Roy *et al.*, 2004; Kaur *et al.*, 2017). Limited availability of genomic resources in Berseem hampered the utilization of molecular markers in genetic improvement programme. Therefore, molecular markers were developed and validated in Berseem (Verma *et al.*, 2017, Chandra 2011). Genetic diversity in Berseem and related *Trifolium* species were studied by using isozymes (Malaviya *et al.*, 2005) and molecular markers (Kalia *et al.*, 2009).

Future prospects and conclusion

Berseem being an important forage crop providing nutritional security to the animals by producing high quantum of quality green forage, also contributing to the sustainability of rice-wheat cropping system. Being an introduced crop in India, genetic improvement in this crop is hampered by narrow genetic base and lack of variability in desirable traits. Further introduction of germplasm from its origin place, development of interspecific hybrids and induction of mutations will further contribute in broadening the genetic base of berseem. Desirable variants developed through interspecific hybridization and mutation could be utilized in development of improved cultivars. Development of inbreds and further establishment of heterotic pool will help in development high biomass producing synthetic and composite population and hybrids. There is a scope to further strengthen the genomic resources by developing more SSR markers, molecular linkage map and mapping of forage quality and biomass contributing traits which could be utilized to speed up conventional breeding programme going on in different research institutes/universities.

References

- Badr A, El-Shazly HH, Watson LE, 2008. Origin and ancestry of Egyptian clover (*Trifolium alexandrinum* L.) as revealed by AFLP markers. *Genet Resour Crop Evol* 55:21–31
- Bakheit B.R. (1989) Pollination and seed setting in different genotypes of Egyptian clover Berseem (*Trifolium alexandrinum* L.) Assiut Journal of Agricultural Sciences. 20: 1, 199-208
- Bakiheit. B.R. (1996) Development of a New Multifoliate Strain of Berseem Clover (*Trifolium alexandrinum* L.) in Egypt, *Journal of Agronomy and Crop Science*. 177, 139-141.
- Beri SM, M. S. Sohoo M. S., H. L. Sharma, HL (1985a) Estimates of natural cross pollination in Egyptian clover. *Euphytica*, 34 (1): 147-151
- Beri SM, M. S. Sohoo, H. L. Sharma (1985b). Mode of pollination and seed setting in Egyptian clover. *Euphytica*, 34(3): 745-750.
- Bhaskar R.B., Malavlya D. R., Roy A. K. and Kaushal P. 2002. Evaluation of exotic *Trifolium* accessions for disease incidence and resistance. *Abstr. Nat. Symp. On Grassland and Fodder Research in the New Millennium held at IGFRI, Jhansi*. Pp. 31-32.
- Bhowal M., Cherian K. J. and Das L. (2011) Direct organogenesis in fodder crop *Trifolium alexandrinum* L., *Journal of Environmental Research And Development*, Vol. 5 (4):892-897.
- Chandra, A. (2011) Use of EST database markers from *M. truncatula* in the transferability to other forage legumes. *J. Environ. Biol.* 32, 347-354.
- Chaudhry AR, Inam-ul-Haq and Nadeem Rehman A (1992) Cultural approach towards control of Berseem root rot, *Pak, J. Agric. Science*, 29 (1): 65-68.
- Choo TM, Reinbergs E, Kasha KJ (1985) Use of haploids in breeding barley. *Plant Breed Rev* 3: 220–252

