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Synthesis and Evaluation of Gyps vulture Multidrug Resistant Protein 4 (MRP4/ABCC4) Specific Multiple Antigenic Peptide (MAP) for its Immunoreactivity

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ABSTRACT

Keywords

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Efforts are in progress to conserve critically endangered *Gyps* spp. vultures by identifying and removing the vulture-unsafe drugs from the food chain. Several drugs namely diclofenac, aceclofenac, ketoprofen and nimesulide are found to cause hyperuricemia and fatal nephrotoxicity in *Gyps* vultures. Research on development of an *in vitro* model for screening drugs in practice and the new molecules for their hyperuricemic potential can facilitate the conservation efforts. Multi drug resistant protein 4 (MRP4/ABCC4) is an efflux transporter on the luminal side of proximal renal tubular membrane that plays key role in urate as well as drug transport. In the present study, we report development and use of *Gyps himalayensis* MRP4 specific multiple antigenic peptide (MAP). Sequences of linear peptide and N terminal immunodominant region of *Gyps himalayensis* MRP4 was predicted *in silico*. Tetrameric MAP was synthesized in solid phase synthesis (SPPS) using Wang resin and employed for immunization of rabbit for obtaining polyclonal serum. N terminal region (1-737 amino acids) of vulture MRP4 predicted to contain immunodominant epitopes was expressed in *E. coli* BL21 cells and purified in Ni-NTA affinity chromatography. The ~24 kDa purified protein could be detected in immunoblots using anti-MAP serum indicating its potential in detection of MRP4 expression. This hyperimmune serum can be exploited for its use in detection of MRP4 expression in avian renal tubular cells or MRP4 expressing cells before their use for screening drugs for their role in urate transport.

Introduction

In South Asian subcontinent, three resident *Gyps* species vultures, *Gyps indicus*, *Gyps bengalensis* and *Gyps tenuirostris* are 'Critically endangered' and migratory species *Gyps himalayensis* moved from 'Least concern'

to 'Near threatened' category (<https://www.iucnredlist.org/search?taxonomies=22675030&searchType=species>)(Prakash *et al.*, 2017). *Gyps* vultures got exposed to the non-steroidal anti-inflammatory drug diclofenac through food chain and experienced fatal nephrotoxicity that proved to be the major

cause of catastrophic decline in their population (Cuthbert *et al.*, 2014, 2015). Since the detection diclofenac as the culprit drug in dead birds (Oaks *et al.*, 2004; Green *et al.*, 2004) as well as experimentally treated vultures (Swan *et al.*, 2006a and b), several other drugs like ketoprofen (Naidoo *et al.*, 2017), fluxiclin (Zorrilla *et al.*, 2015), aceclofenac (Galligan *et al.*, 2016), nimesulide (Cuthbert *et al.*, 2015) are reported to produce hyperuricemia and toxicity akin to that of diclofenac in vultures. Meloxicam being the only proven vulture-safe molecule (Swan *et al.*, 2006a; Swarup *et al.*, 2007) and increasing evidence of many molecules being detected to be nephrotoxic in *Gyps* vultures emphasizes the need for toxicity testing of veterinary drugs in practice and the new molecules in order to ensure some vulture-safe drugs. However, testing all molecules *in vivo* is not practical due to limited number of these critically endangered species (Prakash *et al.*, 2017). Furthermore, *Gyps* vultures are highly sensitive to drugs like diclofenac as compared to other avian species like chicken, crow and turkey vulture (Rattner *et al.*, 2008; Naidoo *et al.*, 2011); the possibility of screening drugs in surrogate species is also ruled out.

Diclofenac toxicity in *Gyps* vulture has been shown to be associated with decreased uric acid excretion (Naidoo and Swan, 2009). Urate transport across proximal renal tubular cells is considered to be unidirectional in birds that results from coordinated efforts of organic anion transporter 1/3 (OAT1/OAT3) on basolateral side and multidrug resistant protein 4 (MRP4/ABCC4) on the luminal side (Dudas *et al.*, 2005; van Aubel *et al.*, 2005; Batallie *et al.*, 2008).

The core structure of MRP4, a typical ABC transporter comprises of two membrane-spanning domains, each consisting of six trans-membrane helices with two cytosolic ATP-binding domains (Borst *et al.*, 2007).

