

Review Article

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Trichoderma in Agriculture: An Overview of Global Scenario on Research and its Application

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ABSTRACT

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The non-judicious use of agrochemicals has created a vicious impact on the environment and its inhabitants which is a major concern among society these days; although, thinking and attitude towards its use has changed and there is a growing interest to develop its alternative. On the other hand, the use of microorganisms that antagonize plant pathogens (biological control) is a risk-free approach and the species of Trichoderma are well recognized as agents for plant disease control in addition to its promontory effect on plant growth. Presently a large number of Trichoderma based products are available in the market globally which are dominated by *T. harzianum* and *T. viridae*. This review is focused on various mechanisms of bio-control employed by Trichoderma, current status of research and its applications in agriculture with the objective to create awareness about this fungus in society to popularize its beneficial service to the environment and its future development.

Introduction

The organism that suppresses the pathogen or its activity is referred as biological control agent (BCA) and phenomenon known as bio-control or biological control. The species of Trichoderma are universally known for their bio-control potential in addition to plant growth and development (Vinale *et al.*, 2014). An effective and potential BCA (microbial pesticide) must have some unique characteristics for successful bio-control of plant diseases i.e. i) good shelf life and survival in the soil in its inactive or active

form, ii) host specificity, iii) easy to multiply in laboratory and iv) ecofriendly and cost effective. The phenomenon of bio-control employed by Trichoderma is governed by a variety of mechanism either its direct presence at the infection site or passively due to its activities which triggers and activates the various biochemical pathways in plant system. Hence, lot of research has been focused on characterizing the unique mechanism(s) used by a particular BCA operating in different experimental situations. The variety of registered microbial agents *viz.* bacteria (*Pseudomonas aureofaciens*,

Pseudomonas fluorescens, *Bacillus subtilis*, *Agrobacterium radiobacter*, *Streptomyces griseoviridis*), nematodes, protozoa viruses and fungi (*Ectomycorrhizae* sp., *Cladosporium* sp. *Ampelomyces quisqualis*, *Coniothyrium minitans*, *Aspergillus niger* IARI isolate) including *Trichoderma* species (*T. harzianum*, *T. viridiae*, *T. virens*) are being utilized in plant disease control as the bio-pesticide (Gardener and Fravel, 2002; Koul, 2011). In this review, various mechanisms employed by *Trichoderma*, present scenario of research and its application are being discussed.

Direct mechanism

Mycoparasitism/Hyperparasitism

More than seventy reports on mycoparasitic ability of *Hypocrea/Trichoderma* species are documented against phytopathogenic fungi such as *Alternaria alternata*, *Botrytis cinerea*, *Rhizoctonia solani*, *Sclerotinia sclerotiorum*, *Pythium* spp., *Phytophthora* sp. and *Fusarium* spp. (Harman 2006; Druzhinina *et al.*, 2011; Atanasova *et al.*, 2013; Bae *et al.*, 2016) and it is one of the chief mechanisms involved in the antagonism and bio-control efficacy of *Trichoderma* spp. (Sharon *et al.*, 2001). This process involves tropic growth of the bio-control agent towards the target organism, coiling and finally enzymatic dissolution of cell wall / cell membrane of the target organism where not only an antagonist, but also target pathogen responsible to govern the nature of antagonism. In a study, it was reported that some strains of *T. harzianum* have ability to parasitize on nematodes and their egg masses also where it coiled around the second stage juveniles of *Meloidogyne javanica* and penetrated them by forming haustoria like structures (Sahebani and Hadavi, 2008). *Trichoderma longibrachiatum* was also reported to manage *Heterodera avenae* (Zhang *et al.*, 2014a) and

Meloidogyne incognita (Zhang *et al.*, 2015) where it showed its bio-control potential.

Antibiosis

It is an antagonistic association between two microorganisms, in which one is adversely affected due to metabolites or antibiotics released by another. The chemical and analytical reports portray that 373 different secondary metabolites such as peptaibols, NRPs, volatile and non-volatile terpenes, pyrones and nitrogen containing compounds obtained from species of *Trichoderma* revealed their great potential to produce antibiotics and secondary metabolites (Mukherjee *et al.*, 2012a; Mukherjee *et al.*, 2012b; Crutcher *et al.*, 2013; Bae *et al.*, 2016; Contreras-Cornejo *et al.*, 2016). Gliotoxin and glyoviridin from *T. virens* (Wilhite *et al.*, 2001), viridin, alkylpyrones, isonitriles, polyketides, peptaibols, diketopiperazines and sesquiterpenes isolated from *Trichoderma* spp. (Singh *et al.*, 2001b; Reino *et al.*, 2008), trichodermin (Tijerino *et al.*, 2011; Shentu, *et al.*, 2013) and 6-Pentyl-2H-pyran-2-one (Garnica-Vergara *et al.*, 2015) are some of the examples which are found effective against target pathogen *in-vitro* and/or *in-situ*.

