

Review Article

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## Dealing with Zinc and Iron Deficiency in Rice: Combine Strategies to Fight Hidden Hunger in Developing Countries

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### ABSTRACT

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Zinc and Iron are essential micronutrient for both plant growth and human health but it is often reported to be deficient in regions where rice is use as staple food. Although significant progresses are made in understanding genetic and molecular mechanism of micronutrient acquisition but these need to be characterize to increase the bioavailability of these micronutrients. Biofortification is suggested to be a sustainable and cost-effective approach in this perspective and for that combination of various agronomic and genetic strategies should be put in place without delay.

#### Introduction

Rice is the primary staple food for more than half the world's population and together they directly supply more than 50% of all calories consume by the entire human population (Jia-Yang *et al.*, 2014). Total rice production is increases to 751.9 million tonnes worldwide (FAO, 2017) and among that 90 percent is produce and consume in developing countries. But unfortunately, about 870 million people are suffering from chronic undernourishment globally (Da Silva *et al.*, 2013) and vast majority of them are from developing countries where rice is closely associated with food security and political stability. So, improving the micronutrient status of rice is very important to tackle key nutrition and health related problems of these large numbers of populations, most notably developing countries.

Among the various micronutrients, iron (Fe) and zinc (Zn) are important for both plant growth and human health. In developing countries, iron and zinc deficiencies are reported to be the sixth and fifth highest health risk factor respectively (Freitas *et al.*, 2016; Sharma *et al.*, 2013) causing a high mortality rates. So, overcoming these nutritional deficiencies is need of hour.

Various strategies to improve micronutrient status include food supplementation, food fortification and biofortification (Masuda *et al.*, 2013). Among them biofortification is appears to be the most feasible, sustainable and economical as poor families of developing countries cannot afford other strategies (Nakandalage *et al.*, 2016). For this, selection of effective genetic and crop management approach is of utmost importance.

## Importance of zinc

### Role in plants

Zinc is one of the key micronutrient involve in regulating various biological and physiological processes in plants. In rice tissues, typical zinc concentration is around 35 to 100 ppm and deficiency symptoms appear when concentration drops below 20 ppm. Zinc deficiency affects photosynthesis due to altered chloroplast pigments (Table 1) (Samreen *et al.*, 2017) and results in short internodes, decrease in leaf size and delayed maturity, sterile spikes, leaves with brown botches and streaks (Abdullah, 2015).

Further it reduces pollen viability leading to fewer grain set and severe yield penalties worldwide (Disante *et al.*, 2010).

### Impact in human health

Zinc is one of the important trace elements whose role in human health is undisputable. Cellular zinc homeostasis is important for proper release and action of insulin (Rutter *et al.*, 2016), modulating oxidative stress and various age-related disorder (Prasad, 2013). Insufficient intake of zinc in humans include emotional disorder, weight loss, dysfunctions, atherosclerosis, several malignancies, alopecia, diarrhea (Rutter *et al.*, 2016, Chasapis *et al.*, 2012) decline in immune competence and certain neurological and physiological problem (Roohani *et al.*, 2013).

## Importance of iron

### Role in plants

Iron is one of the important micronutrient that

requires to maintain proper metabolic and physiological processes in plants. It acts as cofactor for many enzymes and proteins of mitochondria and chloroplast and hence it has major role in life sustaining processes like photosynthesis and respiration. It has role in scavenging of ROS and act as key element to ensure electron flow through the PSII–b6f/Rieske–PSI complex in chloroplast (Zargar *et al.*, 2015). Further insufficient iron uptake leads to iron deficiency symptoms such as interveinal yellowing and chlorosis of emerging leaves, less dry matter production, reduced sugar metabolism enzymes (El-Jendoubi *et al.*, 2014; Das, 2014), seed dormancy (Murgia *et al.*, 2017).

### Impact in human health

Iron is the most abundant transition metal involve in various biological processes. Almost two-thirds of the body iron is found in the hemoglobin present in circulating erythrocytes, 25% is contained in a readily mobilizable iron store and the remaining 15% is bound to myoglobin in muscle tissue and in a variety of enzymes involved in the oxidative metabolism and many other cell functions (IOM, 2001).

Abnormal iron homoeostasis can induce cellular damage through hydroxyl radical production which can cause the oxidation and modification of lipids, proteins, carbohydrates, DNA and leads to various neuro generative diseases like Alzheimer's disease and Parkinson's disease (Ward *et al.*, 2014). Further iron deficiency anaemia is a major problem affecting around 2 billion people in both developed and developing countries (WHO, 2016).