Cell-culture based screening can provide an alternative approach for testing the molecules for nephrotoxic potential. HEK293 and SF9 cells expressing human MRP4 for studying the effect of drugs on urate transport have been employed (van Aubel *et al.*, 2005; El Sheikh *et al.*, 2008). Likewise, use of cells expressing vulture MRP4 for screening of drugs for their urate transport inhibitory potential can be explored, for which means to characterize MRP4 expression needs to be established. In an earlier study (Batallie *et al.*, 2008), MRP4 protein in chicken proximal renal tubular cells was detected by using a custom affinity-purified polyclonal antibody designed against peptide C-N-G-Q-L-A-T-D-S-S-L-D-P-S-S localized in a region near C-terminal end of the chicken MRP4 sequence. We have observed that *ABCC4/MRP4* is transcribed as two transcript variants in chicken and vulture and both of them produce functional protein (Barik, 2016). These variants are likely to produce proteins differing in 19 amino acids at C-terminal end, thus the antibody serving as a tool for MRP4 detection should be able to identify both the transcript variants. Multiple antigenic peptides (MAPs) provide potent antigen to produce mono-specific serum with high titre (Dyson *et al.*, 1988; Tam, 1988). Here, we report designing, synthesis and evaluation of N-terminal specific multiple antigenic peptide specific polyclonal serum for immune-detection of vulture MRP4.

Materials and Methods

***In silico* analysis of the N-terminal region of MRP4 of *Gyps himalayensis* to determine the immunodominant and signal peptide containing region**

N-terminal region (1-737 amino acids) of predicted MRP4 of *Gyps himalayensis* (Accession KX168697) was analysed using online tools from IEDB Analysis Resource

(<http://tools.iedb.org/bcell/>) for linear epitope prediction (Larsen *et al.*, 2006; Jespersen *et al.*, 2017) and hydrophilicity (Parker *et al.*, 1986). The topology of the protein was checked by using online platform from Stockholm Bioinformatics Centre (<http://phobius.sbc.su.se/index.html>) using the phobius algorithm (Käll *et al.*, 2007).

Synthesis and purification of multiple antigenic peptide (MAP)

Preparation of Fmoc-Cys-Wang

100 mg Wang resin was dissolved in 3 ml of Dimethylformamide (DMF) and poured into a clean Merrifield flask. In a separate 5 ml tube, 234.20 mg Fmoc-cysteine was dissolved in 3 ml DMF and 34 μ l of N,N'-Diisopropyl carbodiimide (DIPC) was added to the solution and incubated at 4°C for 20 min. After incubation, the entire solution containing cysteine was poured into the Merrifield flask after draining out the excess DMF and then 0.1 equivalent of catalytic p-dimethyl aminopyridine (DMAP) was added to it. The flask was properly sealed and kept in constant shaking (200 RPM) at 4°C for overnight. The product so obtained is Fmoc-Cys-Wang.

Coupling of Di-Fmoc-Lys to NH₂-Cys-Wang

The excess DMF was drained out and the flask was washed twice with DMF, di-chloro methane (DCM) and DMF, respectively. Then capping of free functional groups on Wang resin was done by incubating the resin in 2 ml acetylation mixture [DMF: Acetic anhydride: DIEA (193: 6: 1, v/v)] at 4°C and constant shaking (200 rpm) for 20 min. After incubation, excess solution was drained out and the flask was washed with DMF and DCM as above. Then, 4 ml of 20 % piperidine in DMF (V/V) was added to the flask and again incubated at 4°C and constant shaking (200 rpm) for 20 min. The flask washing with

DMF and DCM was performed as mentioned above. In an Eppendorf tube, 118.14 mg of Di-Fmoc-Lys, 27.06 mg of 1-Hydroxybenzotriazole (HOBt) and 72.06 mg of 2-(1H-Benzotriazole-1-yl)-oxo-1,2,3,3-tetramethyluronium hexa-fluorophosphate (HBTU) was weighed and dissolved in DMF and stored at 4°C for 20 min. Then 150 μ l of N,N-Diisopropyl ethylamine (DIEA) was added to the tube. The content of the entire tube was now emptied into the Merrifield flask and incubated for 2 h at 4°C for coupling. Thereafter, again the acetylation mixture was prepared and added to the flask after emptying it and kept for constant shaking (200 rpm) at 4°C for 30 min. Thereafter, the flask was washed thrice with DMF and then 4 ml of 20 % piperidine in DMF was added to the flask and incubated at 4°C for 20 min in constant shaking. The resultant product in this step is Di-Fmoc-Lys bound to NH₂-Cys-Wang.

Preparation of branched Lysine four arm of the MAP core

In the constructed MAP, branching was achieved to form four arm structure using di-Fmoc-Lys-OH to provide lysine core over which chosen peptide sequence was synthesized using Fmoc(N α -9-Fluorenylmethyloxycarbonyl) solid-phase peptide synthesis (Fig. 2).

The flask was washed twice with DMF, di-chloro methane (DCM) and DMF, respectively. In an Eppendorf tube 236.28 mg of di-Fmoc-Lys, 54.12 mg of HOBt and 147.29 mg of HBTU was weighed and dissolved in DMF and stored at 4°C for 20 min. Further steps were carried out as described above to yield the formation of four dendritic arms of lysine bound to di-Fmoc-Lys-NH₂-Cys-Wang.