Competition and rhizosphere competence

The most abundant non-pathogenic plant-associated microbes are generally thought to protect the plant by rapid colonization and thereby exhausting the limited available substrates so that none are available for pathogens to grow. Being a living organism, competition is considered as a 'classical' mechanism of *Trichoderma* in biological control. The omnipresence of *Trichoderma* in agricultural and natural soils throughout the world proves that it must be an excellent competitor for space and nutritional resources. Tsahouridou and Thanassoulpoulos (2002) found the *T.*

koningii as rhizosphere competent when treated tomato seeds with conidial suspension of *T. koningii* were sown which resulted in reduced damping-off pathogen.

Sclerotia colonization and parasitization

The phytopathogens like *Rhizoctonia solani*, *Sclerotium rolfsii* and *Sclerotinia* spp. have tremendous ability to survive in soil for decades through hardy structures 'sclerotia' which are highly resistant to fungicides and microbial degradation and hence, play important role in disease cycle of the pathogen (Metcalf *et al.*, 2004, Abdulla *et al.*, 2008). The idea of sclerotia parasitization or killing them using microbial antagonist may be a key point for successful biological control. However, species of *Trichoderma* are also known for their ability to degrade cellulose and chitin (Balakrishnan *et al.*, 2012, Barakat *et al.*, 2013). In previous studies it was reported that the sclerotia of *Rhizoctonia solani* and *S. rolfsii* are colonized by *Trichoderma*, completely rotted and do not germinate if parasitized by *Trichoderma* (Rawat and Tewari, 2010; Sharma, 2017).

Indirect mechanism

Induction of host resistance

Besides the ability to attack directly or inhibit the growth of plant pathogens, some of the bio-control strains of *Trichoderma* sp. are known to strongly induce systemic resistance (ISR) and host plant defense against a variety of plant pathogens which is mediated by jasmonic acid (JA)/ethylene (Harman 2004; Pal and Gardener, 2006). In a study, the isolates of endophytic *Trichoderma* from tropical environments reported to induce resistance against *Phytophthora capsici* in hot pepper and delayed the disease onset (Bae *et al.*, 2011). Yedidia *et al.*, (2003) gave evidence of induction of a systemic response

by root application of *T. asperellum* which protect the cucumber plant against angular leaf spot. Induced resistance in tomato plant (cv. Sida cultivar) against bacterial leaf spot (*Xanthomonas campestris* pv. *vesicatoria*) with 69.32% reduction in disease was reported by Saksirirat *et al.*, (2009) after 14 days post inoculation of *T. harzianum* (T9). A protective effect of *Trichoderma* on plants was strongly inhibited when treated with an inhibitor of ethylene which confirmed the essential role of ethylene signal in ISR (Shoresh *et al.*, 2005). Moreover, there are some other reports which evidenced that species of *Trichoderma* are able to induce plant defense mechanisms (Shoresh, *et al.*, 2010; Contreras-Cornejo *et al.* 2011; Sharma, 2017). Research is initiated from last decade to characterize the determinants and biochemical pathways of induced resistance correlated with bio-control agents including *Trichoderma* and other non-pathogenic microbes.

Plant growth promotion

Bio-control agents, both fungal and bacterial, are reported to induce the growth of various crops. These responses may be due to the individual and /or cumulative effect of (i) suppression of deleterious root micro-flora, (ii) production of growth hormones or growth factors (Contreras-Cornejo *et al.* 2011, 2014a; Martínez-Medina *et al.*, 2014) and/or (iii) increased nutrient uptake through solubilization and sequestration of nutrients and/or enhanced root growth (Viterbo *et al.*, 2002). Species of *Trichoderma* are unique groups of rhizospheric microorganisms associated with certain beneficial effects to enhance plant growth and development. The research by several independent research groups have observed the increased growth of various crops with the use of antagonist *Trichoderma* spp. (Sharma *et al.*, 2012; Contreras-Cornejo *et al.*, 2013; Hermosa *et*

al., 2013; Samolski *et al.*, 2013; Qi and Zhao, 2013; Stewart and Hill, 2014). In a study conducted by Zhang *et al.*, (2014a) revealed that *Trichoderma longibrachiatum* was found fairly effective for promoting plant growth and nematode control in wheat. Increased shoot and roots length, weight, of pod weight and number of nodules per plant was reported in peanut with *T. harzianum* among the four *Trichoderma* as compare to control treatment (Kamaruzzaman *et al.*, 2016). The significant increase in yield (29% and 36%) of wheat (variety Raj 3765) was achieved by Sharma *et al.*, (2012) with the application of *Trichoderma harzianum* (Th3) in Jaipur and Kota respectively in the field demonstrations at farmer's field conducted during 2008-2011 Martínez-Medina *et al.*, (2014) reported that auxin production was induced and a decrease in cytokinins and abscisic acid content by *Trichoderma* isolates that promoted the plant growth. The biological activities of *Trichoderma* spp. in soil help in bioavailability of various nutrients and minerals either through chelation or solubilization, and thus make them available at the root surface of the plants (Menezes-Blackburn *et al.*, 2014) which results in increased plant growth.

