

Original Research Article

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Diversity Analysis and Hybridity Test of Drought Responsive Wheat Cultivars and their F₁'s on the Basis of SDS-PAGE Profiles

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ABSTRACT

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The genetic polymorphism revealed by biochemical markers (protein and isozymes) in plant population have been extensively used to, assess the genetic variability, confirm hybridity of F₁s, assure purity of cultivar and establish relationship between species. Electrophoretic protein analysis holds potential utility in breeding programmes of wheat through helping in the unequivocal characterization of varieties used. Sixteen wheat genotypes along with their 32 F₁'s were characterized by using protein profile polymorphism in seeds through SDS-PAGE. The total number of bands varied from 9-17 with arrange of 3000 kd to 20,100 kd. The similarity coefficient ranged from 0.985-0.632 among parental lines. The dendrogram constructed on the basis of similarity coefficient clearly divided the genotypes into two major clusters. In most of the crosses the protein profile was more likely to the maternal parent with few bands from the male parent. Presence and/or absence of unique bands were also recorded in crosses which might be because of cis/trans regulation for gene expression and protein synthesis.

Introduction

Drought is generally accepted to be the most widespread abiotic stress experienced by crop plants, and is becoming an increasingly severe problem in many regions of the world. Each year, drought strikes more than half of the area sown to wheat in the developing world. Thirty seven percent of the area of developing countries consists of semi-arid environment in which available moisture constitutes a primary constraint on wheat production. If prediction is right, an even large expanse of land will become parched every year owing to global

warming, urbanization and deforestation. As conditions worsen, farmers will need varieties that tolerate drought and are more responsive to farming practices with better water use efficiency. The effects of drought on crop plants are complex and variable and are greatly accentuated by a number of interacting factors. The onset of drought in general has been observed to reduce germination, emergence, hypocotyl length, water use efficiency and the mobilization of dry matter reserve even at early growth stages (Richards and Thurling, 1978). Wheat is one of the major staple food crops throughout the world.

Because of modern breeding, it has been realized that the genetic diversity in wheat has increasingly narrowed. Narrow genetic base hamper the pace of crop improvement programme and ultimately gain. It is difficult to breed for adaptation to abiotic and biotic stresses with the population having narrow genetic base (Ahmed *et al.*, 2008). Therefore, it becomes necessary to assess the genetic diversity in wheat in order to enrich the gene pool and broaden the genetic base for future breeding programme. Morphological traits are being used to assess the genetic diversity but they are highly influenced by environmental fluctuations. Use of biochemical marker for genetic diversity studies have received much attention because of cheap, easy in set up and abundance in nature (Masood *et al.*, 2004).

Protein markers could be very successfully used to verify the identity of a variety. If a suitable protocol is standardized, it can be used with equal reproducibility at any time and any location. So such markers can be used to identify parents and to test hybridity of F₁s (Akhare *et al.*, 2007). Among the biochemical markers the SDS-PAGE is widely used due to its simplicity and effectiveness for describing the genetic structure of crop germplasm. Identification of different germplasm of is essential when diverse accessions of crop germplasm are to be characterized, newly developed cultivars are to be registered and purity of a variety is to be determined.

Use of biochemical markers for maintaining the genetic purity of line and hybridity of F₁s has also been reported earlier (Bhatt and Khanna, 2006; Pallavi *et al.*, 2010; Geetha and Balamurugan, 2011; Sharma *et al.*, 2015). Therefore, the present investigation was undertaken to study utility of biochemical marker as an alternative method for assessment of diversity and distinguishing the F₁s and parents.

Materials and Methods

SDS-PAGE discontinuous system was used to observe the protein banding pattern of the parents and their respective F₁'s. The procedure followed was as per the protocol of Laemmli (1970) with some modifications suggested by Lawrence and Shepherd (1980).

Experimental material

The experimental material used for the study consisted of thirteen drought tolerant, three drought susceptible wheat genotypes and their thirty two F₁'s produced by crossing of these diverse genotypes.

Experimental method

Extraction of total seed protein

The total seed storage protein extraction was done according to the method described by Singh *et al.*, (1991). One grain (about 30 mg flour) was crushed to fine powdered form and then added 400 µl extraction buffer (pH 6.8) containing 2 per cent β-mercaptoethanol and vortexed briefly for 1 to 2 minutes. The tubes were incubated at 60⁰C for 30 minutes and cooled to room temperature. 25µl dye was added to each tube and the contents were centrifuged for 10 minutes at 10,000 rpm. The supernatant was used for loading the sample in the well of the gel for electrophoresis.

Preparation of gels

The 8.0% stacking and 10% separating gels was prepared from the stock solutions.

APS (10 %) was added to separating gel mix just before pouring the gel solution between the plates. 20 µl sample was loaded in each well and the electrodes were connected with the power supply. Initially the current was set at 100 v and 26 mA current for 60 minutes.

Later on it was increased to 40 mA with constant voltage at 120 v.

Staining, destaining and documentation of gel

Gel was transferred to the staining solution tray after completion of electrophoresis for overnight. Next day 3% NaCl solution was used for destaining as described by Sreeramulu and Singh (1994). The photographs of gels were taken on Alpha Imager gel documentation system.

Analysis of gels

The Rf value was calculated with the help of Alpha Imager, a software for gel documentation.

$Rf = \text{Path travelled by the bands} / \text{Path travelled by the dye front}$

The protein profiles were also score on the basis of presence and absence of bands. The 0-1 binary data was analyzed by the NT-SYS 2.0 software for diversity analysis.

Results and Discussion

Seed storage proteins are encoded by many multigeneic loci and the production of a single locus comprised of several electrophoretically separable bands. The procedure has been standardized according to crop for different protein fractions. In present study thirteen drought tolerant and three drought susceptible genotypes of wheat and their thirty two F₁s were used for diversity analysis and characterization by total soluble seed proteins separated on SDS-PAGE. The protein banding profile and its zymogram is presented in figure 1, 2 and 3. Rf values were calculated for each band. The details of banding profile on the basis of Rf value was given in Table1.

The total number of bands varied from 9-17 with Rf value from 0.11 to 0.76. Most of the bands present in maternal parent were found in F₁ while there were few bands those were common in paternal parent and F₁s. There was presence of few unique bands in F₁s those were not found in the either of the parents. Three bands of Rf value 0.10, 0.14 and 0.16 were found only in crosses and absent in either of the parents. A reverse situation was also observed where bands present in both the parents were not found in F₁s. Bands of Rf value 0.10, 0.15, 0.28, 0.31, 0.40 and 0.76 were found in the parents but not found in the respective crosses.

Diversity analysis

Electrophoresis of protein is a powerful tool for identification of genetic diversity and the SDS-PAGE is predominantly considered as a consistent technology because seed storage protein are highly independent of environmental fluctuations (Javid *et al.*, 2004, Iqbal *et al.*, 2005). Diversity is pre-requisite for planning a breeding programme. The banding profile (Fig 1) was scored on absence and presence (0-1) of bands and the data was analyzed by NT-SYS software for diversity analysis. The diversity in seed storage protein has also been reported by Khan *et al* (2002), Kakai and Kahrizi (2011) in wheat. The parental lines in present investigation showed wide range of variability. The similarity coefficient ranged from 0.985-0.632 (Fig 4). The minimum similarity coefficient was found between PBW 175 and NIAW 34 (0.632) Followed by VL 804 and NIAW 34 (0.647). The maximum similarity was found between VL 804 and VL 802 (0.985) and VL 802 and UP 2572 (0.985). Low value of similarity is an indicative of high level of diversity. The high level of diversity is because of their diverse origin. Highly diverse genotypes can be used as potential donors for improvement programme and can also be used for the

development of diverse gene pools for exploitation of heterosis. Jangid *et al.*, (2017) used SDS-PAGE for diversity studies among released varieties and crosses developed through diallel mating design in mustard.