Coupling of MAP

Each successive Fmoc-amino acid to be used

in peptide synthesis was activated by HOBt and HBTU and coupled in the presence of DIEA one by one until the desired length of the multimeric peptide was obtained.

Freeing MAP from resin

Final chemical deprotection and cleavage was done by treating resin bound peptides with the cleavage mixture [Trifluoroacetic acid: m-cresol: Thioanisole: Ethanedithiol: Water (82.5: 5: 5: 5: 2.5, v/v)]. Lastly, cleaved MAP was precipitated with chilled (dry) diethyl ether, vacuum dried and stored under dry conditions until further used for immunisation.

Generation of hyperimmune serum in rabbit against MAP

After obtaining due permission from Institute Animal Ethics Committee, one four months old New Zealand white female rabbit weighing 3.0 kg was procured from Laboratory Animal Resources, ICAR-Indian Veterinary Research Institute, Bareilly, Uttar Pradesh, India. All the guidelines of CPCSEA were strictly followed to minimize stress and discomfort and the animal was monitored daily for any disease condition. Pre-immunization serum was collected aseptically from the marginal ear vein of rabbit. Emulsion consisting of an equal volume of MAP (250 µg of antigen) and Freund's incomplete adjuvant (FIA) (Santa Cruz) was formulated. The rabbit was immunized with the emulsion prepared parenterally by intramuscular route at 2 sites (0.5 ml/site).

Subsequent boosters were administered at 7th, 14th and 28th days post-first immunization using an emulsion made of an equal volume of MAP (100 µg of antigen) and FIA. Finally, 10 ml of blood was collected from ear vein on the 35th day post-first immunisation. The serum after extraction from blood was stored at -20°C till further use.

Construction of recombinant plasmid (pRSET-rNvMRP4) containing coding region for short N-terminal stretch of MRP4

Based on *in silico* analysis, the stretch of amino acids from 533 to 698 residues of complete vulture MRP4 (Accession KX168697) was selected as the antigenic region (rNvMRP4) to be expressed in frame with 6X-Histidine tag of the vector backbone. Gene specific primer pair including Forward 5'-AAAGGATCCGGAGCTACACTGAGTG-3' and Reverse 5'-GAAGAA TTCTTTT CCCTCAGAACGACT-3' was designed to amplify respective 512 bp coding region using pCDNAV5/His-MRP4 (Barik, 2016) as template and synthesized at Eurofins, Bangalore, India. Forward and reverse primer incorporated *Bam* HI and *Eco* RI restriction sites, respectively to facilitate directional cloning in prokaryotic expression vector pRSET-A (Invitrogen).

PCR reaction consisted of both forward and reverse primers (50pmol) 2.5 µl each, 1:10 diluted pCDNAV5/His-MRP4 (Barik, 2016) as template 0.5 µl; and Phusion high fidelity master mix with GC buffer (New England Biolabs) 25 µl and final volume made up to 50 µl using nuclease free water. PCR reaction conditions were as follows: initial denaturation 98°C for 30 sec, followed by 30 repeated cycles of 98°C for 7 sec, 58°C for 30 sec, 72°C for 30 sec and final extension at 72°C for 10 min. The 512 bp amplicon was purified using PCR clean up kit (Qiagen) and double digested with *Bam* HI (New England Biolabs) and *Eco* RI (MBI Fermentas). The ligation mixture containing double digested PCR product (3 µl), pRSET A vector (1 µl), 10X ligation buffer 1 µl and T4 DNA ligase 1 µl (New England Biolabs) and the volume adjusted to 10 µl using nuclease free water was incubated at 4°C overnight. The ligation mixture was transformed to *E. coli* BL21 (Novagen) competent cells using the TSS

method (Chung *et al.*, 1989) and grown at 37°C overnight on LB agar plates containing chloramphenicol (34 µg/ml), ampicillin (100 µg/ml) and 1% glucose.