Tolerance to abiotic stress

Abiotic stress (salt, drought etc.) play crucial role for crop cultivation as they limit in plant growth and subsequent yield in many areas. Some rhizospheric strains of *Trichoderma* are found fairly effective to induce plant growth under abiotic stresses (Bae *et al.*, 2009; Mastouri *et al.*, 2010; Shukla *et al.*, 2012). Due to higher production and activity of ROS-scavenging enzymes (superoxide dismutase, catalase and ascorbate peroxidase) induced in tomato plants colonized with *T. harzianum* T22 under water-deficit stress condition results in enhanced plant growth (Mastouri *et al.*, 2012). Under drought conditions, the cacao plants colonized with

endophytic fungus *Trichoderma hamatum* isolate DIS 219b exhibited delayed drought-induced changes in stomatal conductance and net photosynthesis (Bae *et al.*, 2009). In a glass house study where mustard plants were raised with *T. virens* (Isolate PB23) treated seeds and subjected to water stress. The plants were able to thrive well and showed delayed response under water-deficit stress (Sharma and Singh, 2014). *Trichoderma asperelloides* - T203 was found to ameliorate seed germination under saline stress conditions due to its acquired tolerance to salt stress which is supposed to link with ACCD activity (Brotman *et al.*, 2013). Inoculation of *A. thaliana* plants with *Trichoderma* resulted in significantly higher tolerance to salinity stress as compare to untreated plants which occur probably due to accumulation of antioxidant and stress hormone (Contreras-Cornejo *et al.*, 2014b).

Siderophores

Siderophores (Gr. "iron-bearers") are low molecular weight, ferric specific ligands which is to supply iron to the cell (Neilands, 1981) and iron is an essential element for various enzymatic reactions occur in living organisms although; naturally abundant ferric ion is almost insoluble in oxygenated environments and thus unavailable for microbial growth. Siderophores mediate the limited amount of iron in the rhizosphere, deprive pathogens of iron which produce less siderophores or different siderophores with lower binding coefficients and finally suppress their growth. Many reports have been published showing siderophore involvement in the suppression of plant pathogenic fungi (Vinale *et al.*, 2013). *T. virens* produces three types of hydroxamate siderophores: a monohydroxamate (*cis*- and *trans*-fusarinines), a dipeptide of *trans*-fusarinine (dimerum acid), and a trimer disepsipeptide (Lorito *et al.*, 1993).

Metabolism of germination stimulants

Propagules (sclerotia) of soil borne phytopathogens may germinate only in response to root exudates (germination stimulants) of host plants (Metcalf *et al.*, 2004) however, some of the non-pathogenic organisms may also prefer to utilize these compounds. Thus, a relatively new concept 'inhibition of germination stimulants' is supposed to be an indirect mechanism of bio-control and correlated with those non-pathogenic organisms where mycoparasitism / antibiosis is not established. Howell (2002) reported that control of pre-emergence damping off of cotton seedlings incited by *P. ultimum* and/or *Rhizopus oryzae* by *T. virens* (Strain G6, G6-5) or protoplast fusions of *T. virens* / *T. longibrachiatum* was due to metabolism of germination stimulants released by the cotton seed.

Application methods and factors affecting efficacy

The main problem in developing *Trichoderma* based bio-pesticides is that it represents a living system which should remain sufficiently viable for a period until it reaches to farmers (Vidhyasekaran and Muthamilan, 1995). Here, formulation helps to retain viability of bio-agent during preparation, storage and favours survival in the environment after application. Furthermore, it is quite essential to have an efficient, economic and ecologically viable delivery system or mode of application of bio-control agents in soil ecosystem and fairly good work has been done on this aspect under different conditions. Inoculation of seeds, corms, bulbs, tuber, etc. with antagonists prevents seed/corm decay and seedling blight. *T. harzianum*, *T. virens* and *T. viride* are effective seed protectants against *Pythium* spp. and *Rhizoctonia solani* (Jayaraj and Radhakrishana, 2003). According to Dutta

and Das, (2002), soil application of *Trichoderma* spp. at the time of seedling planting reduced the collar rot in tomato as compared to seed treatment. Several reports depict the effectiveness of bio-control as foliar spray against different plant pathogens however, success of these antagonists depends largely on their ability to colonize on leaf/sheath surface (Lo *et al.*, 1997).