**Table.1** Chlorophyll contents (mg kg<sup>-1</sup>) on dry weight basis in mungbean varieties at different concentrations of Zn in solution culture

Zn treatment	V1	V2	V3	V4	Mean±St.dv
Control	35.7f	73.45de	93.12 cd	105.93c	78.55b 30.63
1µM	36.81f	145.30b	210.82a	221.01a	153.5a 84.71
2 µM	64.54e	146.07b	210.57a	226.08a	161.9a 73.52
Mean±St.dv	45.69c	123.6b	171.5a	184.4a	
	16.34	41.71	67.88	67.95	

V1 = Ramazan, V2 = Swat mungI, V3 = NM92, V4 = KMI.St. d = standard deviation. The mean followed by similar letter (s) are not significantly different at *P* = 0.05.

**Table 2.** Effect of different forms of foliar Zn fertilization on the percentages of solubility, retention, transported and uptake efficiency of Zn among three rice cultivars

Treatments	Cultivars <sup>a</sup>					
	Hai7	Bing91185	Biyuzaonuo	Hai7	Bing91185	Biyuzaonuo
	Solubility (%)			Retention (%)		
Control	29.17 c	30.58 c	25.70 b	14.53 c	14.91b	14.39 c
Zn-EDTA	30.75 c	31.21 bc	26.06 b	14.82 bc	14.96 b	15.04 bc
Zn-Citrate	30.90 bc	31.13 bc	26.20 b	14.93 bc	15.78 b	15.11 bc
ZnSO4	32.68 a	32.24 a	28.54 a	16.68 ab	18.29a	16.80 ab
Zn-AA	31.64 ab	32.62 a	27.73 a	17.54 a	18.80a	17.53 a
Zn effect by f-test <sup>b</sup>	*	*	***	*	***	*
	Transport (%)			Uptake efficiency (%)		
Control	9.35 b	16.09 b	8.99 c	6.95 c	9.48 b	6.01 b
Zn-EDTA	9.61 b	16.23 b	9.57 c	7.52 bc	9.73 b	6.39 b
Zn-Citrate	11.08 ab	16.15 b	9.96 bc	8.02 b	9.94 b	6.57 b
ZnSO4	13.27 a	18.43 ab	13.53 a	9.79 a	11.84 a	8.66 a
Zn-AA	13.05 a	19.11 a	12.42 ab	9.68 a	12.39 a	8.30 a
Zn effect by f-test <sup>b</sup>	**	*	*	***	***	***

aDifferent letters after number in the same column designated significant difference by LSDP,0.05.b Significant effects: NS = not significant at P.0.05\*at P,0.05; \*\*at P,0.01;\*\*\*at P,0.001.

**Table.3** Main effects of cultivation system, genotype, and Fe application on shoot dry weight, shoot Fe concentration, and shoot Fe content of rice at tillering stage

Main effects and factors within main effects	Shoot dry weight (kg ha <sup>-1</sup> )	Shoot Fe concentration (mg kg <sup>-1</sup> )	Shoot Fe content (g ha <sup>-1</sup> )
Cultivation system			
Aerobic	935 a	294 b	27 b
Flooded	825 b	393 a	32 a
Genotype			
Qiuguang	659 c	378 a	25 b
K150	788 bc	295 b	23 b
Han72	759 bc	361 ab	27 b
89B-271-17(hun)	898 b	332 ab	30 ab
Han277	1059 a	348 ab	36 a
Han297	1119 a	348 ab	36 a
Average	880	344	30
Fe application			
0 (kg ha <sup>-1</sup> )	819 b	328 a	25 b
30 (kg ha <sup>-1</sup> )	941 a	358 a	34 a

For each main effect, values in a column followed by the same letter are not significantly different ( $P > 0.05$ ).

**Table4** Zn concentrations in shoot and root of rice under different water regimes and Zn source treatments

Genotype	Zn treatment	Shoot Zn concentration (mg/kg)		Root Zn concentration (mg/kg)	
		CF	AWD	CF	AWD
Nipponbar	Control	50.2b	54.6b	86.9c	90.7c
	ZnSO4	60.7a	63.6a	130.6a	143.9a
	Zn-EDTA	59.6a	61.3a	119.5b	117.7b
	Mean	56.9B	59.8B	112.3A	117.5B
Jiaxing27	Control	62.0b	65.3b	96.2c	102.1b
	ZnSO4	68.7a	72.6a	144.5a	153.5a
	Zn-EDTA	66.2ab	71.5a	130.0b	149.9a
	Mean	65.6A	69.8A	123.6A	135.1A