The dendrogram constructed on the basis of similarity coefficient clearly divided the genotypes into two major clusters (Fig 6). Both the clusters comprised of 8 genotypes each at 0.71 similarity coefficient. Group I can be subdivided into 2 sub groups. Subgroup I a with genotypes VL-80.2, 804, UP2572 whereas subgroup I b comprised of PBW 175, PBW 65, PBW 373, UP 2425 and UP 2338. The genotypes in subgroup I a were suitable for cultivation in the hills of Uttarakhand. PBW175, PBW 65, PBW 373, UP 2425, UP 2338 are recommended for plains areas. All the Genotypes having gene for drought/heat tolerance belongs to the group II. Group II is further subdivided into 3 subgroups. Group II a comprised of 3 genotypes viz; WH 730, Job666 and NI 5439. NP-846, NIAW34 and HI 385 belong to subgroup II b and PBN-51 and Halna constituted II c. Such clustering helps in grouping the genotype in diverse gene pools for future breeding programmes. The application of protein profiling in germplasm characterization and their use as genetic markers have been widely and effectively used to determine the diversity, taxonomy and evolutionary aspects of several crops (Sharma *et al.*, 2015; Gafoor *et al.*, 2002; Ahmad *et al.*, 2010; Ullah *et al.*, 2016 and Singh *et al.*, 2017)

Hybridity of F₁s and identification of cultivars

The parents and their F₁'s were compared for their protein banding pattern to observe the hybrid nature of F₁'s (Akhare *et al.*, 2007 and Pallavi *et al.*, 2010). The total number of bands varied from 9-17 with a range of 3000 kd to 20,100 kd. The results showed that the

presence of bands from both the parents was found in the F₁'s that showed that the F₁'s were the result of the cross between drought tolerant and drought susceptible parents (Fig 5). A combined dendrogram of parents and F₁s (Fig 7) showed that the protein profile of almost all the F₁s are intermediate to their parental profiles but made cluster with the female parent. The reason being most of the bands were contributed by the female parent. In few crosses the profile of F₁s was identical to their respective female parent. Such a condition limits the application of SDS-PAGE for identification and characterization. Findings of Jaiswal and Agrawal 1990 also reported similar findings that banding pattern of protein extracted from seeds could not present a clear spectrum. In some crosses new bands were observed that were not present in either of the parents, whereas there were cases where, the bands found in both the parents were absent in the crosses. Such unique cases were an indication towards the new nuclear-cytoplasmic interaction for expression of proteins.

Protein profiling of selected genotypes have been found effective to differentiate the different genotypes. The tolerant and susceptible parents could be differentiated on the basis of bands profile. The bands of Rf value 0.26, 0.31, 0.53, 0.63 and 0.72 were present in tolerant parents, while bands of Rf value 0.09, 0.17 and 0.44 were present in drought susceptible genotypes. So these protein bands having specific Rf value may be useful for separating the two types. Identification of wheat genotypes including ILC-195, CM-2000 and CM-98/99 has also been reported by protein markers (Zeb *et al.*, 2006). The utilization of wheat endosperm protein through SDS-PAGE was reported as a valuable tool for assessment of genetic diversity and cultivar identification that help in wheat breeding programme (Khan and Ali, 2014).