For successful colonization of *Trichoderma* in rhizosphere / phyllosphere, the minimum inoculum is crucial and subsequent action to initiate the various mechanisms through which bio-agents inhibit, restrict even kill the pathogen directly or indirectly. Adequate amount of soil moisture, temperature near ambient and definite range of P^H (5.5-7.5) is needed for growth and multiplication of *Trichoderma* species to perform their bio-control activity in soil. Against *Rhizoctonia solani* in pea and rajma *Trichoderma harzianum* is more effective in acidic P^H > pH 6.5 (Singh *et al.*, 2006). Changes of 4°C or 5 per cent RH are associated with variability in disease suppression by *Trichoderma* that may range from 25-100 per cent (Hannusch and Boland, 1996).

Global and Indian scenario of bio-pesticides

Present share of bio-pesticides is only 5 % of the total crop protection market globally which was valued about \$3 billion worldwide (Olson, 2015). Majority of *Trichoderma* based bio-pesticides globally are dominated by *T. harzianum* and *T. viride* (Gardener and Fravel, 2002; Koul, 2011 and Table 1). In India, only 15 types of bio-pesticides with more than 970 products are registered and more than 63 Indian private companies including Pest Control (Pvt) Ltd; Multiplex Biotech Ltd., International Panacea, Biotech International Ltd; T. Stanes; etc. have their registered products as bio-pesticide.

Table.1 List of microbial pesticides developed from *Trichoderma* sp.

Bio-agent	Target pest	Action	Brand name	Producer
<i>Trichoderma harzianum</i>	Effective against variety of soil borne pathogens and wound pathogens	Mycoparasitic, Cell wall degrading enzymes (CWDE), ISR	Root Shield	BioWorks Inc, New York
			BioTrek 22g	Wilbur-Ellis Co., Fresno CA
			Supresivit	Borregaard, Denmark
			T-22G	Bioworks Inc, EU
			T-22HB	
			Trianum P	Koppert Biological Systems
			Binab	<i>BINAB Bio Innovation</i> AB, Sweden
			Trichodex	Makhteshim Chemical Works LTD, Israel
<i>Trichoderma viride</i>	Effective against soil borne pathogens causing root rot diseases	Mycoparasitic, Anibiosis, PGP	Ecosom TV	AgriLife, India
			Tricon	Green Max, India
			Trieco	Ecosense labs, India
			Bioderma/ Protector	Biotech International Ltd., New Delhi
			Ecofit	Eastman Kodak Company, New York
<i>Trichoderma</i> sp. OR <i>T. harzianum</i> + <i>T. viride</i>	Effective against <i>Armillaria</i> and, <i>Botryoshaeria</i> and other root pathogens	Mycoparasitic, Competition	Trichodry	Agrimm Technologies Ltd., New Zealand
			Vinevax	
			Trichoflow	
			Trichopel	
			Trichoject	
			Trichoseal	
<i>Trichoderma polysporum</i> + <i>T. harzianum</i>	Against wood decay and wounds of ornamental, shade, and forest trees		Binab T	Binab Bio-Inovation, EFTR AB, Sweden
<i>Gliocladium virens</i> GL-21	Damping off and root rot pathogens of nursery ornamental	Mycoparasitic, Antibiosis, Competition, CWDE	SoilGard	Certis USA

(Gardener and Fravel, 2002; Koul, 2011)

Microbial pesticides market in India dominated by sale of *Trichoderma viride*, *Pseudomonas fluorescens* and *Bacillus thuringiensis* and *Trichoderma* alone shared 52.5 % in the Indian bio-fungicides market in 2015 (Ken research Report, 2015). Funding for the research and production of bio pesticides and their promotion also is being led by the Department of Biotechnology (DBT) and Indian Council for Agricultural Research (ICAR) who support 22 and 31 bio-control production facilities respectively in India. For reducing complications in registration, some amendments are made in Insecticide Act of 1968 for speedy development of bio-pesticides. In India, the bio-control products are promoted by the Governments through subsidies, demonstration, training etc. and strict quality control protocols are followed to maintain its quality and efficacy. However, the major constraints for popularizing and marketing of bio-control agents (BCAs)/bio-pesticides In India can be directly correlated with factors such as lack of awareness about the use of bio-pesticides and related products, high cost and time consuming registration procedures, unreliable quality standards and product availability. Moreover, pesticide market is led by unorganized stakeholders and synthetic pesticides are easily availability than bio-pesticides.