Expression, purification and characterization of recombinant rNvMRP4

These transformed cells were grown in LB broth supplemented with 1% glucose and antibiotics (100µg/ml ampicillin and 35µg/ml chloramphenicol) on shaker-cum-incubator at 37°C and 180 rpm. When the optical density of the culture at 600nm reached 0.6, isopropyl beta-D-thiogalactoside (Invitrogen) was added at to a final concentration of 1 mM followed by incubation at 37°C for four hours at 180 rpm to induce protein expression. The cells were pelleted and lysed in lysis buffer (6M Guanidium Hydrochloride, 100mM NaH₂PO₄, 10 mM Tris-Chloride, pH 8.0) by mild agitation for 1 hour at room temperature at 80 rpm. The lysate was centrifuged at 10000 rpm for 30 min at 4°C and the supernatant so obtained was incubated with Ni-NTA agarose (Qiagen) for 1 hour at room temperature with mild shaking at 80 rpm. After incubation, the entire slurry was loaded into the polypropylene column. The flow through was collected and the column was washed two times with wash buffer (8M Urea, 100 mM NaH₂PO₄, 10 mM Tris-Chloride, pH 6.3). The bound protein was then eluted by two buffers, one with pH 5.9 (8M Urea, 100mMNaH₂PO₄, 10 mM Tris-Chloride) and then with pH 4.5 (8M Urea, 100 mM NaH₂PO₄, 10 mM Tris-Chloride). All elutes, flow through and washes were collected and analysed in 15% Sodium Dodecyl Sulphate–Polyacrylamide Gel Electrophoresis (SDS-PAGE). The concentration of the expressed protein was determined by densitometry against known BSA standards as described by Pecora *et al.*, (2009). The identity of the recombinant protein (rNvMRP4) was confirmed by mass spectrometry. Expressed protein visualized as ~24 kDa band in SDS-PAGE was sliced. The

protein samples were trypsin digested and peptides extracted according to standard techniques. Peptides were analysed by a MALDI-TOF mass spectrometer at Sandor Life Sciences, Hyderabad, India. Spectra were analysed to identify the protein of interest using Mascot sequence matching software (Matrix Science).

Evaluation of MAP specific hyperimmune serum

The polyclonal antiserum against MAP raised in rabbit was tested for reactivity with recombinant MRP4 in dot blot assay and then confirmed in Western blot using semi-dry blot system (Towbin *et al.*, 1979). Briefly, recombinant rNvMRP4was subjected to 15% SDS-PAGE and protein bands were transferred to nitrocellulose membrane (Bio-Rad) using constant current (0.8 mA/cm² of the gel) for 90 min. The membrane was blocked in 3% BSA in phosphate buffer saline (PBS) at 37°C for 1 hour and then incubated with rabbit antiserum raised against vulture MRP4 specific MAP diluted in 0.8% PBS-Tween (1:500) for 1 hour at 37°C. After incubation, the membrane was washed and treated with anti-rabbit IgG HRPO conjugate (1:1000) for 1 hour and then the blot was developed with 3,3'-Diaminobenzidine (DAB) as substrate.

Results and Discussion

Present study reports the development of MAP and expression of partial recombinant MRP4 of Himalayan griffon vulture and also the evaluation of their use to detect the expression in immunoblots. The solid phase peptide synthesis discovered by Merrifield (1963) had unveiled new horizons for widespread applications of synthetic peptides as reagents and biologicals in chemical, biological and biomedical investigations. The short comings of some short peptides to elicit immune response due to their small sizes was soon

overcome by preparing copies of the epitopic peptide over lysine core matrix referred to as multiple antigenic peptide (Tam 1988). The 13 amino acids (646 to 658) long stretch (T-P-N-L-K-S-S-R-S-R-T-F-S) chosen for MAP construction. The tetrameric MAP was constructed on Wang resin by solid phase peptide synthesis (Fig. 2). The ultimate yield of MAP was 54.9 mg after final cleavage from the resin. This stretch was likely to serve as good B cell linear epitope eliciting better immune response that could detect recombinant N-terminal partial MRP4 as antigen.

Earlier, we characterized the complete coding sequence of vulture *MRP4/ABCC4* (KX168697) and also identified its two transcript variants of 4050 bp and 3993 bp mRNA sequences coding for polypeptides of 1349 and 1330 amino acids, respectively that differed in 19 amino acids at C-terminal end (Barik, 2016). The MRP4 sequence reported by Bataillie *et al.*, (2008) to be responsible for urate transport in *Gallus gallus* (Accession NM_001030819.1) lacked the stretch of 19 amino acids. The N-terminal amino acids 1-737 are translated in all transcript variants of vulture *MRP4/ABCC4*. Based upon *in silico* analysis, the stretch of the amino acids selected from 533 to 698 was predicted to be fairly immune-dominant, hydrophobic and lying in the cytoplasm not within the trans-membrane region (Fig. 1a, b and c).

Signal peptide predictions *in silico* also suggested that this selected region does not contain any cleavage site (Fig. 1c) indicating thereby that no portion will be lost as signal sequence during localization of the protein after expression. Thus, the stretch from 533 to 698 amino acids was expressed as recombinant N-terminal partial MRP4 (rNvMRP4). For amplification of 512 bp partial sequence from pCDNA5/His-MRP4 using expression primers, 58°C was found to

be the optimum annealing temperature. The PCR product was cloned in prokaryotic expression vector and the resulting recombinant plasmid pRSET-rNvMRP4 was characterized in PCR and restriction analysis (Fig. 3a and b). Visualization of 512 bp amplicon in PCR (Fig. 3a) and linearised 3412bp recombinant plasmid in RE digestion (Fig. 3b) confirmed the presence of insert. Presence of *Bam* HI site at the start of the insert and that of *Bgl* II site in the vector backbone, yielded a linear product of equal length upon digestion confirming the recombinant construct.