Within a column, means followed by different letters are significantly different at  $P < 0.05$  according to Duncan's multiple range test. Lower-case and upper-case letters indicate comparisons among three Zn treatments and between two genotypes, respectively

**Table 5** Iron and Zn concentrations in individual plant tissues of transgenic progeny classified as high-yield(CHY) and low-yield(CLY)in the OE-*OsNAS*/IR64 and OE-*OsNAS*/Esp progenies

Progeny type	Concentration (mg g <sup>-1</sup> DW)											
	Root		Stem/sheath		Non-flag leaf		Flag leaf		Panicle		Grain	
	Fe	Zn	Fe	Zn	Fe	Zn	Fe	Zn	Fe	Zn	Fe	Zn
<i>OE-OsNAS/IR64</i>												
NS (n D 3)	6267	18.7	273	7.8	237	10.2	205	9.8b	89b	16.9b	14.4b	15.3b
+C HY (n D 6)	7350	18.5	258	12.0	251	10.6	2131	1.3b	107ab	11.1b	18.0b	23.2b
+LY(n D 4)	9150	22.5	283	26.2	253	15.6	193	16.6a	122a	34.6a	28.8a	55.9a
<i>P</i> -value (progenytyp)	n.s.	n.s.	n.s.	.n.s.	n.s.	.n.s.	n.s.	**	*	**	**	***
<i>OE-OsNAS/Esp</i>												
NS (n D 3)	8000	25.0	207	9.0	295	13.2	295	15.2	155	10.0	16.1b	20.2b
+ HY (n D 10)	8120	43.4	234	19.0	337	13.7	296	17.9	157	15.2	28.6a	38.3a
+ LY(n D 3)	6333	59.0	243	16.0	423	10.53	201	4.2	173	16.13	6.1a	63.0a
<i>P</i> -value (progenytyp)	n.s.	n.s.	n.s.	.n.s.	n.s.	.n.s.	n.s.	n.s.	n.s.	n.s.	***	***

n.s.,not significant. Within each column, values with different letters represent significant differences between progeny type at the 5% level by Hochberg's GT2 test. The values given are means. \* $P < 0.05$ , \*\* $P < 0.01$ , \*\*\* $P < 0.001$ . NS, null segregants; DW, dry weight

### Agronomic strategy for improving iron and zinc uptake

#### Application of fertilizers

Nitrogen (N) is an essential macronutrient (Sarwar *et al.*, 2010) which helps to improve translocation of other micronutrients like iron and zinc in various plants. Better N nutrition promotes protein synthesis, which is a major sink for Fe and Zn and enhances the expression

Zn and Fe transporter proteins, such as ZIP family transporters (Cakmak *et al.*, 2010), YSL protein synthesis and nitrogenous compounds formation, such as NA and DMA, both of which participate in Zn and Fe transport in rice (Slamet-Loedin *et al.*, 2015). So, application of N fertilizer could improve Fe and Zn in rice grains but effect varies depending on genotypic different and rate or method of application. Split application of nitrogen fertilizer in proper time corresponding to plant requirement found to be

effective and help to increase Fe content of rice grain and enhance rice grain nutritional value (Fei *et al.*, 2008). N fertilizer rate combined with Zn application method show a clear increase in both grain yield and Zn content as the N fertilizer level increased from 200 to 300 kg/ha. Fe and Zn content in different parts of rice plant may be affected by nitrogen fertilizer thus increasing the nitrogen fertilizer up to 160kg/ha has reported to improve Fe and Zn concentrations in brown rice by 28.96%, and 16.0% for IR64 and by 22.16% and 20.21% for IR68144 compared with control (Hao *et al.*, 2007).

An estimation of soil Zn and application of Zn fertilizer to Zn deficit soil is important for Zn biofortification (Mallikarjuna Swamy *et al.*, 2016). But the response to Zn fertilizer has been shown to differ across rice genotypes, methods of application and soil conditions (White *et al.*, 2011). Foliar application of Zn fertilizers has shown better results than soil application for increasing grain Zn concentration, but the magnitude of this increase is not consistent across genotypes (Table 2) (Mabesa *et al.*, 2013). Application of Fe fertilizer is direct and effective method for enhancing Fe content in rice grain (Li *et al.*, 2016). Among the various iron forms chelated iron sulphates results in higher root iron concentrations while a higher leaf iron concentration is observed when iron citrate is used. Effects of foliar application of different forms of iron fertilizer at different plant developmental stages are studied in rice and it is shown that application of the synthetic chelating agents like DTPA-Fe form at the anthesis stage results in about 20% increase in iron content of polished rice grains (He *et al.*, 2013). In addition to grain iron concentration, iron fertilization positively influences the grain zinc concentration in rice and wheat (Zeidan *et al.*, 2010, Zaigham *et al.*, 2014)