Future prospects

Research findings with the use of molecular, biochemical, physiological and genetical study tools have explored a wide area of research on various mechanisms employed by *Trichoderma* in its biocontrol activities. Future research should be carried out efficiently on participatory mode with public-private sector approach for encouraging investment in bio-pesticide and bio-fertilizer enterprises. Additional funding for innovative

R&D for improvement of a particular microbial bio-pesticide by using unique sources (nanosilica, chitosan) will be needed. Development of consortium of two or more species or strains of *Trichoderma* in addition to liquid formulations is another area for increasing self-life and efficient delivery system at farmer's field.

References

- Abdullah, M.T., Ali, N.Y. and Suleman P. (2008). Biological control of *Sclerotinia sclerotiorum* (Lib.) de Bary with *Trichoderma harzianum* and *Bacillus amyloliquefaciens*. *Crop Prot.*, 27: 1354-1359.
- Atanasova, L., Le-Crom, S., Gruber, S. Coulpier, F., SeidlSeiboth, V., Kubicek, C.P. and Druzhinina, I. (2013). Comparative transcriptomics reveals different strategies of *Trichoderma* mycoparasitism. *BMC Genomics*, 14: 121.
- Bae, H., Roberts, D.P., Lim, H.S., Strem, M.D., Park, S.C., Ryu, C.M., Melnick, R.L., Bailey, B. (2011). Endophytic *Trichoderma* isolates from tropical environments delay disease onset and induce resistance against *Phytophthora capsici* in hot pepper using multiple mechanisms. *Mol. Plant Microbe Interact.*, 24(3): 36-51.
- Bae, H., Sicher, R.C., Kim, M.S., Kim, S.H., Strem, M.D., MeInice, R.L. and Bailey, B.A. (2009). The beneficial endophyte *Trichoderma hamatum* isolate DIS 219b promotes growth and delays the onset of the drought response in *Theobroma cacao*. *J. Exp. Bot.*, 60(32): 3279-3295.
- Bae, S.J., Mohanta, T.K., Chung, J.Y., Ryua, M., Park, G., Shim, S., Hong, S.B., Seo, H., Bae, D.W., Bae, I., Kima, J.J. and Bae, H. (2016). *Trichoderma*

- metabolites as biological control agents against *Phytophthora* pathogens. *Biol Control*, 92: 128–138.
- Balakrishnan Sowmya, Gomathi, D., Kalaiselvi, M., Ravikumar, G., Arulraj, C. and Uma, C. (2012). Production and purification of chitinase by *Streptomyces* sp. from soil. *J. Advanced Scientific Research.*, 3(3): 2-29.
- Barakat, F. M., Abada, K. A., Abou-Zeid, N. M. and El-Gammal, Y. H. E. (2013). Effect of volatile and non-volatile compounds of *Trichoderma* spp. on *Botrytis fabae* the causative agent of faba bean chocolate spot. *J. Agric. Res.*, 1(3): 42-50.
- Contreras-Cornejo, H. A., Macias-Rodriguez, L., del-Val, E., and Larsen, J. (2016). Ecological functions of *Trichoderma* spp. and their secondary metabolites in the rhizosphere: interactions with plants. *FEMS Microbiol. Ecol.* 92(4): fiw036.
- Contreras-Cornejo, H. A., Macías-Rodríguez, L. I., Alfaro-Cuevas, R., and López-Bucio, J. (2014a). *Trichoderma* spp. improve growth of *Arabidopsis* seedlings under salt stress through enhanced root development, osmolite production, and Na⁺ elimination through root exudates. *Mol. Plant-Microbe Interact.*, 27: 503-514.
- Contreras-Cornejo, H. A., Macías-Rodríguez, L. I., Herrera-Estrella, A., and López-Bucio, J. (2014b). The 4-phosphopantetheinyl transferase of *Trichoderma virens* plays a role in plant protection against *Botrytis cinerea* through volatile organic compound emission. *Plant Soil.*, 379: 261-274.
- Contreras-Cornejo, H. A., Ortiz-Castro, R., and López-Bucio, J. (2013). “Promotion of plant growth and the induction of systemic defence by *Trichoderma*: physiology, genetics and gene expression,” In *Trichoderma: Biology and Applications*, eds P. K. Mukherjee, B. A. Horwitz, U. S. Singh, M. Mukherjee, and M. Schmoll (Wallingford: CABI), 173–194.
- Contreras-Cornejo HA, Macías-Rodríguez L, Beltrán-Peña E, Alfredo Herrera-Estrella A, López-Bucio J (2011). *Trichoderma*-induced plant immunity likely involves both hormonal- and camalexin-dependent mechanisms in *Arabidopsis thaliana* and confers resistance against necrotrophic fungus *Botrytis cinerea*. *Plant Signal Behav.*, 6(10): 1554-1563.
- Crutcher, F.K., Parich, A., Schuhmacher, R., Mukherjee, P.K., Zeilinger, S. and Kenerley, C.M. (2013). A putative terpene cyclase, *vir4*, is responsible for the biosynthesis of volatile terpene compounds in the biocontrol fungus *Trichoderma virens*. *Fungal Genet. Biol.*, 56: 67-77.
- Druzhinina IS, Seidl-Seiboth V, Herrera-Estrella A, Horwitz BA, Kenerley CM, Monte E, Mukherjee PK, Zeilinger S, Grigoriev IV, Kubicek CP (2011). *Trichoderma*: the genomics of opportunistic success. *Nat Rev Microbiol.* 9(10): 749-59.
- Dutta, P. and Das, B.C. (2002). Management of collar rot of tomato by *Trichoderma* spp. and chemicals. *Indian Phytopathol.*, 55 (2): 235-237.
- Gardener, M.B.B., and Fravel, D.R. (2002). Biological control of plant pathogens: Research, commercialization, and application in the USA. Online. *Plant Health Progress*, doi: 10.1094/PHP-2002-0510-01-RV.
- Garnica-Vergara, A., Barrera-Ortiz, S., Muñoz-Parra, E., Raya-González, J., Méndez-Bravo, A., Macías-Rodríguez, L., Francisco Ruiz-Herrera