The SDS-PAGE analysis of the expressed rNvMRP4 revealed that ~24 kDa protein was eluted in pH 4.5 buffer (Fig. 4a). Upon MALDI-TOF analysis, the protein was found to align with *Gallus gallus* Multi Drug Receptor proteins (Fig. 4b). Besides MRP4, the peptides also matched with MRP1, MRP2 and P-Glycoprotein due to conserved nature of these transporters in their N-terminal regions (<https://www.ncbi.nlm.nih.gov/Structure/cdd/cddsrv.cgi?uid=330224>).

Densitometric analysis against known dilutions of BSA standards resolved in SDS-PAGE gels was preferred to quantify the recombinant protein as some minor bacterial proteins also get eluted in Ni-NTA column during purification.

The concentration of purified rNvMRP4 was found to be 1 µg/µl. The rabbit hyperimmune serum raised against this purified rNvMRP4 was evaluated in dot blot assay. Absence of reactivity in the pre-immune serum suggested the specific reactivity of the hyperimmune serum (Fig. 5a). Western blot (Fig. 5b) revealed that the hyperimmune serum was able to detect the expressed ~24 kDa N-terminal epitopic region of vulture MRP4/ABCC4.

Fig.1 *In silico* analysis of N-terminal region (1-737 amino acids) of predicted Multi Drug Resistant Protein 4 (MRP4) of *Gyps himalayensis* (Accession KX168697). (a) Linear epitope prediction. (b) Hydrophilicity plot. (c) Topology check and signal peptide cleavage site prediction