Table.1 Protein profile of wheat parental lines and F₁s by SDS PAGE

S No	Name of cross	Range of bands in F ₁ s	Bands in F ₁ s	Common bands in F ₁ s and female parent	Bands in female Parent	Common bands in F ₁ s and male	Band in male parent	Common in all	Unique bands in	Absence of bands in F ₁ s
1	VL804/UP 2425	0.13 to 0.72	0.13, 0.16, 0.20, 0.22, 0.26, 0.31, 0.37, 0.40, 0.50, 0.53, 0.63 and 0.72	0.13, 0.20, 0.22, 0.26, 0.31, 0.37, 0.40, 0.50, 0.53, 0.63 and 0.72	0.13, 0.20, 0.22, 0.26, 0.31, 0.37, 0.40, 0.50, 0.53, 0.63 and 0.72	0.13, 0.22, 0.37, 0.50	0.11, 0.13, 0.18, 0.19, 0.22, 0.37, 0.39, 0.47, 0.50, 0.54, 0.62 and 0.76	0.13, 0.22, 0.37 and 0.50	0.16	
2	VL804/PBW 373	0.13 to 0.72	0.13, 0.16, 0.20, 0.22, 0.26, 0.31, 0.37, 0.40, 0.50, 0.53, 0.63, 0.72	0.13, 0.20, 0.26, 0.31, 0.40, 0.53, 0.63 and 0.72	0.13, 0.20, 0.22, 0.26, 0.31, 0.37, 0.40, 0.50, 0.53, 0.63 and 0.72	0.13, 0.22, 0.37, 0.50	0.09, 0.13, 0.18, 0.22, 0.37, 0.39, 0.47, 0.50, 0.54, 0.62 and 0.76.	0.13, 0.22, 0.37 and 0.50	0.16	
3	VL 802/UP 2338	0.13 to 0.72	0.13, 0.16, 0.20, 0.22, 0.26, 0.31, 0.37, 0.40, 0.50, 0.53, 0.63 and 0.72	0.13, 0.20, 0.26, 0.31, 0.37, 0.40, 0.53, 0.63 and 0.72.	0.13, 0.20, 0.22, 0.26, 0.31, 0.37, 0.40, 0.50, 0.53, 0.63 and 0.72	0.22, 0.50	0.11, 0.18, 0.19, 0.22, 0.28, 0.30, 0.39, 0.47, 0.50, 0.54 and 0.62.	0.22 and 0.50	0.16	
4	VL 802/PBW 373	0.13 to 0.76.	0.13, 0.20, 0.22, 0.26, 0.31, 0.37, 0.40, 0.53, 0.50, 0.63, 0.72, 0.76	0.13, 0.20, 0.22, 0.26, 0.31, 0.37, 0.40, 0.50, 0.53, 0.63 and 0.72	0.13, 0.20, 0.22, 0.26, 0.31, 0.37, 0.40, 0.50, 0.53, 0.63 and 0.72.	0.76	0.09, 0.13, 0.18, 0.22, 0.37, 0.39, 0.47, 0.50, 0.54, 0.62 and 0.76	0.13, 0.22, 0.37 and 0.50		
5	VL 802/UP 2425	0.13 to 0.76.	0.13, 0.20, 0.22, 0.26, 0.31, 0.37, 0.40, 0.50, 0.53, 0.63, 0.72, 0.76	0.13, 0.20, 0.26, 0.31, 0.40, 0.53, 0.63 and 0.72	0.13, 0.20, 0.22, 0.26, 0.31, 0.37, 0.40, 0.50, 0.53, 0.63 and 0.72	0.76	0.11, 0.13, 0.18, 0.19, 0.22, 0.37, 0.39, 0.47, 0.50, 0.54, 0.62 and 0.76	0.13, 0.22, 0.37 and 0.50		
6	UP 2572/PBW 373	0.13 to 0.72	0.13, 0.20, 0.26, 0.31, 0.37, 0.40, 0.50, 0.53, 0.54, 0.63, 0.72	0.13, 0.20, 0.26, 0.31, 0.40, 0.53, 0.63 and 0.72	0.13, 0.20, 0.26, 0.31, 0.37, 0.40, 0.50, 0.53, 0.63 and 0.72	0.54	0.09, 0.13, 0.18, 0.22, 0.37, 0.39, 0.47, 0.50, 0.54, 0.62 and 0.76	0.13, 0.37 and 0.50		
7	UP 2572/UP 2425	0.13 to 0.72	0.13, 0.16, 0.20, 0.26, 0.37, 0.40, 0.50, 0.53, 0.54, 0.63 and 0.72.	0.20, 0.26, 0.40, 0.53, 0.63 and 0.72.	0.13, 0.20, 0.26, 0.31, 0.37, 0.40, 0.50, 0.53, 0.63 and 0.72	0.54	0.11, 0.13, 0.18, 0.19, 0.22, 0.37, 0.39, 0.47, 0.50, 0.54, 0.62 and 0.76.	0.13, 0.37 and 0.50	0.16	
8	UP 2572/UP 2338	0.13 to 0.72	0.13, 0.22, 0.20, 0.26, 0.37, 0.40, 0.50, 0.53, 0.54, 0.63 and 0.72	0.13, 0.20, 0.26, 0.37, 0.40, 0.50, 0.53, 0.63 and 0.72	0.13, 0.20, 0.26, 0.31, 0.37, 0.40, 0.50, 0.53, 0.63 and 0.72	0.22 and 0.54	0.11, 0.18, 0.19, 0.22, 0.28, 0.30, 0.31, 0.39, 0.47, 0.50, 0.54 and 0.62.	0.50	-	0.31
9	PBW 65/PBW 373	0.11 to 0.76	0.11, 0.19, 0.22, 0.30, 0.37, 0.40, 0.47, 0.50, 0.62 and 0.76	0.11, 0.19, 0.30 and 0.40	0.11, 0.19, 0.22, 0.30, 0.37, 0.40, 0.47, 0.50, 0.53, 0.62 and 0.76.		0.09, 0.13, 0.18, 0.22, 0.37, 0.39, 0.47, 0.50, 0.54, 0.62 and 0.76	0.22, 0.37, 0.47, 0.50, 0.62 and 0.76		
10	PBW 65/UP 2425	0.11 to 0.76	0.11, 0.18, 0.19, 0.22, 0.30, 0.37, 0.30, 0.47, 0.50, 0.62 and 0.76	0.30 and 0.40	0.11, 0.19, 0.22, 0.30, 0.37, 0.40, 0.47, 0.50, 0.53, 0.62 and 0.76.	0.18	0.11, 0.13, 0.18, 0.19, 0.22, 0.37, 0.39, 0.47, 0.50, 0.54, 0.62 and 0.76	0.11, 0.19, 0.22, 0.37, 0.47, 0.50, 0.62 and 0.76		
11	PBW 65/UP 2338	0.11 to 0.76	0.11, 0.19, 0.22, 0.30, 0.37, 0.40, 0.47, 0.50,	0.37 and 0.76	0.11, 0.19, 0.22, 0.30, 0.37, 0.40, 0.47, 0.50,		0.11, 0.18, 0.19, 0.22, 0.28, 0.30, 0.39,	0.11, 0.19, 0.22, 0.30, 0.40, 0.47,		