- Chowdhury J. B., Mehta R. K., Joshi A. B. 1966. Pollination in Berseem. Indian Journal of Genetics and Plant Breeding, 26 (1): 118-120.
- Chowdhury, J. B., Mehta, R. K., and Joshi, A. B. 1966. Pollination in Berseem. Indian Jour. Genet. Plant Breed. 26: 118-120.
- Das Gupta NC (1943) Green Berseem as a substitute for concentrates or economic feeding of dairy cattle. Indian journal of veterinary science and animal husbandry. 13:196-213.
- Dhaliwal, J.S. and Atwal A.S. (1976). Note on the effect of air temperature, relative humidity and wind velocity on bees visiting Berseem at Ludhiana. Indian Journal of Agriculture Sciences, 46: 50-51.
- Dixit OP, U. P. Singh and J. N. Gupta (1989) Significance of Pollination in Seed Setting Efficiency of Berseem (*Trifolium alexandrinum* L.), Journal of Agronomy and Crop Science, 162(2): 93-96
- Ellison NW, Liston A, Steiner JJ, Williams WM, Taylor NL (2006) Molecular phylogenetics of the clover genus (*Trifolium*-Leguminosae). Mol Phylog Evol 39:688-70.
- Gupta, Pc; Karwasra, Sso, 1982: Epicoccum leaf spot—a new disease of Berseem *Trifolium alexandrinum* Egyptian clover. Indian phytopathology 35(3): 538-539
- Jobshy, Z.M., E.I. Syed, A. Rammah and MA. Satter, 1981. Pathogenicity and control of three fungi associated with damping off and root rot of Egyptian clover *Trifolium alexandrinum*, Res. Bull. No. 1674, p. 14.
- Kaila V, V. K. Sood, H. K. Chaudhary, J. C. Bhandari, Archit Sood and R. K. Mittal (2009) Morphological and RAPD markers – mediated assessment of genetic diversity amongst various *Trifolium* species and identification of potential ideotypes for genetic upgradation of Berseem under changed climate in mid-hills of north-west Himalayas. Indian J. Genet., 69(4): 1-6 (2009)
- Kaur, A., Kaur, K.P., Kalia, A. *et al.*, (2017). Generation of interspecific hybrids between *Trifolium vesiculosum* and *T. alexandrinum* using embryo rescue Euphytica 213: 253. <https://doi.org/10.1007/s10681-017-2042-x>
- Kaushal P, DR Malaviya, Roy AK, Aparna Tiwari, B Kumar (2005) *Trifolium alexandrinum* × *T. resupinatum* – Interspecific Hybrids Developed through Embryo Rescue. Plant Cell Tissue and Organ Culture 83(2):137-144
- Kaushal P, A. Tiwari, A.K. Roy, D.R. Malaviya and B. Kumar (2006) In vitro regeneration of *Trifolium glomeratum*. Biologia Plantarum 50 (4): 693-696
- Kaushal P, D. R. Malaviya, A. K. Roy and B. Kumar (2004) In Vitro Response of Immature Zygotic Embryos of *Trifolium alexandrinum* (Egyptian clover), Plant Cell Biotechnology and Molecular Biology, 5(3-4): 109-114
- Knight WE (1985) Miscellaneous annual clovers. In Taylor NL (edn) Clovers Science and Technology. Agron Monog 25:547-561
- Kumar, B., Malaviya, D.R., Roy A.K. and Kaushal P. (2003). Protein profile and species relationship in *Trifolium*. Indian J. Genet., 63(1): 41-44.
- Lange O and MT Schifino-Wittmann (2000) Isozyme variation in wild and cultivated species of the genus *Trifolium* L. (Leguminosae). Ann. Bot. 86: 339-345.
- Malaviya D.R. A. K. Roy P. Kaushal B. Kumar A. Tiwari C. Lorenzoni (2004) Development and characterization of interspecific hybrids of *Trifolium alexandrinum* X *T. apertum* using embryo rescue. Plant Breeding 123:536 – 542. DOI.10.1111/j.1439-0523.2004.01042.x
- Malaviya DR, B. Kumar, A.K. Roy, P. Kaushal and Aparna Tiwari (2005). Estimation of variability of five enzyme systems among wild and cultivated species of *Trifolium*, Genetic Resources and Crop Evolution (2005). 52: 967-976
- Malaviya DR, Roy AK, Aparna Tiwari, P

- Kaushal, B Kumar (2006) *In vitro* Callusing and Regeneration in *Trifolium resupinatum*—A Fodder Legume Cytologia 71(3):229-235. DOI10.1508/cytologia.71.229
- Malaviya, D.R., Pandey, K.C., Roy, A.K. and Kaushal, P. (1999). Role of honey bees in seed setting of Egyptian clover. *Crop Improvement*, 26(2): 204-207.
- Malaviya, D.R., Roy, A.K., Kaushal, P., Bhaskar, R.B. and Kumar B. (2004). Evaluation of *Trifolium* species for defining multiple use gene pool for tropical *Trifolium* species. *Indian J. Genet.*, 64(3): 251-252
- Mehta R., Swaminathan M. (1957) Studies on induced polyploids in forage crops. 1. Survey of previous work. *Ind. J. Genet. PL Breed* 17:27-57.
- Narayanan, E. S., Sharma, P. L., Phadke, K. G. 1961. Studies on requirements of various crops for insect pollination – insect pollinators of erseem – Berseem clover (*Trifolium alexandrinum*) with particular reference to honey bees and their role in seed setting. *Indian Bee Jour.* 23(4/6): 23-30.
- Pandey KC and Roy AK 2011. Forage Crops Varieties. IGFRI Jhansi (India).
- Putiyevsky E, Katznelson J (1973) Cytogenetic studies in *Trifolium* spp. Related to Berseem. I. Intra- and inter-specific hybrid seed formation. *Theor Appl Genet* 43:351–358
- Roy AK, Aparna Tiwari, DR Malaviya, P Kaushal, B Kumar (2005) Regeneration Efficiency in *Trifolium apertum* as Influenced by Explant Media Interaction. *Cytologia* 70(2):153-159
- Roy AK, D R Malaviya, P Kaushal, A Chandra, U P Singh (2009) Descriptors for Tropical Forage Legume – Egyptian clover/Berseem *Trifolium alexandrinum* L., IGFRI, Jhansi
- Roy AK, D. R. Malaviya and P. Kaushal (2005) Pollination behaviour among different breeding populations of Egyptian clover. *Plant Breeding*, 124 (2):171–175
- Roy AK, D. R. Malaviya, P. Kaushal, B. Kumar, A. Tiwari (2004). Interspecific hybridization of *Trifolium alexandrinum* with *T. constantinopolitanum* using embryo rescue. *Plant Cell Rep*, 22:705–710
- Roy AK, DR Malaviya and P Kaushal (2005) Pollination Berseem among different breeding populations of Egyptian clover. *Plant Breeding* 124: 171-175.
- Roy AK, DR Malaviya, P Kaushal (2011) Generation of Interspecific Hybrids of *Trifolium* Using Embryo Rescue Techniques. *Methods in molecular biology* (Clifton, N.J.) 710:141-51. DOI10.1007/978-1-61737-988-8_12
- Roy, A.K., Malaviya, D.R. and Kaushal, P. (2005). Pollination Berseem among different breeding populations of Egyptian clover. *Plant Breeding*, 124(2):171-175.
- Roy, A.K., Malaviya, D.R., Kaushal, P., Chandra, A and Singh, U.P. (2009). Descriptors for tropical forage legume Egyptian clover (*Trifolium alexandrinum* L.), *Indian Grasses and Forest Research Station, Jhansi, India.*
- SevaNayak D., Singh, T. and Radhakrishna A. (2017). Effect of drought stress on biomass and drought adaptive traits in Berseem (*Trifolium alexandrinum* L.), National symposium “new directions in managing forage resources and livestock productivity in 21st century: challenges and opportunity” P4-17
- Shipra Agarwal, Bhupendra Singh, Alka and P Kumar. Salinity Tolerance of some cultivars of Berseem (*Trifolium alexandrinum* L.) during Germination and Early Seedling Growth. *Vegetos. Vol. 23* (1): 63 – 82 (2010)
- Shukla GP and BD Patil (1985) Breeding Egyptian Clover – A Review. *Forage Research* 11(1): 1-19.
- Sidhu BS and PD Mehndiratta (1976) Genetic analysis of factors influencing forage yield *Berseem. Indian J. Agric.*, 46: 168-170.
- Sikka S. M., Mehta R. K., Swaminathan M. S. (1959) Studies on Induced Polyploids in