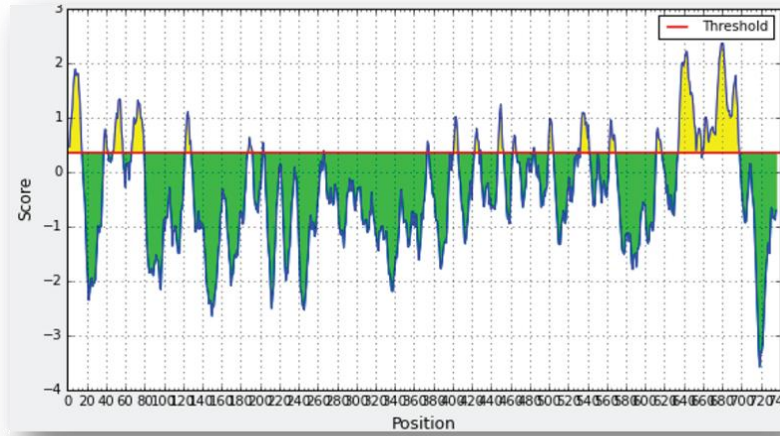


Fig. 1a

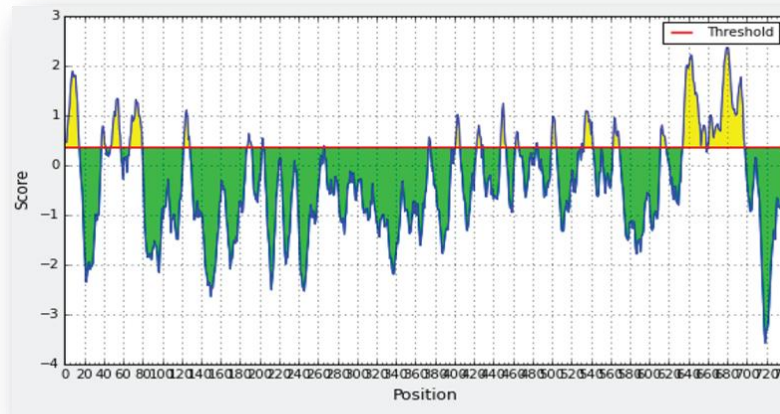


Fig. 1b

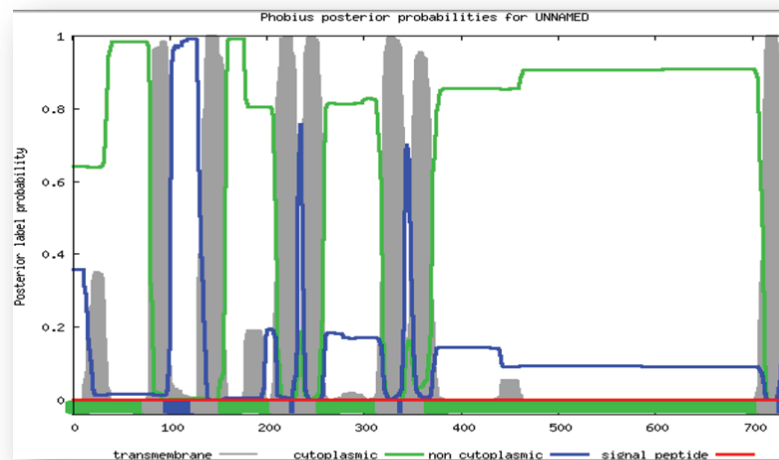


Fig. 1c

Fig.2 Schematic representation of the synthesized Multi Antigenic Peptide. Amino acids in box represent the MAP core bound to Wang resin shown in circle. The 13 amino acid peptide arms are marked in red [646-658 amino acids of N-terminal region of Multi Drug Resistant Protein 4 (MRP4) of *Gyps himalayensis* (Accession KX168697)]

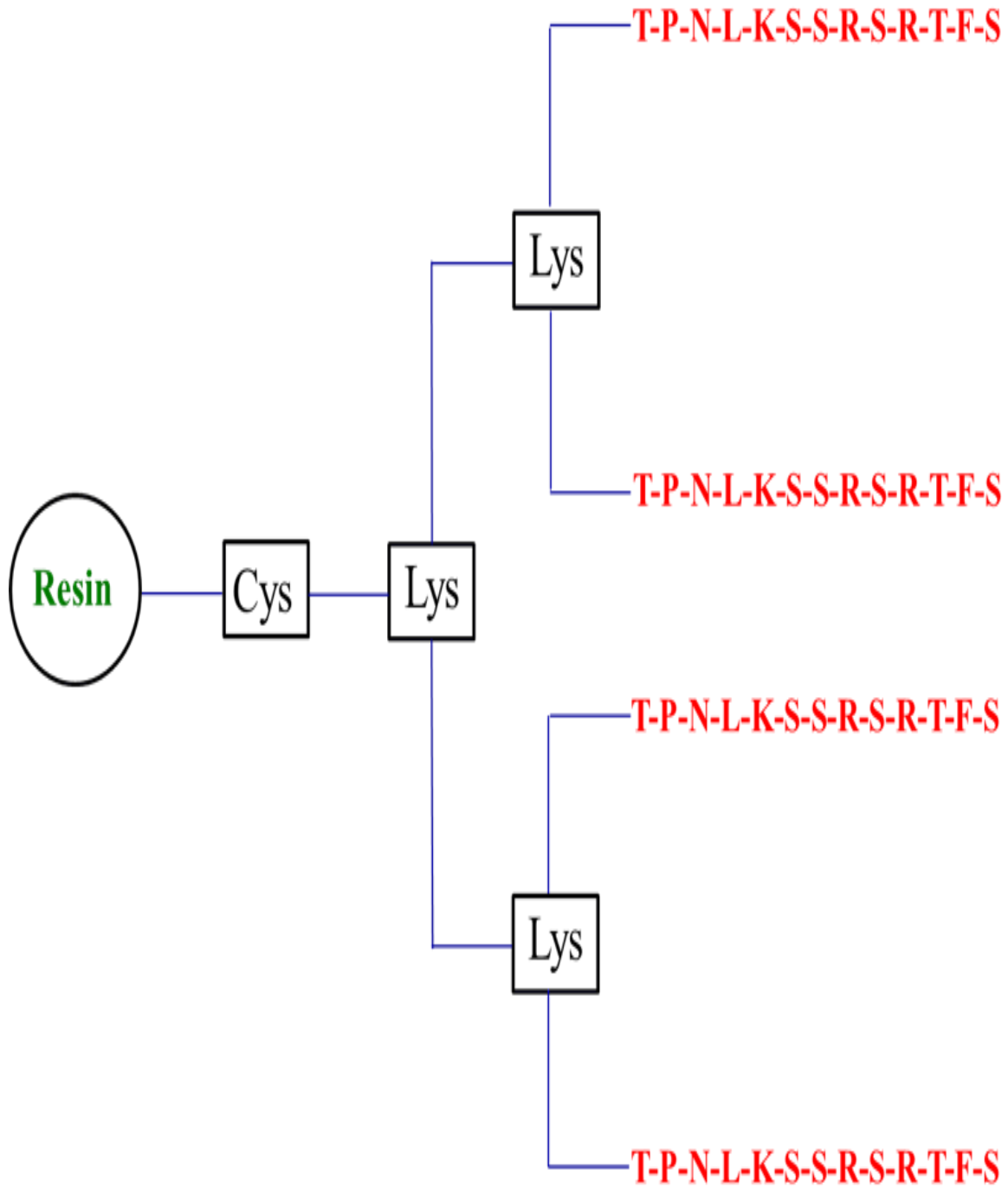


Fig.3 Characterization of recombinant plasmid (pRSET-rNvMRP4) containing partial coding region pertaining to N-terminal region of Multi Drug Resistant Protein 4 (MRP4) of *Gyps himalayensis*. (a) 1% agarose gel (1X TAE) electrophoresis of 500 bp amplicon in PCR using pRSET-rNvMRP4 as template [M: 100 bp plus ladder; Lane 1: pRSET-rNvMRP4]. (b) 1% agarose gel (1X TAE) electrophoresis of pRSET-rNvMRP4 digested with *Bam* HI (Lane 2) and *Bgl* II (Lane 3) against uncut pRSET-rNvMRP4 (Lane 1) [M: 1 kb plus ladder]