			0.62, 0.76		0.53, 0.62 and 0.76		0.47, 0.50, 0.54 and 0.62	0.50 and 0.62		
12	PBW 175/UP 2338	0.11 to 0.62	0.11, 0.18, 0.19, 0.22, 0.28, 0.30, 0.37, 0.47 and 0.50	0.37	0.11, 0.18, 0.19, 0.22, 0.28, 0.30, 0.37, 0.40, 0.47, 0.50, 0.54, 0.62 and 0.76	0.30	0.11, 0.18, 0.19, 0.22, 0.28, 0.30, 0.39, 0.40, 0.47, 0.50, 0.54 and 0.62.	0.11, 0.18, 0.19, 0.22, 0.28, 0.47 and 0.50		0.40
13	PBW 175/UP 2425	0.11 to 0.62	0.11, 0.18, 0.19, 0.28, 0.37, 0.47, 0.50, 0.54 and 0.62	0.28	0.11, 0.18, 0.19, 0.22, 0.28, 0.37, 0.40, 0.47, 0.50, .54, 0.62 and 0.76		0.11, 0.13, 0.18, 0.19, 0.22, 0.37, 0.39, 0.40, 0.47, 0.50, 0.54, 0.62 and 0.76.	0.11, 0.18, 0.19, 0.37, 0.47, 0.50, 0.54 and 0.62		0.40 and 0.76
14	PBW 175/PBW 373	0.11 to 0.76	0.11, 0.18, 0.22, 0.28, 0.37, 0.40, 0.47, 0.50, 0.54, 0.62 and 0.76	0.11, 0.19, 0.28, and 0.40	0.11, 0.18, 0.19, 0.22, 0.28, 0.37, 0.40, 0.47, 0.50, 0.54, 0.62 and 0.76		0.09, 0.13, 0.18, 0.22, 0.37, 0.39, 0.47, 0.50, 0.54, 0.62 and 0.76	0.18, 0.22, 0.37, 0.47, 0.50, 0.54, 0.62 and 0.76		0.40
15	WH 730/UP 2425	0.13 to 0.76	0.13, 0.19, 0.23, 0.28, 0.33, 0.37, 0.39, 0.41, 0.46, 0.49, 0.54, 0.60 0.63, 0.72 and 0.76	0.13, 0.23, 0.28, 0.33, 0.41, 0.46, 0.49, 0.60, 0.63 and 0.72	0.13, 0.19, 0.23, 0.28, 0.33, 0.37, 0.39, 0.41, 0.46, 0.49, 0.54, 0.60, 0.63, 0.72 and 0.76		0.11, 0.13, 0.18, 0.19, 0.22, 0.37, 0.39, 0.47, 0.50, 0.54, 0.62 and 0.76	0.19, 0.37, 0.39, 0.54 and 0.76		-
16	WH 730/UP 2338	0.13 to 0.76	0.13, 0.19, 0.23, 0.28, 0.33, 0.37, 0.39, 0.41, 0.46, 0.49, 0.54, 0.60, 0.63, 0.72 and 0.76	value 0.13, 0.23, 0.33, 0.37, 0.41, 0.46, 0.49, 0.54, 0.60, 0.63, 0.72 and 0.76	0.13, 0.19, 0.23, 0.28, 0.33, 0.37, 0.39, 0.41, 0.46, 0.49, 0.54, 0.60, 0.63, 0.72 and 0.76		0.11, 0.18, 0.19, 0.22, 0.28, 0.30, 0.39, 0.47, 0.50, 0.54 and 0.62	value 0.19, 0.28 and 0.39		
17	JOB 666/UP 2338	0.13 to 0.76	0.13, 0.19, 0.21, 0.23, 0.30, 0.33, 0.37, 0.39, 0.41, 0.46, 0.49, 0.54, 0.60, 0.63, 0.72 and 0.76	0.13, 0.21, 0.23, 0.33, 0.37, 0.41, 0.46, 0.49, 0.60, 0.63, 0.72 and 0.76	0.13, 0.15, 0.21, 0.23, 0.28, 0.30, 0.33, 0.37, 0.39, 0.41, 0.46, 0.49, 0.60, 0.63, 0.72 and 0.76	0.19	0.11, 0.18, 0.19, 0.22, 0.28, 0.30, 0.39, 0.47, 0.50, 0.54 and 0.62	0.30, 0.39 and 0.54		0.28
18	JOB 666/UP 2425	0.11 to 0.76	0.13, 0.14, 0.19, 0.21, 0.23, 0.37, 0.39, 0.41, 0.49, 0.60, 0.54, 0.63, 0.72, 0.76	0.13, 0.21, 0.23, 0.41, 0.49, 0.60, 0.63 and 0.72	0.13, 0.15, 0.21, 0.23, 0.28, 0.30, 0.33, 0.37, 0.39, 0.41, 0.46, 0.49, 0.54, 0.60, 0.63, 0.72 and 0.76	0.19	0.11, 0.13, 0.18, 0.19, 0.22, 0.37, 0.39, 0.47, 0.50, 0.54, 0.62 and 0.76	0.37, 0.39, and 0.76	0.14	
19	NI 5439/UP 2425	0.13 to 0.76	0.13, 0.14, 0.19, 0.21, 0.23, 0.30, 0.33, 0.37, 0.39, 0.41, 0.49, 0.60, 0.63, 0.72, 0.76	0.21, 0.23, 0.30, 0.33, 0.41, 0.49, 0.60, 0.63 and 0.72	0.13, 0.21, 0.23, 0.30, 0.33, 0.37, 0.39, 0.41, 0.49, 0.60, 0.63, 0.72 and 0.76.	0.19	0.11, 0.13, 0.18, 0.19, 0.22, 0.37, 0.39, 0.47, 0.50, 0.54, 0.62 and 0.76	0.13, 0.37, 0.39 and 0.76	0.14	
20	NI 5439/UP 2338	0.13 to 0.76	0.13, 0.14, 0.19, 0.21, 0.23, 0.30, 0.33, 0.37, 0.39, 0.41, 0.49, 0.60, 0.63, 0.72 and 0.76	0.13, 0.21, 0.23, 0.33, 0.37, 0.41, 0.49, 0.60, 0.63, 0.72 and 0.76	0.13, 0.21, 0.23, 0.30, 0.33, 0.37, 0.39, 0.41, 0.49, 0.60, 0.63, 0.72 and 0.76	0.19	0.11, 0.18, 0.19, 0.22, 0.28, 0.30, 0.39, 0.47, 0.50, 0.54 and 0.62.	0.30, 0.39	0.14	
21	NP 846/UP 2338	0.11 to 0.76	0.11, 0.13, 0.14, 0.19, 0.21, 0.26, 0.28, 0.35, 0.39, 0.41, 0.46, 0.49, 0.52, 0.59, 0.67, 0.72 and 0.76	0.11, 0.13, 0.14, 0.19, 0.21, 0.26, 0.28, 0.35, 0.39, 0.41, 0.46, 0.49, 0.52, 0.59, 0.67, 0.72 and 0.76	0.11, 0.13, 0.14, 0.19, 0.21, 0.26, 0.28, 0.35, 0.39, 0.41, 0.46, 0.49, 0.52, 0.59, 0.67, 0.72 and 0.76		0.11, 0.18, 0.19, 0.22, 0.28, 0.30, 0.39, 0.47, 0.50, 0.54 and 0.62	0.11, 0.19 and 0.39		
22	NP 846/UP 2425	0.11 to 0.76	0.11, 0.13, 0.14, 0.19, 0.21, 0.26, 0.28, 0.35,	value 0.11, 0.13, 0.14, 0.19, 0.21, 0.26,	0.11, 0.13, 0.14, 0.19, 0.21, 0.26, 0.28, 0.35, 0.39,		0.11, 0.13, 0.18, 0.19, 0.22, 0.37, 0.39, 0.47,	0.11, 0.13, 0.19, 0.39 and 0.76		