- L. and López-Bucio, J. (2016). The volatile 6-pentyl-2H-pyran-2-one from *Trichoderma atroviride* regulates *Arabidopsis thaliana* root morphogenesis via auxin signaling and Ethylene Insensitive 2 functioning. *New Phytol.*, 209: 1496–1512.
- Hannusch, D.J. and Boland, G.J. (1996). Influence of air temperature and relative humidity on biological control of white mold of bean (*Sclerotinia sclerotiorum*). *Phytopathology*, 86 (2): 156-162.
- Harman, G.E., Howell, C.R., Viterbo, A., Chet, I. and Lorito, M. (2004). *Trichoderma* species-Opportunistic, avirulent plant symbionts. *Nature Rev.*, 2: 43-56.
- Hermosa, R., Rubio, M.B., Cardoza, R.E., Nicolás, C., Monte, E., and Gutiérrez, S. (2013). The contribution of *Trichoderma* to balancing the costs of plant growth and defense. *International Microbiology*, 16: 69–80.
- Howell C.R., (2002). Cotton seedling preemergence damping off incited by *Rhizopus oryzae* and *Pythium* spp. and its biological control with *Trichoderma* spp. *Phytopathology*. 92: 177-180
- Jayaraj, J. and Radhakrishnan, N.V. (2003). Development of UV-induced carbendazim-resistant mutants of *Trichoderma harzianum* for integrated control of damping-off disease of cotton caused by *Rhizoctonia solani*. *Zeitschrift für Pflanzenkrankheiten und Pflanzenschutz*, 110 (5): 449-460.
- Kamaruzzaman, M., Rahman, M.M. and Ahmad, M.U. (2016). Efficacy of four selective *Trichoderma* isolates as plant growth promoters in two peanut varieties. *International Journal of Biological Research*, 4(2): 152-156.
- Ken research Report (2015). Courtesy: India Biopesticide Market Outlook: Growth, trends and Market forecast to 2020.
- Koul A. (2011). Microbial biopesticides: opportunities and challenges. CAB Reviews: Perspectives in Agriculture, Veterinary Science, Nutrition and Natural Resources, 2011, 6(56): 1-26.
- Lo, C.T., Nelson, E.B. and Harman, G.E. (1997). Improved biocontrol efficacy of *Trichoderma harzianum* 1295-22 for foliar phases of turf diseases by use of spray applications. *Plant Dis.*, 81 (10): 1132-1138.
- Martínez-Medina, A, Maria Del Mar Alguacil, Jose A. Pascual, Saskia, C.M. and Van Wees (2014). Phytohormone profiles induced by *Trichoderma* isolates correspond with their biocontrol and plant growth-promoting activity on melon plants. *J Chem Ecol*, 40:804–815.
- Mastouri, F., Bjorkman, T., and Harman, G. E. (2012). *Trichoderma harzianum* enhances antioxidant defense of tomato seedlings and resistance to water deficit. *Mol. Plant Microbe Interact.*, 25, 1264-1271.
- Mastouri, F., Bjorkman, T., and Harman, G. E. (2010). Seed treatment with *Trichoderma harzianum* alleviates biotic, abiotic, and physiological stresses in germinating seeds and seedlings. *Phytopathology*, 100: 1213–1221.
- Menezes-Blackburn, D., Jorquera, M.A., Gianfreda, L., Greiner, R., de la Luz Mora, M. (2014). A novel phosphorus biofertilization strategy using cattle manure treated with phytase-nanoclay complexes. *Biol Fertil Soils*, 50: 583-592.
- Metcalf, D.A., Dennis, J.J.C. and Wilson, C.R. (2004). Effect of inoculum