### **Water management**

Rice is a semi aquatic crop grown under lowland condition but as the fresh water crisis increasing day by day, rice is now grown under

various irrigation management options like always aerobic, always anaerobic and many variations along the aerobic-anaerobic spectrum (Bouman *et al.*, 2007). In aerobic conditions, rice is grown as a dry field-crop in irrigated not in flooded, fertile soils (Gao *et al.*, 2006). But shifting from anaerobic to aerobic condition has benefits and risking of micronutrient status of grains in different soil types which need to be understand. In aerobic conditions nitrogen is uptake as nitrate which may cause an imbalance in the cation/anion ratio, resulting in exudation of OH<sup>-</sup> into the rhizosphere with a subsequent rise in soil pH and redox potential. A higher redox potential can accumulate much more oxidized Fe<sup>3+</sup> which is not readily available for plant uptake (Zuo *et al.*, 2011).

While in flooding condition, Fe- oxides are dissolved when the Fe<sup>3+</sup> is reduced to Fe<sup>2+</sup> which weakens the oxide stability and increases its water-solubility (Kirk, 2004).

This releases much more Fe into the soil solution which is nearly sufficient for plant uptake. In both aerobic and flooded condition, application of ferrous sulphate significantly increases shoot Fe concentration and shoot Fe content at tillering stage but at physiological maturity, grain iron is found significantly lower in aerobic than in flooded plots (Table 3) (Xiaoyun *et al.*, 2012).

Under anaerobic conditions, Zn forms as insoluble zinc sulphide (Bostick *et al.*, 2001) and insoluble carbonate mixtures (Kirk, 2004) which plant cannot uptake. While increase oxidation under aerobic condition decrease Zn precipitation as ZnS (Carbonell-Barrachina *et al.*, 2000) and further increase availability of iron oxidizing/reducing bacteria, AM fungi associated with root-induced rhizosphere processes such as exudation of Zn chelators and have positive effect on nutrient availability (Gao *et al.*, 2017).

Alternative wetting and drying (AWD) is one of the promising water saving technology which is widely adapted in many rice producing

countries (Lampayan *et al.*, 2015). It combines both the beneficial effects of aerobic and anaerobic cultivation system which potentially decrease water inputs by 5%–35% when compared with Continuous flooding (CF) with the yield of rice grain either being maintained (Chapagain *et al.*, 2010).

Although for iron, it does not seem to be promising for increasing iron content in grain (Nortona *et al.*, 2017) but shows effective for increment of grain zinc content alone or when combine with various zinc fertilizer treatments (Table 4) (Wang *et al.*, 2014).

### **Breeding and transgenics approach**

Plant breeding (e.g., genetic biofortification) approach is thought to be the cost effective and eco- friendly approach for improving micronutrient status of rice in developing countries. For developing variety with high micronutrient, germ plasm screening is done initially to find out the genetic variation among the existing genetic resource (Slamet-Loedin *et al.*, 2015, Howarth *et al.*, 2017). There is abundant genetic variation for the grain Zn and Fe concentration in both brown and polished grains in the rice germplasm. Different wild relatives, landraces, aus and aromatic accessions, deep water rice and coloured rice are the best sources of high grain Zn. Wild species of rice such as *O. nivara*, *O. rufipogon*, *O. latifolia*, *O. officinalis*, and *O. granulata* also contain high amounts of Zn, around 2–3 fold higher than in the cultivated rice with Zn concentration varying from 37 mg/kg to 55 mg/kg in non-polished grains (Impa *et al.*, 2013; Anuradha *et al.*, 2012; Banerjee *et al.*, 2010).