			0.39, 0.41, 0.46, 0.49, 0.52, 0.59, 0.67, 0.72 and 0.76	0.28, 0.35, 0.39, 0.41, 0.46, 0.49, 0.52, 0.59, 0.67, 0.72 and 0.76	0.41, 0.46, 0.49, 0.52, 0.59, 0.67, 0.72 and 0.76		0.50, 0.54, 0.62 and 0.76			
23	NIAW 34/PBW 373	0.10 to 0.76	0.10, 0.13, 0.14, 0.21, 0.33, 0.35, 0.39, 0.46, 0.49, 0.52, 0.59, 0.67, 0.72 and 0.76	0.10, 0.13, 0.14, 0.21, 0.33, 0.35, 0.39, 0.46, 0.49, 0.52, 0.59, 0.67, 0.72 and 0.76	0.10, 0.13, 0.14, 0.21, 0.33, 0.35, 0.39, 0.46, 0.49, 0.52, 0.59, 0.67, 0.72 and 0.76		0.09, 0.13, 0.18, 0.22, 0.37, 0.39, 0.47, 0.50, 0.54, 0.62 and 0.76.	0.13, 0.39 and 0.76		
24	NIAW 34/UP 2338	0.11 to 0.76	0.10, 0.13, 0.21, 0.33, 0.35, 0.39, 0.46, 0.49, 0.52, 0.59, 0.67 and 0.72, 0.76	0.10, 0.13, 0.21, 0.33, 0.35, 0.49, 0.52, 0.59, 0.67 and 0.72	0.10, 0.13, 0.14, 0.21, .33, 0.35, 0.39, 0.46, 0.49, .52, 0.59, 0.67, 0.72 and 0.76		0.11, 0.18, 0.19, 0.22, 0.28, 0.30, 0.39, 0.47, 0.50, 0.54 and 0.62.	0.39, 0.46 and 0.76		
25	HI 385/PBW 373	0.10 to 0.76	0.10, 0.13, 0.14, 0.19, 0.21, 0.28, 0.33, 0.35, 0.39, 0.46, 0.49, 0.52 0.67, and 0.72, 0.76	0.13, 0.14, 0.19, 0.21, 0.28, 0.33, 0.46, 0.49, 0.52 and 0.72, 0.76	0.13, 0.14, 0.19, 0.21, 0.28, 0.33, 0.35, 0.39, 0.46, .49, 0.52, 0.59, 0.67, 0.72 and 0.76		0.09, 0.13, 0.18, 0.22, 0.37, 0.39, 0.47, 0.50, 0.54, 0.62 and 0.76	0.13, 0.35, 0.39, 0.67 and 0.76	0.10	
26	HI 385/UP 2338	0.11 to 0.76	0.11, 0.13, 0.14, 0.19, 0.21, 0.28, 0.33, 0.35, 0.39, 0.46, 0.49, 0.52, 0.59, 0.67 and 0.72	0.13, 0.14, 0.19, 0.21, 0.33, 0.39, 0.49, 0.52, 0.59, 0.67 and 0.72	0.13, 0.14, 0.19, 0.21, .28, 0.33, 0.35, 0.39, 0.46, .49, 0.52, 0.59, 0.67, 0.72 and 0.76	0.11	0.11, 0.18, 0.19, 0.22, 0.28, 0.30, 0.39, 0.47, 0.50, 0.54 and 0.62	0.28, 0.35, 0.39 and 0.46		
27	HI 385/UP 2425	0.11 to 0.76	0.11, 0.13, 0.14, 0.19, 0.21, 0.28, 0.33, 0.35, 0.39, 0.46, 0.49, 0.52, 0.67 and 0.72, 0.76	0.14, 0.21, 0.33, 0.46, 0.49, 0.52 and 0.72	0.13, 0.14, 0.19, 0.21, 0.28, 0.33, 0.35, 0.39, 0.46, 0.49, 0.52, 0.59, 0.67, 0.72 and 0.76	0.11	0.11, 0.13, 0.18, 0.19, 0.22, 0.37, 0.39, 0.47, 0.50, 0.54, 0.62 and 0.76	0.13, 0.19, 0.28, 0.35, 0.39, 0.67 and 0.76		
28	PBN 51/UP 2425	0.12 to 0.76.	0.12, 0.17, 0.22, 0.28, 0.30, 0.35, 0.46, 0.49, 0.58, 0.67 and 0.72	0.30, 0.46, 0.49, 0.58 and 0.72	value 0.12, 0.17, 0.22, 0.28, 0.30, 0.35, 0.46, 0.49, 0.58, 0.67, 0.72 and 0.76		0.12, 0.17, 0.20, 0.22, 0.28, 0.35, 0.40, 0.44, 0.54, 0.67 and 0.76.	0.12, 0.17, 0.22, 0.28, 0.35, 0.67 and 0.76		
29	PBN 51/UP 2338	0.11 to 0.76.	0.12, 0.17, 0.20, 0.22, 0.28, 0.30, 0.35, 0.46, 0.49, 0.58, 0.67 and 0.72, 0.76	0.28, 0.30, 0.49 and 0.72	0.12, 0.17, 0.22, 0.28, 0.30, 0.35, 0.46, 0.49, 0.58, 0.67, 0.72 and 0.76	0.20	0.10, 0.12, 0.17, 0.20, 0.22, 0.35, 0.40, 0.46, 0.54, 0.58, 0.67 and 0.76.	0.12, 0.17, 0.20, 0.22, 0.35, 0.46, 0.58, 0.67 and 0.76	0.15	
30	Halna/UP 2425	0.12 to 0.76	0.12, 0.17, 0.20, 0.22, 0.28, 0.35, 0.40, 0.44, 0.46, 0.49 and 0.58, 0.67, 0.72	0.46, 0.49 and 0.58	0.10, 0.12, 0.15, 0.17, .20, 0.22, 0.28, 0.35, 0.40, 0.44, 0.46, 0.49, 0.58, 0.67, 0.72 and 0.76		0.12, 0.17, 0.20, 0.22, 0.28, 0.35, 0.40, 0.44, 0.54, 0.67 and 0.76	0.12, 0.17, 0.20, 0.22, 0.28, 0.35, 0.40, 0.44, 0.46, 0.49, 0.58, 0.67 and 0.72		
31	Halna/UP 2338	0.12 to 0.76.	0.12, 0.17, 0.20, 0.22, 0.28, 0.35, 0.40, 0.44, 0.46, 0.49, 0.58, 0.67, 0.72 and 0.76	0.28, 0.44, 0.49, and 0.72	0.10, 0.12, 0.15, 0.17, 0.20, 0.22, 0.28, 0.35, 0.40, 0.44, 0.46, 0.49, 0.58, 0.67, 0.72 and 0.76		0.10, 0.12, 0.17, 0.20, 0.22, 0.35, 0.40, 0.46, 0.54, 0.58, 0.67 and 0.76.	0.12, 0.17, 0.20, 0.22, 0.35, 0.40, 0.46, 0.46, 0.58, 0.67 and 0.76		0.10
32	Halna/PBW 373	0.12 to 0.76	0.12, 0.17, 0.20, 0.22, 0.28, 0.35, 0.40, 0.44, 0.46, 0.49, 0.58, 0.67 and 0.76	0.12, 0.20, 0.28, 0.40, 0.44, 0.46, 0.49 and 0.58	0.10, 0.12, 0.15, 0.17, 0.20, 0.22, 0.28, 0.35, 0.40, 0.44, 0.46, 0.49, 0.58, 0.67, 0.72 and 0.76		0.11, 0.15, 0.17, 0.22, 0.35, 0.54, 0.67 and 0.76.	0.17, 0.22, 0.35, 0.67 and 0.76		0.15