- density of *Sclerotium cepivorum* on the ability of *Trichoderma koningii* to suppress white rot of onion. *Plant Disease*, 88:287-291.
- Mukherjee, P.K., Horwitz, B.A. and Kenerley, C.M. (2012a). Secondary metabolism in *Trichoderma* – a genomic perspective. *Microbiology*, 158: 35-45.
- Mukherjee PK, Buensanteai, N., Moran-Diez, ME. (2012a). Functional analysis of nonribosomal peptide synthetases (NRPSs) in *Trichoderma virens* reveals a polyketide synthase (PKS)/NRPS hybrid enzyme involved in the induced systemic resistance response in maize. *Microbiology*, 158: 155-165.
- Neilands, J. B. (1981). Microbial iron compounds. *Annu. Rev. Biochem.*, 50: 715-731.
- Olson, S. (2015). An analysis of the biopesticide market now and where is going. *Outlooks Pest Manag.*, 26: 203-206.
- Pal, K.K. and Gardener M.B. (2006). Biological Control of Plant Pathogens *The Plant Health Instructor*. DOI: 10.1094/PHI-A-2006-1117-02.
- Qi W. Z., Zhao L. (2013). Study of the siderophore-producing *Trichoderma asperellum* Q1 on cucumber growth promotion under salt stress. *J. Basic Microbiol.* 53 355–364.
- Rawat, R. and Tewari, L. (2010). Transmission electron microscopic study of the cytological changes in *Sclerotium rolfii* parasitized by a biocontrol fungus *Trichoderma* species. *Mycology: An International J. Fungal Biology.*, 1(4): 237-241.
- Reino, J.L., Guerrero, R.F., Hernández-Galán, R. and Collado, I.G. (2008). Secondary metabolites from species of the biocontrol agent *Trichoderma*. *Phytochem. Rev.*, 7: 89-123.
- Sahebani, N. and Hadavi, N. (2008). Biological control of the root-knot nematode *Meloidogyne javanica* by *Trichoderma harzianum*. *Soil Biology and Biochemistry*, 40, 2016-2020.
- Saksirirat, W., Chareerak, P. and Bunyatrachata, W. (2009). Induced systemic resistance of biocontrol fungus, *Trichoderma* spp. against bacterial and gray leaf spot in tomatoes. *Asian J. Food Agro-Industry*, 2: S99-S104.
- Samolski, I., Rincón, A., Pinzón, L.M. *et al.*, (2012). The *qid74* gene from *Trichoderma harzianum* has a role in root architecture and plant biofertilization. *Microbiology*, 158(1): 29-38.
- Sharma, K.K. (2017). Induction of systemic resistance (ISR) against sheath blight of rice caused by *Rhizoctonia solani* Kuhn using biological seed treatment with *Trichoderma*. *Journal of Applied and Natural Science*, 9(3): 1861-1865.
- Sharma, K.K. (2017). Qualitative enzyme assay and sclerotia parasitization by fungal antagonist *Trichoderma*. *The Bioscan* (Supplement on Plant Pathology), 11(4): 2867-2872.
- Sharma, K.K. and Singh U.S. (2014). Induction of water stress tolerance of mustard plants using *Trichoderma* as seed treatment. *Journal of Applied and Natural Science*, 6(2): 436-441.
- Sharma, P., Patel, A.N, Saini, M.K. and Swati Deep (2012). Field Demonstration of *Trichoderma harzianum* as a Plant Growth Promoter in Wheat (*Triticum aestivum* L). *Journal of Agricultural Science*, 4(8): 65-73.
- Sharon, E., Bar-Eyal, M., Chet, I., Herrera-Estrella, A., Kleifeld, O. and Spiegel, Y. (2001). Biological control of the root knot nematode *Meloidogyne javanica* by *Trichoderma harzianum*. *Phytopathology*, 91: 687-693.