The world's first Zn enriched rice variety is released in 2013 by the Bangladesh Rice Research Institute (BRRI dhan62), which is claimed to contain 20–23mgZnkg<sup>-1</sup> for brown rice (Harvest plus, 2015) while another variety by Directorate of Rice Research (DRR-Dhan 45) is released in India with over all mean zinc content of 22.6ppm in polished rice, develop

through conventional breeding without compromising yield using the material from Harvest Plus (Balasubramanian, 2016). While in case of iron, rice germplasm has a very narrow genetic variability for endosperm iron content. Iron content changes depending on varieties, IR64 (12.58-12.88mg/Kg), Jasmine 85 (12.84-18.50 mg/Kg) and OMCS2000 (11.77-14.78 mg/Kg) and about 2/3 of iron is lost through milling (Tran *et al.*, 2004). Other advance strategy like mutation breeding also gaining importance in this regard. A number of IR64 mutants produced by the treatment with Sodium azide, a mutagen, is reported to have high Zn. Three IR64 mutant lines viz., M-IR-180, M-IR-49 and M-IR-175 has more than 26 mg kg<sup>-1</sup> Zn in polished rice as against 16 mg kg<sup>-1</sup> in IR64 has been reported (Jeng *et al.*, 2012) A combinatorial approach using both hybridization and induced mutation is also found to be effective to develop new cultivar expressing several improved traits like improve aroma and high iron content (Cua, 2016).

Although various approaches are trying from last 15 years to reach the 30% EAR (Estimated Average Requirement) nutritional targets for iron and zinc concentrations in polished rice grains (Bouis *et al.*, 2011) but still it remains a major challenge. This 30% EAR was calculated as 13 μg g<sup>-1</sup> Fe and 28 μg g<sup>-1</sup> Zn in polished grains taking into account of 90% micronutrient retention after processing and 10% bioavailability for Fe and 25% bioavailability for Zn (Trijatmiko *et al.*, 2016). In this aspect transgenic approach can be a better option.

Several studies exhibit the associated increase in Fe and Zn content in rice grain by over expression or activation of various transporters genes. Over expression of three rice NAS homologous proteins, (OsNAS1, OsNAS2, and OsNAS3) resulted in 2-fold increase in Fe and Zn concentration in polished rice (Sasaki *et al.*, 2014) while over expression of *OsHMA3* enhance the uptake of Zn by up regulating the ZIP family genes in the roots (Johnson *et al.*, 2011).

Combined improvement of iron, zinc and  $\beta$ -carotene content in rice endosperm are improve by expressing Arabidopsis *Nicotianamine Synthase 1* (*Atnas1*), Bean *Ferritin* (*PvFERRITIN*), bacterial *Carotene Desaturase* (*CRTI*) and maize *PHYTOENE Synthase* (*ZmPSY*) in a single genetic locus (Singh *et al.*, 2017).

High yielding rice line with Zn and Fe biofortified in polished grains can also be develop by overexpressing *OsNAS2* in various genotypes (Table 5) (Singh *et al.*, 2017) Further field evaluation of transgenic events is also reported to be successful without a yield penalty or altered grain quality where *NASFer-274* containing rice (*OsNAS2*) and soybean ferritin (*SferH-1*) genes is use in a single locus insertion (Cua, 2016).

Iron and zinc deficiency are the most common type of micronutrient malnutrition where population of all groups in all the region of world is get affected. So, for effective and sustainable solution of this problem a complete understanding of iron and zinc uptake, translocation and further allocation to reproductive organs is needed. Agronomic interventions for increment of micronutrient status are effective but it is erratic, depends on cultivar and environment.

Genetic intervention is a cost effective and sustainable strategy but for that further exploitation of wide genetic variety of rice germplasm is necessary. Consequently, new combined agronomic and genetic strategy should be developed to address this problem of malnutrition for people whose staple diet is rice.

## References

Abdullah, A.S., 2015. Zinc Availability and Dynamics in the Transition from Flooded to Aerobic Rice Cultivation. *J Plant Biol Soil Health*. 2(1): 2-5.

Anuradha, K., Agarwal, S., Batchu, A.K., Babu, A.P., Swamy, B.P.M., Longva, T., Sarla,

N. 2012. Evaluating rice germplasm for iron and zinc concentration in brown rice and seed dimensions. *J Geophys Res*. 4:19–25.

Balasubramanian.2016. Biofortification: Micronutrient built in grains. The Hindu <http://www.thehindu.com/sci-tech/science/Biofortification-Micronutrient-built-in-grains/article14572744.ece>. Accessed 16 April 2016

Banerjee, S., Sharma, D.J., Verulkar, S.B., Chandel, G. 2010. Use of in silico and semi quantitative RT-PCR approaches to develop nutrient rich rice (*Oryza sativa* L) India. *J Biotechnol*. 9:203–212.

Bostick, B.C., Hansel, C.M., La Force, M.J., Fendorf, S. 2001. Seasonal fluctuations in zinc speciation within a contaminated wetland. *Environ Sci Technol*. 35: 3823–3829.