- Forage Crops II. Colchicine Treatment Methods for Berseem and Senji. *Indian Journal of Genetics and Plant Breeding*, 19(1): 90-97
- Singh, T., Malaviya, D. R. and Kaushal P (2015) Genetic analysis of some morphological traits in Egyptian clover (*Trifolium alexandrinum* L.) Sustainable use of grassland resources for forage production, biodiversity and environmental protection: Extended Abstracts 23rd International Grassland Congress, Editors: A K Roy, R V Kumar, R K Agrawal, S K Mahanta, J B Singh, M M Das, K K Dwivedi, G. Prabhu, N. K. Shah. Paper ID: 1467
- Singh, T., Radhakrishna, A., SevaNayak D. and Malaviya D.R. (2017). Identification and characterization of novel variability developed through wide hybridization in Egyptian clover (*Trifolium alexandrinum* L.), National symposium “new directions in managing forage resources and livestock productivity in 21st century: challenges and opportunity” P2-24
- Sohoo, M.S.; Beri, S.M.; Bhardwaj, B.L. (1985) Radiation induced late flowering and high yielding mutant in Berseem (*Trifolium alexandrinum* L.). *Journal of Nuclear Agriculture and Biology*; v. 14(4) p. 136-138
- Taylor NL (1985) Clovers around the World. In TL Taylor (ed): *Clover Science and Technology*. Agron Monog 25:1–6
- Verma P, A Chandra, AK Roy, DR Malaviya, P Kaushal, D Pandey, S Bhatia (2015) Development, characterization and cross-species transferability of genomic SSR markers in Berseem (*Trifolium alexandrinum* L.), an important multi-cut annual forage legume. *Mol Breeding* (2015) 35:23
- Verma, JS and Mishra SN, 1995. Advances in Forage Improvement in Upper Gangetic Plains. In: *New Vistas in Forage Production*. Hazra CR, Mishri B (eds). All India Coordinated Project for Research on Forage Crops, Indian Grassland and Fodder Research Institute Jhansi-284003 (U P), India, pp 83-96.
- Zayed E.M., Magda I. Soliman, G. A. Ramadan and M. M. Tarrad (2010) Molecular characterization of two cultivars of Egyptian clover (*Trifolium alexandrinum* L.). *Range Mgmt. and Agroforestry* 31 (2): 140-143, 2010
- Zohary M (1972) A revision of the species of *Trifolium* sect. *Trifolium* (Leguminosae), II Taxonomic treatments. *Candollea* 27:99–158
- Zohary M (1972) Origins and evolution in the genus *Trifolium*. *Bot Not* 125:501–511
- Zohary M, Heller D (1984) The genus *Trifolium*. The Israel Academy of Sciences and Humanities, Jerusalem.

How to cite this article:

Tejveer Singh, A. Radhakrishna, D. Seva Nayak and Malaviya, D.R. 2019. Genetic Improvement of Berseem (*Trifolium alexandrinum*) in India: Current Status and Prospects. *Int.J.Curr.Microbiol.App.Sci*. 8(01): 3028-3036. doi: <https://doi.org/10.20546/ijcmas.2019.801.322>