Fig.1 Protein banding profile of parents and F₁'s by SDS-PAGE

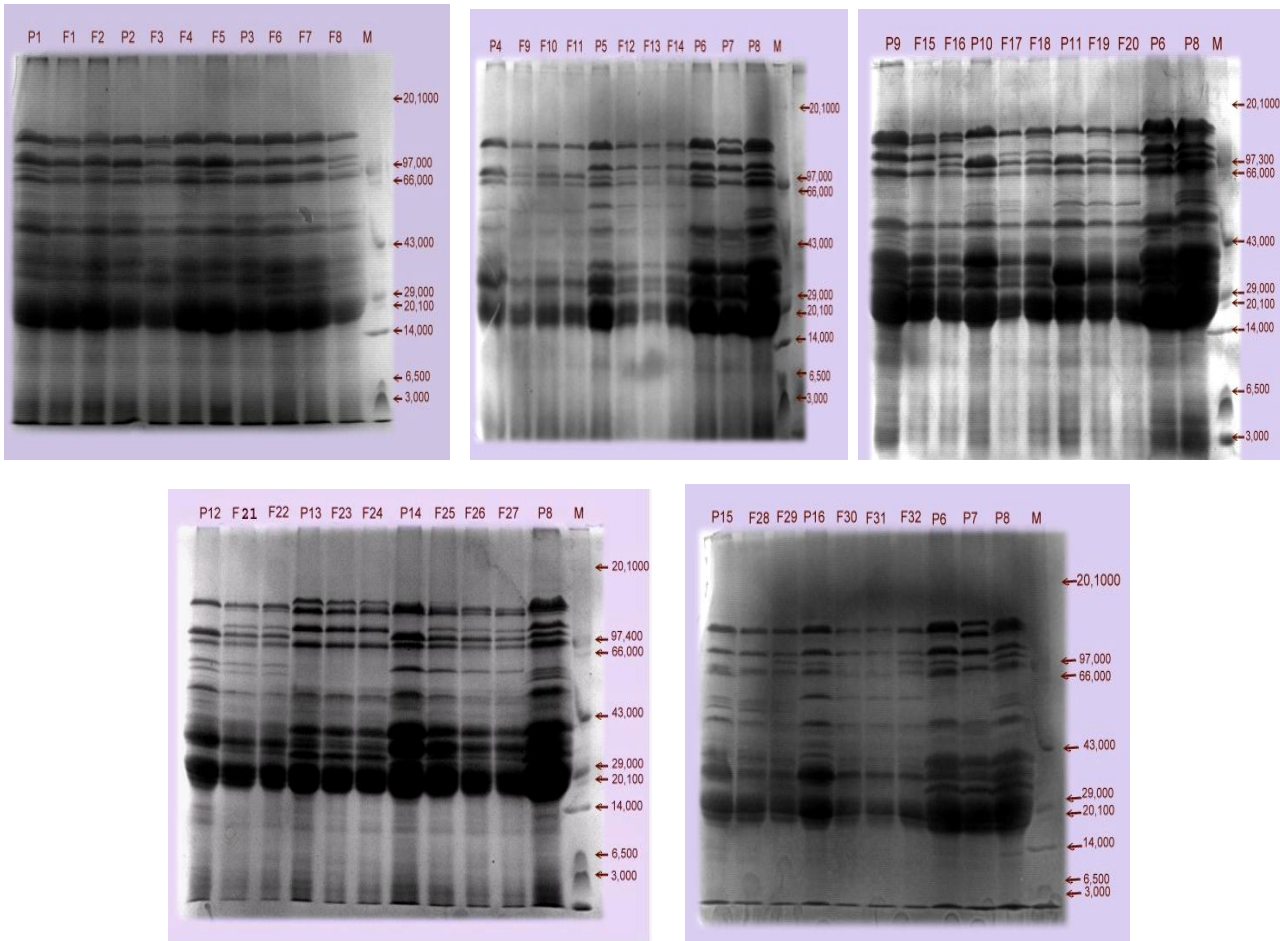


Fig.2 Protein banding profile of parents and F₁'s by SDS-PAGE.

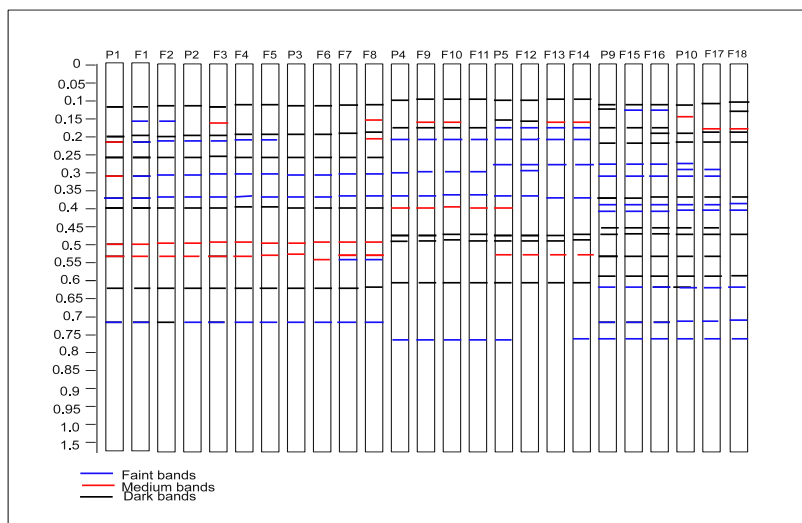


Fig. 3: Protein banding profile of parents and F₁'s by SDS-PAGE.

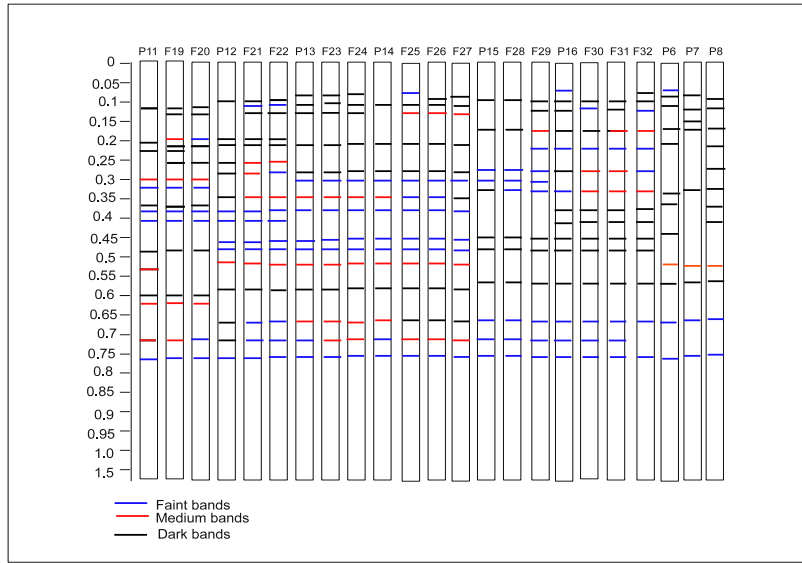


Fig.4 Similarity index of parental lines

1.00 0																	
0.98 5	1.00 0																
0.970 5	0.98 5	1.00 0															
0.79 4	0.80 8	0.79 4	1.00 0														
0.77 9	0.79 4	0.77 9	0.89 7	1.00 0													
0.75 0	0.73 5	0.75 0	0.72 0	0.73 5	1.00 0												
0.72 0	0.70 5	0.72 0	0.69 1	0.67 6	0.91 1	1.00 0											
0.77 9	0.76 4	0.77 9	0.75 0	0.67 6	0.88 2	0.94 1	1.00 0										
0.64 7	0.66 1	0.67 6	0.67 6	0.66 1	0.77 9	0.75 0	0.75 0	1.00 0									
0.70 5	0.69 1	0.70 5	0.67 6	0.63 2	0.77 9	0.77 9	0.80 8	0.88 2	1.00 0								
0.69 1	0.67 6	0.69 1	0.69 1	0.67 6	0.82 3	0.79 4	0.79 4	0.92 6	0.95 5	1.00 0							
0.70 5	0.72 0	0.70 5	0.76 4	0.72 0	0.75 0	0.75 0	0.75 0	0.76 4	0.79 4	0.80 8	1.00 0						
0.70 5	0.72 0	0.70 5	0.70 5	0.72 0	0.69 1	0.66 1	0.73 5	0.73 5	0.76 4	0.75 0	0.91 1	1.00 0					
0.76 4	0.77 9	0.76 4	0.88 2	0.92 6	0.77 9	0.72 0	0.75 0	0.70 5	0.70 5	0.72 0	0.70 5	0.64 7	1.00 0				
0.77 9	0.79 4	0.77 9	0.83 8	0.88 2	0.76 4	0.73 5	0.76 4	0.66 1	0.72 0	0.70 5	0.72 0	0.66 1	0.95 5	1.00 0			
0.75 0	0.73 5	0.72 0	0.83 8	0.91 1	0.73 5	0.70 5	0.70 5	0.66 1	0.66 1	0.70 5	0.72 0	0.66 1	0.89 7	0.85 2	1.00 0		