Bouis, H.E., Hotz, C., McClafferty, B., Meenakshi, J.V., and Pfeiffer, W.H. 2011. Biofortification: a new tool to reduce micronutrient malnutrition. *Food Nutr Bull*. 32: 31–40.

Bouman, B., Barker, R., Humphreys, E., Tuong, T. P., Atlin, G., Bennett *J et al.*, 2007. Rice: Feeding the billion. In *Water for Food, Water for Life: a Comprehensive Assessment of Water Management in Agriculture* (Eds). Molden D editor (London: Earthscan and Colombo: International Water Management Institute). Pp. 515–549.

Cakmak, I., Pfeiffer, W.H., Mc Clafferty, B. 2010. Biofortification of durum wheat with zinc and iron. *Cereal Chem*. 87: 10–20.

Carbonell-Barrachina, A. A., Jugsujinda, A., Burlo, F., Delaune, R. D., Patrick Jr, W.H. 2000. Arsenic chemistry in municipal sewage sludge as affected by redox potential and pH. *Water Res*. 34:216–224.

Chapagain, T., Yamaji, E. 2010. The effects of irrigation method, age of seedling and spacing on crop performance, productivity and water-wise rice

- production in Japan. *M. Paddy Water Environ.* 8(1): 81-90.
- Chasapis, C.T., Loutsidou, A.C., Spiniopoulou, C.A., Stefanidou, M.E. 2012. Zinc and human health: an update. *Archi. toxicol.* 86:521-534.
- Cua, Q.H. 2016. *Biotechnologies for Plant Mutation Breeding.* Springer International.
- Da Silva, J.G. 2013. Food losses means hunger. The think. Eat. Save. Reduce your footprint-campaign of the save food initiative is a partnership between UNEP, FAO and Messe Dusseldorf. <http://www.unep.org/ourplanet/2013/may/en/pdf/article3.pdf>. Accessed 8 November 2013
- Das, S.K., 2014. Role of micronutrient in rice cultivation and management strategy in organic agriculture—A Reappraisal. *Agricultural Sciences.* 5: 765-769.
- Disante, K.B., Fuentes, D., Cortina, J. 2010. Response to drought of Zn-stressed *Quercus suber* L. Seedlings. *Environ Expr Bot.* 70:96-103.
- El-Jendoubi, H., Vazquez, S., Calatayud, A., Vavpetic, P., Vogel-Mikus, K., Pelicon, P., Abadía, J., Abadía, A., Morales, F. 2014. The effects of foliar fertilization with iron sulfate in chlorotic leaves are limited to the treated area. A study with peach trees (*Prunus persica* L. Batsch) grown in the field and sugar beet (*Beta vulgaris* L.) grown in hydroponics. *Front Plant Sci.* 5: 1-16.
- FAO. 2017. Rice market monitor. [http://www.fao.org/fileadmin/templates/est/COMM\\_MARKETS\\_MONITORING/Rice/Images/RMM/RMMAPR17H.pdf](http://www.fao.org/fileadmin/templates/est/COMM_MARKETS_MONITORING/Rice/Images/RMM/RMMAPR17H.pdf). Accessed 11 April. 2017
- Fei, X., Zhong, W., Yun-jie, G., Gang, C., Peng, Z. 2008. Effects of nitrogen application time on caryopsis development and grain quality variety Yangdao6. *Rice Science.* 15(1): 57-62.
- Freitas, B.A., Lima, L.M., Moreira, M.E., Priore, S.E., Henriques, B.D., Carlos, C.F., Sabino, J.S., Franceschini, Sdo. C. 2016. Micronutrient supplementation adherence and influence on the prevalence of anemia and iron, zinc and vitamin A deficiencies in preemies with a corrected age of six months. *Clinics.* 71(8):440-448.
- Gao, X., Chunqin, Z., Xiaoyun, F., Fusuo, Z., and Ellis, H. 2006. From flooded to aerobic conditions in rice cultivation: Consequences for zinc uptake. *Plant and Soil.* 280:41-47.
- Gao, X., Kuyper, T.W., Zou, C., Zhang, F., Hoffland, E. (2007). Mycorrhizal responsiveness of aerobic rice genotypes is negatively correlated with their zinc uptake when nonmycorrhizal. *Plant and Soil.* 290: 283-291.
- Hao, H., Wei, Y., Ying, X., Wu, F. 2007. Effects of Different Nitrogen Fertilizer Levels on Fe, Mn, Cu and Zn Concentrations in Shoot and Grain Quality in Rice (*Oryza sativa*). *Rice Science.* 14(4): 289-294.
- Harvestplus2015. Bangladesh releases new, improved zinc rice variety. Harvest Plus, Washington, DC (2015)
- He, W., Shohag, M.J., Wei, Y., Feng, Y., Yang, X. 2013. Iron concentration, bioavailability, and nutritional quality of polished rice affected by different forms of foliar iron fertilizer. *Food Chem.* 141:4122-4126.
- Howarth, E.B., Amy, S. 2017. Improving nutrition through biofortification: A review of evidence from Harvest Plus, 2003 through 2016. *Global Food Security* 12: 49-58.
- Impa, S.M., Morete, M.J., Ismail, A.M., Schulin, R., Johnson-Beebout, S.E. 2013. Zn uptake translocation and grain Zn loading in rice (*Oryza sativa* L) genotypes selected for Zn-deficiency tolerance and high grain Zn. *J Exp Bot.* 64:2739-2751.
- Jeng, T.L., Lin, Y.W., Wang, C.S., Sung, J.M. 2012. Comparisons and selection of rice mutants with high iron and zinc contents in their polished grains that were mutated from the *indica* type cultivar IR64. *J Food Compos Anal.* 28:149-154.
- Jia-Yang, L., Jun, W., Robert, S.Z. 2014. The 3,000 rice genomes project: new