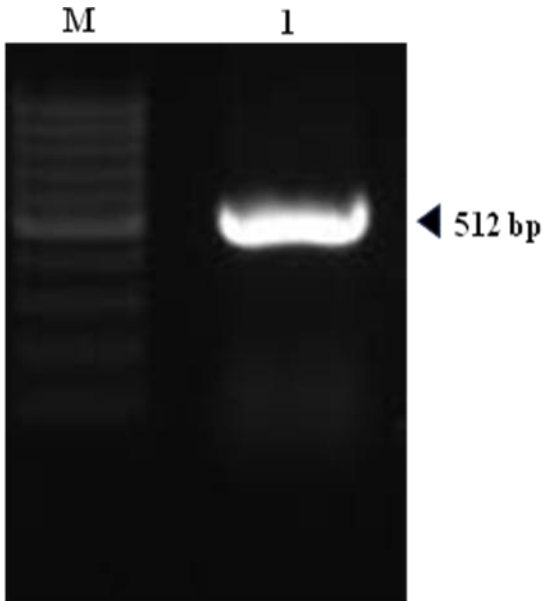


Fig.3a

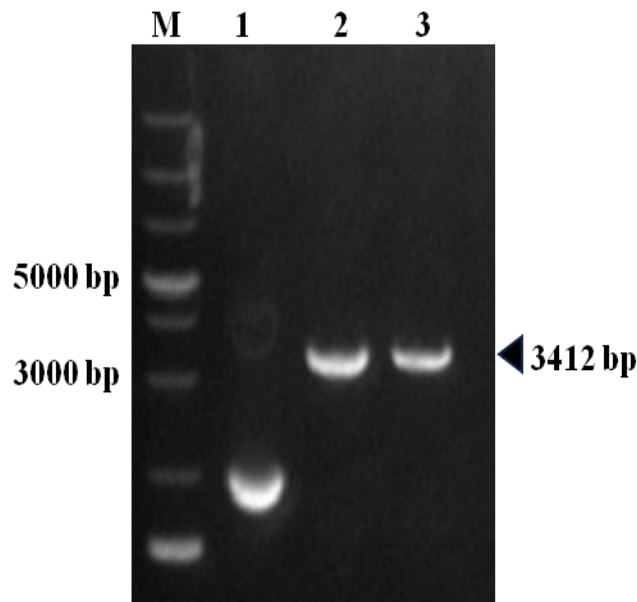


Fig. 3b

Fig.4 Purification and characterization of recombinant protein containing N-terminal region of Multi Drug Resistant Protein 4 (MRP4) of *Gyps himalayensis* (rNvMRP4) (a) 15% SDS-PAGE analysis of the expressed recombinant protein (rNvMRP4) in buffer with pH 4.5 (~24 kDa) using Ni-NTA affinity column chromatography [M: Protein molecular weight marker, L 1-9: Successive elutes in pH 4.5 buffer]. (b) MALDI-TOF analysis of the recombinant protein (rNvMRP4)