- Shentu, X.P., Liu, W.P., Zhan, X.H., Yu, X.P. and Zhang, C.X. (2013). The elicitation effect of pathogenic fungi on trichodermin production by *Trichoderma brevicompactum*. *Sci. World J.*, DOI: 10.1155/2013/607102
- Shoresh, M., Harman, G. E. and Mastouri, F. (2010). Induced systemic resistance and plant responses to fungal biocontrol agents. *Annu Rev Phytopathol.*, 48: 21-43.
- Shoresh, M., Yedidia, I. and Chet, I. (2005). Involvement of jasmonic acid/ethylene signaling pathway in the systemic resistance induced in cucumber by *Trichoderma asperellum* T203. *Phytopathology* 95: 76-84.
- Shukla, N., Awasthi, R.P. and Rawat, L. (2012). Biochemical and physiological responses of rice (*Oryza sativa* L.) as influenced by *Trichoderma harzianum* under drought stress. *Plant Physiol. Biochem.*, 54: 78-88.
- Singh, U.S., Mishra, D.S., Rohilla, R., Singh, A. and Vishwanath. (2001b). Induced resistance: present status and future prospects as disease management strategy. In: Biopesticides and Pest Management (Opender Kaul, G.S. Dhaliwal, S.S. Marwaha, and J. Arora, eds.), Campus Book International, New Delhi. Degradation of fungal cell walls by lytic enzymes of *Trichoderma harzianum*.
- Singh, U.S., Zaidi, N.W., Joshi, D., Varshney, S. and Khan, T. (2006). Current status of *Trichoderma* as a biocontrol agent. In: Ramanujam B, Rabindra RJ (eds) Current status of biological control of plant diseases using antagonistic organisms in India, Project Directorate of Biological Control, Bangalore.
- Stewart, A. and Hill, R. (2014). Applications of *Trichoderma* in Plant Growth Promotion. In: Biotechnology and Biology of *Trichoderma*, Publisher(s): Elsevier BV; Elsevier, Pp: 415-428.
- Tijerino, A., Cardoza, R.E., Moraga, J., Malmierca, M.G. and Vicente, F. *et al.*, (2011). Overexpression of the trichodiene synthase gene *tri5* increases trichodermin production and antimicrobial activity in *Trichoderma brevicompactum*. *Fungal Genet. Biol.*, 48: 285-296.
- Tsahouridou, P.C. and Thanassoulopoulos, C.C. (2002). Proliferation of *Trichoderma koningii* in the tomato rhizosphere and the suppression of damping-off by *Sclerotium rolfsii*. *Soil Biol Biochem.*, 34(6): 767-776.
- Vidhyasekaran, P. and Muthamilan, M. (1995). Development of formulations of *Pseudomonas fluorescens* for the control of chickpea wilt. *Plant Dis.*, 79: 782-786.
- Vinale, F., Nigro, M., Sivasithamparam, K., Flematti, G., Ghisalberti, E.L., Ruocco, M., Varlese, R., Marra, R., Lanzuise, S., Eid, A., Woo, S.L. and Lorito, M. (2013). Harzianic acid: a novel siderophore from *Trichoderma harzianum*. *FEMS Microbiol. Lett.*, 347(2): 123-129.
- Vinale, F., Sivasithamparam, K., Ghisalberti, E.L., Woo, S.L., Nigro, M., Marra, R., Lombardi, N., Pascale, A., Ruocco, M., Lanzuise, S., Manganiello, G. and Lorito, M. (2014). *Trichoderma* secondary metabolites active on plants and fungal pathogens. *Mycol. J.*, 8: 127-139.
- Viterbo, A., Montero, M., Ramot, O., Friesem, D., Monte, E., Liobell, A. and Chet, I. (2002). Expression regulation of the endochitinase *chit 39* from *Trichoderma asperellum* (*T. harzianum* T-203). *Curr. Genet.*, 42: 114-122.
- Wilhite SE, Lumsden RD and Straney DC. (2001). Peptide synthetase gene in

- Trichoderma virens*. *Applied and Environmental Microbiology*, 67: 5055–5062.
- Yedidia, I., Shoresh, M., Kerem, Z., Benhamou, N., Kapulnik, Y. and Chet, I. (2003). Concomitant induction of systemic resistance to *Pseudomonas syringae* pv. Lachrymans in cucumber by *Trichoderma asperellum* (T-203) and accumulation of phytoalexins. *Applied and Environmental Microbiology*. 69(12): 7343-7353.
- Zhang, S., Gan, Y. and Xu, B. (2016). Application of plant-growth-promoting fungi *Trichoderma longibrachiatum* T6 enhances tolerance of wheat to salt stress through improvement of antioxidative defense system and gene expression. *Front Plant Sci.*, 2016(7): 1-11.
- Zhang S. W., Gan Y. T., Xu B. L. (2014a). Efficacy of *Trichoderma longibrachiatum* in the control of *Heterodera avenae*. *BioControl* 59 319–331.
- Zhang S. W., Gan Y. T., Xu B. L. (2015). Biocontrol potential of a native species of *Trichoderma longibrachiatum* against *Meloidogyne incognita*. *Appl. Soil Ecol.* 94 21–29.

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