- opportunities and challenges for future rice research. *Giga Science*. 3(8):1-3.
- Johnson, A. A., Kyriacou, B., Callahan, D.L., Carruthers, L., Stangoulis, J., Lombi, E., Tester, M. 2011. Constitutive overexpression of the OsNAS gene family reveals single-gene strategies for effective iron-and zinc-biofortification of rice endosperm. *PLoS ONE*. 6(24476): 1-11.
- Kirk, G., 2004. *The Biogeochemistry of Submerged Soils*. Chichester: Wiley.
- Lampayan, R.M., Rejesus, R.M., Singleton, G.R., Bouman, B.A.M. 2015. Adoption and economics of alternate wetting and drying water management for irrigated lowland rice. *Field Crop Res*. 170: 95–108.
- Lei, G., Jiadong, C., Ruijie, C., Hubo, L., Hongfei, L., Longxing, T., and Jie, X. 2016. Comparison on cellular mechanisms of iron and cadmium accumulation in rice: prospects for cultivating Fe-rich but Cd-free rice. *Rice*. 9(39): 1-12.
- Mabesa, R.L., Impa, S.M., Grewal, D., Johnson-Beebout. 2013. Contrasting grain zinc response of biofortification rice (*Oryza sativa* L) breeding lines to foliar Zn application. *Field Crop Res*. 2(149):223-233.
- Mallikarjuna Swamy, B.P.M., Rahman, M., Inabangan-Asilo, M.A., Amparado, A., Manito, C., Mohanty, P.C., Reinke, R., Slamet-Loedin, I.H. 2016. Advances in breeding for high grain Zinc in Rice. *Rice*. 9(49): 1-16.
- Masuda, H., Aung, M.S., Nishizawa, N.K. 2013. In biofortification of rice using different transgenic approaches. *Rice*. 6(40): 1-12.
- Murgia, I., and Morandini, P. 2017. Iron deficiency prolongs seed dormancy in *Arabidopsis* Plants. *Front Plant Sci*. 8: 1-5.
- Nakandalage, N., Nicola, S.M., Norton, R.M., Hirotsu, N., Milham, P.J., Seneweera, S. 2016. Improving Rice Zinc Biofortification Success Rates Through Genetic and Crop Management Approaches in a Changing Environment. *Frontiers in Plant Science*. 7(764):1-13.
- Nortona, G.J., Shafaei, M., Travis, A.J., Deacona, C.M., Dankua, J., Ponda, D., Cochranea, N., Lockhart, K *et al.*, 2017. Impact of alternate wetting and drying on rice physiology, grain production, and grain quality. *Field Crops Research*. 205: 1–13.
- Prasad, A.S., 2013. Discovery of Human Zinc Deficiency: Its impact on human health and diseases. *Adv Nutr*. 4(2): 176-190.
- Roohani, N., Hurrell, R., Kelishadi, R., Schulin, R. 2013. Zinc and its importance for human health: An integrative review. *J Res Med Sci*. 18:144-157.
- Rutter, G.A., Chabosseau, P., Bellomo, E.A., Maret, W., Mitchell, R.K., Hodson, D.A., Solomou, A., and Hu, M. 2016. Intracellular zinc in insulin secretion and action: a determinant of diabetes risk? *Proceed Nutrl Soc.*, 75: 61-72.
- Samreen, T., Hamid, H., Saleem, U., Muhammad, U., Javid. 2017. Zinc effect on growth rate, chlorophyll, protein and mineral contents of hydroponically grown mungbeans plant (*Vigna radiata*). *Arabian J Chem*. 10: 1802–1807.
- Sarwar, N., Saifullah., Malhi, S.S., Zia, M.H., Naeem, A., Bibi. S., Farid, G. 2010. Role of mineral nutrition in minimizing cadmium accumulation by plants. *J Sci Food Agric*. 90(6):925–937.
- Sasaki, A., Yamaji, N., Ma, J.F. 2014. Over expression of *OsHMA3* enhances Cd tolerance and expression of Zn transporter genes in rice. *J Exp Bot*. 65:6013–6021.
- Sharma, A., Patni, B., Shankhdhar, D., and Shankhdha, S.C. 2013. Zinc-An indispensable micronutrient. *Physiol. Mol. Biol, Plants*. 19(1): 11–20.
- Singh, S.P., GUISSEM, W., and Bhullar, N.K. 2017. Single genetic locus improvement of iron, zinc and  $\beta$ -carotene content in rice grains. *Scientific Reports*. 7:1-11.
- Slamet-Loedin, I.H., Johnson-Beebout, S.E., Impa, S., Tsakirpaloglou, N. 2015. Enriching rice with Zn and Fe while