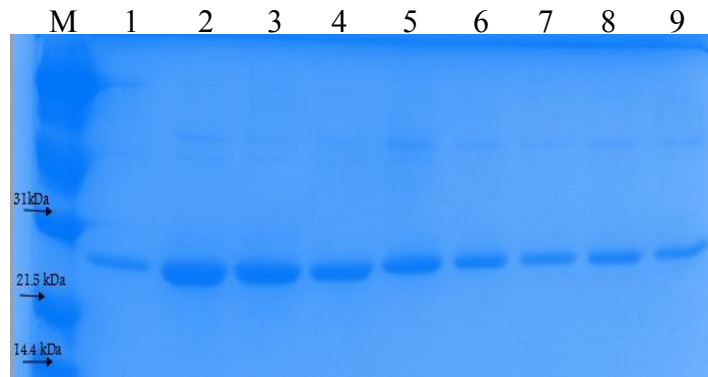


Fig. 4a

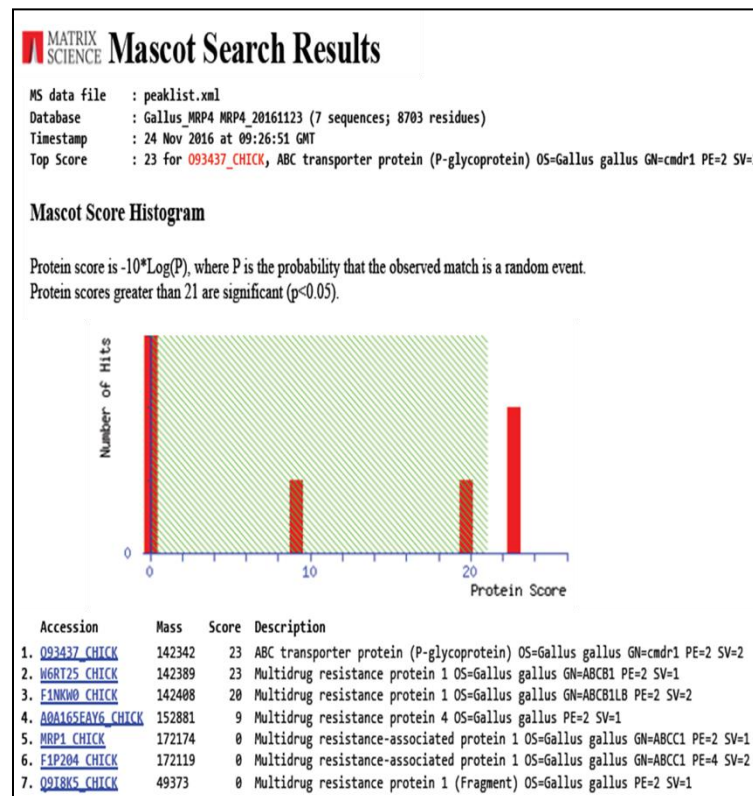


Fig. 4b

Fig.5 Immunoreactivity of the MAP specific hyperimmune serum with N-terminal region of Multi Drug Resistant Protein 4 (MRP4) of *Gyps himalayensis* (rNvMRP4). (a) Dot-blot assay to check the reactivity of the MAP specific hyperimmune serum [Pre: pre-immune serum; Post: pos-immune serum]. (b) Western blot indicating the specific reactivity of MAP specific hyperimmune serum with rNvMRP4 [M: protein marker; L: rNvMRP4]

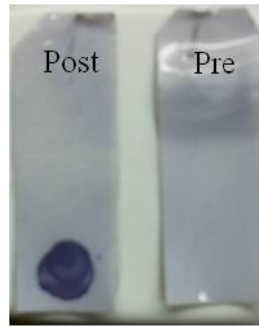


Fig. 5a

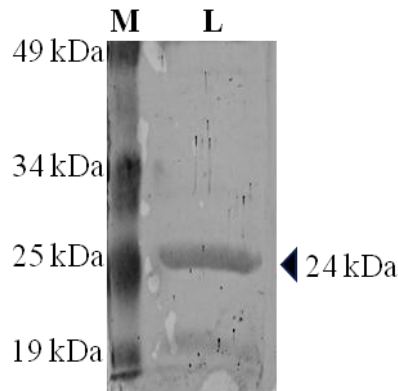


Fig. 5b

Thus, the MAP derived hyperimmune serum was found to be reactive and was successfully used to detect the recombinant linear epitope. Thus, the serum possesses all the required potential to screen for expression of MRP4 in *in vitro* models, though the detection of expression in avian renal tubular cells or MRP4 expressing cells have to be characterized in further studies.

Earlier, Battaille *et al.*, (2008) detected MRP4 protein in chicken proximal renal tubular cells

by using a custom affinity-purified polyclonal antibody designed against C-terminal specific peptide C-N-G-Q-L-A-T-D-S-S-L-D-P-S-S. Except for Cys at N terminal region, this sequence is likely to be present in both the translation products of transcript variants that may aid in detection of MRP4 expression. Short hairpin RNA interference (shRNAi) studies in primary monolayer of chicken proximal tubular cells proved that MRP4 is used for urate secretion in birds (Battaile *et al.*, 2008).

Being specific to N-terminal region, MAP developed in this study can invariably detect products translated from all known transcript variants of vulture *MRP4*. Nevertheless, ABC group of transporters have a conserved N-terminal domain hence the MAP derived hyperimmune serum can also be used for detection of expression of other transporters of ABC superfamily. Besides this, the *in silico* analysis reported here can further be used to design alternative MAPs or recombinant proteins for detection of the members this family of transporters.

In addition to urate, *MRP4* has the ability to transport, a range of endogenous molecules as well as exogenous drugs (Kanamitsu *et al.*, 2017; Murray *et al.*, 2017). Characterization of *MRP4* expression in renal cells/ expressing cell lines can help develop an *in vitro* model for studying transport of endogenous as well as exogenous molecules

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