Fig 5: Combined similarity index of parental lines and F₁s

1.000
1.000 1.000
1.000 1.000 1.000
0.971 0.971 0.971 1.000
0.97 0.971 0.971 1.000 1.000
0.971 0.971 0.971 0.971 0.971 1.000
0.956 0.956 0.956 0.926 0.926 0.956 1.000
0.956 0.956 0.956 0.956 0.956 0.956 0.971 1.000
0.779 0.779 0.779 0.809 0.809 0.779 0.765 0.794 1.000
0.765 0.765 0.765 0.794 0.794 0.765 0.750 0.779 0.985 1.000
0.794 0.794 0.794 0.824 0.824 0.794 0.779 0.809 0.985 0.971 1.000
0.779 0.779 0.779 0.779 0.779 0.779 0.765 0.794 0.941 0.956 0.926 1.000
0.750 0.750 0.750 0.750 0.750 0.779 0.794 0.794 0.912 0.926 0.897 0.941 1.000
0.779 0.779 0.779 0.809 0.809 0.779 0.794 0.824 0.941 0.926 0.926 0.912 0.941 1.000
0.706 0.706 0.706 0.7356 0.735 0.735 0.750 0.750 0.691 0.706 0.706 0.721 0.750 0.721 1.000
0.691 0.691 0.691 0.721 0.721 0.721 0.351 0.735 0.676 0.691 0.691 0.706 0.735 0.706 0.985 1.000
0.706 0.706 0.706 0.735 0.735 0.735 0.750 0.750 0.721 0.735 0.735 0.721 0.721 0.691 0.941 0.956 1.000
0.750 0.750 0.750 0.779 0.779 0.779 0.765 0.765 0.735 0.750 0.750 0.735 0.735 0.706 0.926 0.941 0.926 1.000
0.721 0.721 0.721 0.750 0.750 0.750 0.735 0.735 0.735 0.750 0.750 0.735 0.706 0.676 0.926 0.941 0.956 0.971 1.000
0.721 0.721 0.721 0.750 0.750 0.750 0.735 0.735 0.735 0.750 0.750 0.735 0.706 0.676 0.926 0.941 0.956 0.971 1.000 1.000
0.647 0.647 0.647 0.676 0.676 0.676 0.662 0.662 0.662 0.676 0.676 0.691 0.691 0.662 0.824 0.838 0.794 0.838 0.809 0.809 1.000
0.676 0.676 0.676 0.706 0.706 0.706 0.691 0.691 0.662 0.676 0.676 0.691 0.691 0.662 0.824 0.838 0.794 0.838 0.809 0.809 0.971 1.000
0.676 0.676 0.676 0.706 0.706 0.706 0.691 0.691 0.662 0.676 0.676 0.662 0.662 0.662 0.794 0.809 0.794 0.809 0.809 0.809 0.882 0.882 1.000
0.691 0.691 0.691 0.721 0.721 0.721 0.706 0.706 0.676 0.691 0.691 0.676 0.676 0.676 0.779 0.794 0.809 0.794 0.794 0.794 0.868 0.868 0.985 1.000
0.676 0.676 0.676 0.706 0.706 0.706 0.691 0.691 0.662 0.676 0.676 0.691 0.691 0.662 0.853 0.868 0.824 0.838 0.838 0.838 0.882 0.912 0.941 0.926 1.000
0.676 0.676 0.676 0.676 0.676 0.706 0.691 0.691 0.662 0.676 0.676 0.721 0.721 0.662 0.824 0.838 0.794 0.809 0.809 0.809 0.912 0.941 0.912 0.897 0.941 1.000
0.662 0.662 0.662 0.691 0.691 0.691 0.676 0.676 0.676 0.691 0.691 0.706 0.706 0.676 0.868 0.882 0.838 0.853 0.853 0.853 0.897 0.926 0.897 0.882 0.956 0.956 1.000
0.721 0.721 0.721 0.721 0.721 0.706 0.735 0.735 0.750 0.750 0.794 0.735 0.735 0.721 0.706 0.721 0.706 0.706 0.706 0.750 0.750 0.779 0.794 0.779 0.765 1.000
0.721 0.721 0.721 0.750 0.750 0.721 0.706 0.735 0.735 0.750 0.750 0.765 0.706 0.735 0.721 0.706 0.721 0.706 0.706 0.706 0.750 0.750