- minimizing Cd risk. *Front Plant Sci.* 6(121): 1-9.
- Tran, T., Hoa, C., Nguyen, T., Phong, L. 2004. Effect of milling technology on iron content in rice grains of some leading varieties in the Mekong Delta. *Omonrice.* 12:38–44.
- Trijatmiko, K.R., Duenas, C., Tsakirpaloglou, N., Torrizo, L., Arines, FM., Adeva, C., Balindong, J., Oliva, N., Sapasap, M.V., Borrero, J *et al.*, 2016. Biofortified *indica* rice attains iron and zinc nutrition dietary targets in the field. *Sci Rep.* 6 (19792): 1-13.
- Wang, Y., Wei, Y., Dong, Y., Lu, L., Feng, Y., Zhang, J., Pan, F., Yang, X. 2014. Improved yield and Zn accumulation for rice grain by Zn fertilization and optimized water management. *Biomed & Biotechnol.* 15(4): 365-374.
- Ward, R.J., Zucca, F.A., Duyn, J.H., Crichton, R.R., Zecca, L. 2014. The role of iron in brain ageing and neurodegenerative disorders. *Lancet Neurol.* 13(10):1045-1060.
- Washington, DC: National Academy Press; 2001. IOM. Institute of Medicine. Iron. In: Dietary Reference Intakes for Vitamin A, Vitamin K, Arsenic, Boron, Chromium, Copper, Iodine, iron, Manganese, Molybdenum, Nickel, Silicon, Vanadium, and Zinc., pp. 290–393.
- White, P.J., Braodley, M.R. 2011. Physiological limits to zinc biofortification of edible crops. *Front Plant Sci.* 2(80):1-11.
- WHO (2016). Iron deficiency anaemia. <http://www.who.int/nutrition/topics/ida/en> . Accessed 29 August 2016.
- Xiaoyun, F., Md. Rezaul, K., Xinping, C., Yueqiang, Z., Xiaopeng, G., Fusuo, Z., Chunqin, Z. 2012. Growth and iron uptake of lowland and aerobic rice genotypes under flooded and aerobic cultivation. *Commun Soil Sci Plant Anal.* 43:1811–1822.
- Zaigham, S., Hatem, R., Allah, R. 2014. Combating Mineral Malnutrition through Iron and Zinc Biofortification of Cereals. *Compr Rev Food Sci F.* 13(3)329-347.
- Zargar, S.M., Mahajan, R., Farhat, S., Nazir, M., Mir, R.A., Nazir, M., Salgotra, R.A., Mallick, S.A. 2015. Understanding the role of Iron and Zinc in animals and crop plants from genomics perspective. *Current trends biotechnology pharmacy.* 9(2) 182-195.
- Zeidan, M.S., Mohamed, M.F., Hamouda, H.A. 2010. Effect of foliar fertilization of Fe, Mn and Zn on wheat yield and quality in low sandy soils fertility. *World J Agric Sci.* 6:696–699.
- Zuo, Y., Zhang, F. 2011. Soil and crop management strategies to prevent iron deficiency in crops. *Plant Soil.* 339 83–95.

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