Fig.6 Dendrogram showing diversity among parental lines

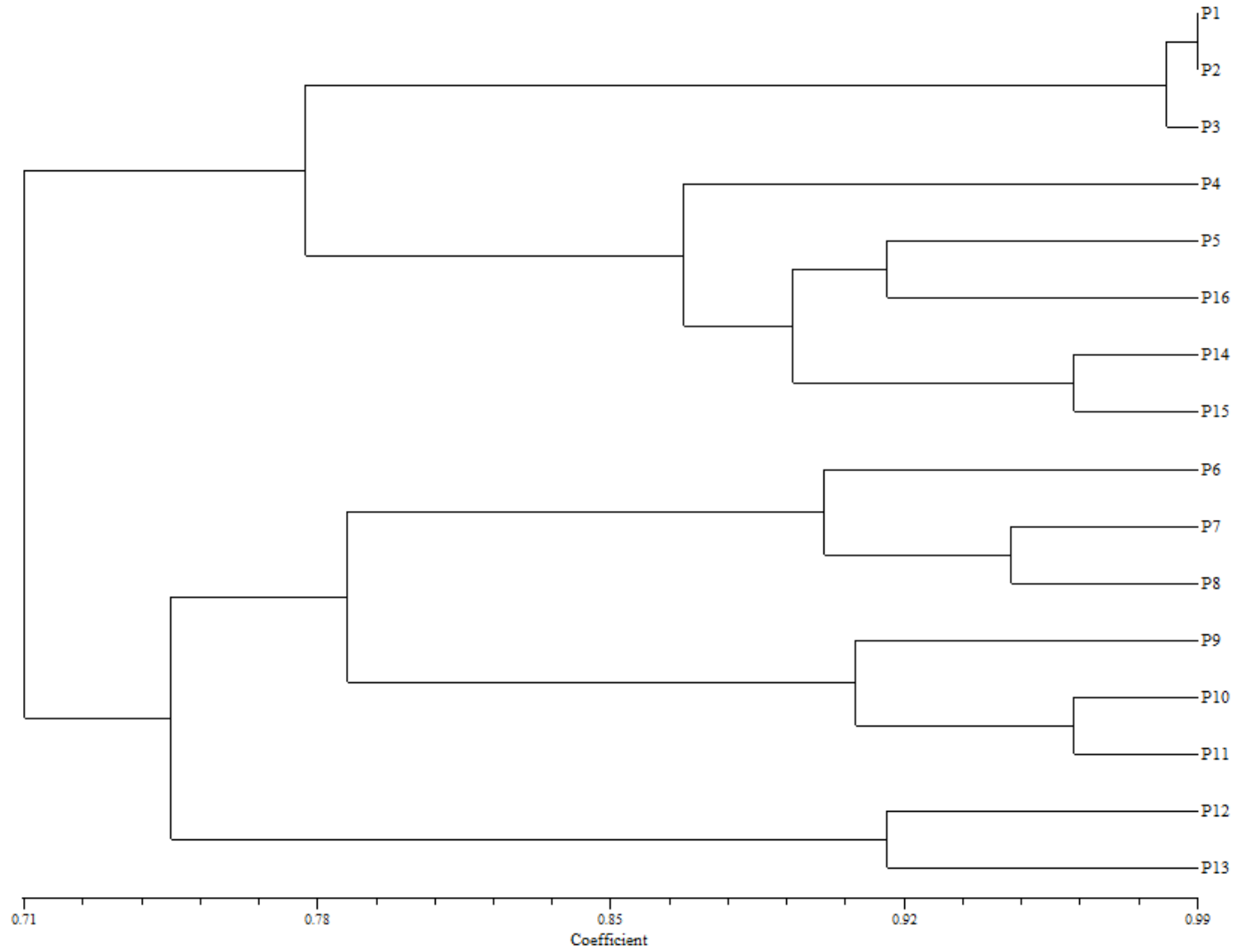
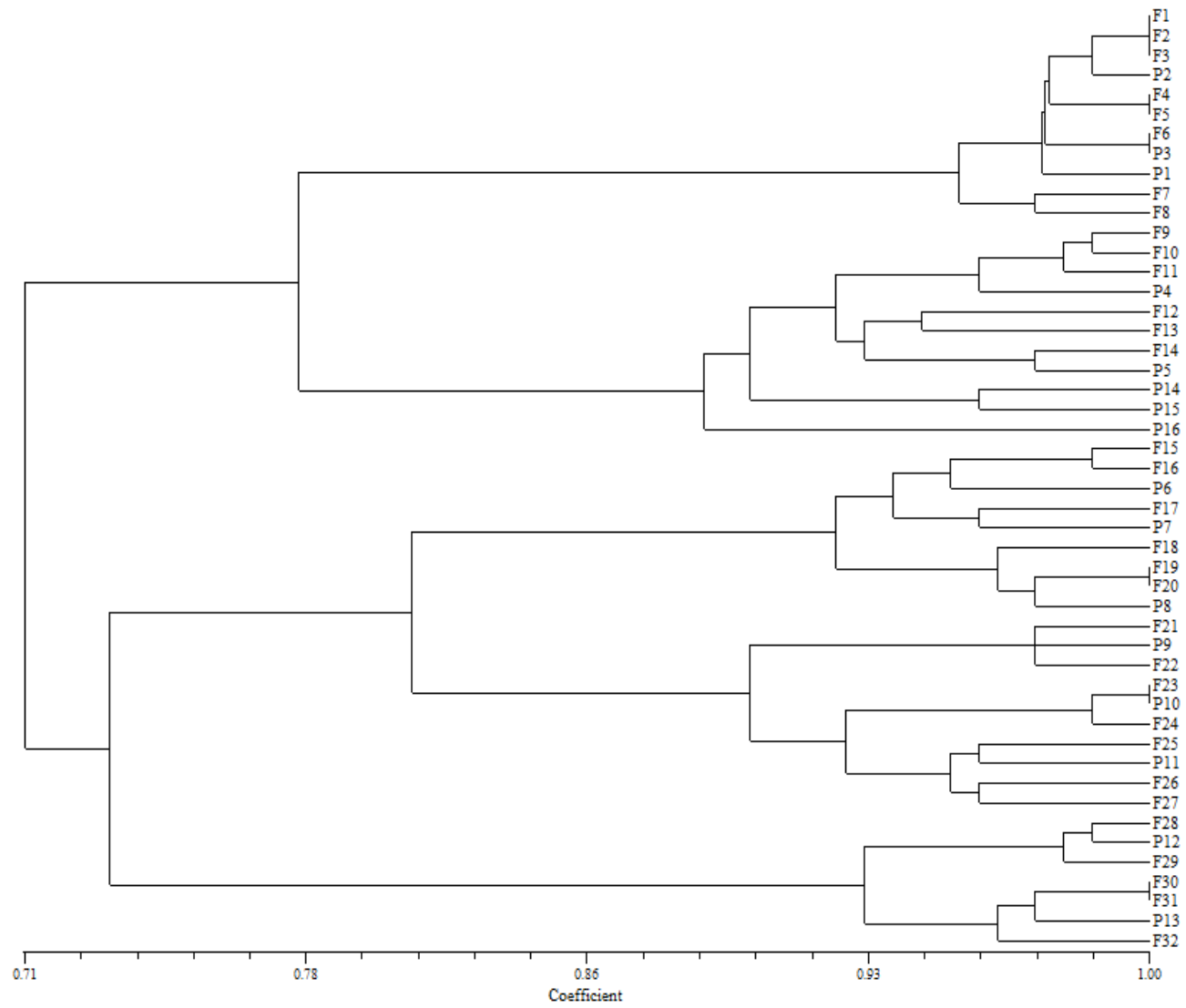


Fig.7 Combine dendrogram of parents and F1s prepared by NT-SYS 2.0



Seed protein pattern can also be used as a promising tool for distinguishing cultivars of particular crop species (Jha and Ohri, 1996; Seferoglu *et al.*, 2006). The SDS-PAGE is considered to be a sensible and reliable technique for species identification.

One band of Rf value 0.72 was present in all the tolerant parents and F₁'s but absent in the drought susceptible parents, so the protein corresponding to 0.72 rf value may be responsible for the drought tolerance. After validation this band can be used as a marker for drought related studies and breeding programmes (Balass *et al.*, 1992; Ekramoddoullah and Tan, 1998; Pattanaik and Kole, 2002 and Kour, 2005). Specific protein profiling for target trait was used for quality assessment of cultivars (Katyal *et al.*, 2018).

Presence of unique bands in some genotypes could be used for the characterization and identification of varieties as well as hybrids. Presence of unique bands in F₁'s is an indicative of new genetic constitution as a result of recombination. New genetic constitution causes formation of new protein and ultimate development of a new band that was not present in either of the parent. The same may be the reason for absence of band where new genetic architecture leads to the no translation. Unique bands were found in cross VL 804/UP 2425, VL804/ PBW 373, VL 802/ UP 2338 and UP 2572/UP 2338 at Rf value 0.16, Job 666/UP 2425, NI5439/UP 2425 and NI 5439/UP 2338 at 0.14, HI 385/ PBW 2338 at 0.10 and PBN-51/UP2338 at Rf value 0.15.

Absence of bands was reported in cross UP 2572/UP 2338 at 0.31, PBW 175/UP 2338 at 0.40, PBW 175/UP 2425 at 0.40, 0.76, PBW 175/PBW 373 at 0.40, JOB 666/UP 2338 at 0.28, Halna/UP 2338 at 0.10 and Halna/PBW 373 at 0.40 Rf value respectively. Anuradh *et*

al., (1990) in pearl millet, Nagaraja *et al.*, (2000) and Chauhan *et al.*, (2002) in sorghum, Limbu *et al.*, (2013) in Indian mustard, Sharma and Krishna (2017) in cow pea and Alice *et al.*, (2017) in chilli mentioned SDS-PAGE as a useful protein marker for varietal identification.

Seed protein is always a reliable method for examination of genetic diversity, characterization and identification of genotypes. In other words it emerged as a powerful tool to assess inter-and intra species variation (Sofalian *et al.*, 2015). Studies on allelic diversity indicated the expansion of SDS-PAGE as biochemical markers for variable purposes (Kakaei, 2018). It is effectively and widely used in insuring the genetic purity of lines as well as hybrids i.e. contributing significantly towards maintenance breeding of cultivars and ensuring the supply of quality seed. In wheat seed storage protein profile could be used as protein marker in the studied of diversity, hybridity, characterization, and identification of adapted cultivars thereby improving the efficiency of wheat breeding programme and varietal development. It is suggested that other complementary biochemical analysis like 2D electrophoresis must be carried out on genotypes with the same electrophoretic pattern in order to have more precise diverse profiles.

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