

## Original Research Article

<https://doi.org/10.20546/ijcmas.2017.606.070>

## Induction of Enzyme Activities in Tuberose Plants Treated with Antagonists and Organic Fungicide under Artificial Inoculation of *Sclerotium rolfsii* Sacc.

G. Ragavi<sup>1</sup>, M.L. Mini<sup>2\*</sup>, I. Yesuraja<sup>3</sup> and K. Sethuraman<sup>4</sup>

<sup>1</sup>Department of Plant Pathology, <sup>2</sup>Department of Biotechnology, Agricultural College and Research Institute, Tamil Nadu Agricultural University, Madurai-625104, Tamil Nadu, India

<sup>3</sup>Horticulture Research Station, Thadiyankudisai, Dindigul - 624 212, Tamil Nadu, India

<sup>4</sup>Maize Research Station, Vagarai, Dindigul - 624 613, Tamil Nadu, India

\*Corresponding author

### ABSTRACT

#### Keywords

Antagonists,  
Peroxidase,  
Polyphenol oxidase,  
*Sclerotium rolfsii*,  
Tuberose.

#### Article Info

Accepted:  
04 May 2017  
Available Online:  
10 June 2017

A pot experiment was conducted to study the antioxidant responses of tuberose treated with biocontrol agents and organic fungicide in combating wilt disease caused by *Sclerotium rolfsii* Sacc. The antagonists, *Trichoderma viride*4 (Tv4), *Bacillus subtilis*1 (Bs1) and *Pseudomonas fluorescense*3 (Pf3) were used alone and in combinations. Mahua cake was used as the source of organic fungicide. The treatment with the commonly used chemical fungicide, hexaconazole was taken as check. Among the fifteen treatments, soil application of Tv4 @2.5 kg ha<sup>-1</sup> + Pf3 @2.5 kg ha<sup>-1</sup> + Bs1 @ 2.5 kg ha<sup>-1</sup> + Mahua cake @ 250 kg ha<sup>-1</sup> recorded the highest enzymatic activities of peroxidase, polyphenol oxidase and phenylalanine ammonia lyase on the fourth day after challenge inoculation with *Sclerotium rolfsii* Sacc. This treatment also recorded the highest phenol content (1.179 µg g<sup>-1</sup> FW) on the fourth day after challenge inoculation with *S. rolfsii*. This treatment recorded the lowest wilt incidence (7.56 %) which indicated 90.41 percent disease reduction over control.

### Introduction

Tuberose (*Polianthes tuberosa* L.) is a commercially important ornamental bulbous plant cultivated in India for cut and loose flower trade and also for the extraction of its highly valued natural flower oil which is one of the most expensive raw materials of high-grade perfumes. It is native of Mexico, from where it has spread to different parts of world and is now one of the most important ornamentals of tropical and sub-tropical areas. It is used for artistic garland, floral ornaments, bouquets and buttonholes. The long flower

spikes are excellent as cut flowers for table decoration (Padaganur *et al.*, 2005).

Tuberose is often attacked by *Sclerotium rolfsii* Sacc. Which is a ubiquitous endemic soil borne plant pathogen. The initial symptom of the disease is flaccidity and drooping of leaves. The leaves become yellow and dry up. The fungus mainly affects the roots and the infection gradually spreads upward through the tuber and collar portion of the stem. Both tubers and roots show rotting

symptoms. Thick cottony growth of the fungus is visible on the rotten stem and on petioles at the soil level. Hexaconazole is a systemic fungicide used for the control of *Sclerotium rolfsii* (Virupakshaprabhu and Hiremath, 2003). But, chemical control is not always effective and there is concern about the side effects of these fungicides on the environmental safety. Natural plant products and biological control, using antagonistic microorganisms, is an alternate approach to control the pathogenic attack. Biological control of soil borne pathogens offers environmentally safe alternative to chemicals. Different species of bacteria *Bacillus subtilis*, *Pseudomonas fluorescense* and fungi *Trichoderma viride* are reported to be effective biocontrol agents against soil borne plant pathogens (Sivasakthi *et al.*, 2014; Zape *et al.*, 2014).

Plants are known to produce various stress compounds when they are exposed to the pathogens (Lavana *et al.*, 2006; Kim *et al.*, 2008). Studies have shown that plants possess an effective antioxidant machinery to combat disease causing pathogen attack (Demidchik, 2012). Activation of a wide array of defense responses slows down or halts infection at certain stages of the host-pathogen interaction. An increase in the activities of phenolics related enzymes and the accumulation of phenolics has been correlated to plant resistance to biotic stresses (Anjum *et al.*, 2012). This study was conducted to explore the changes in some enzyme activities and phenol content in tuberose plants treated with bio-control and organic fungicide after inoculation with *Sclerotium rolfsii*.

## Materials and Methods

### Pathogen source

Six pathogenic isolates of *Sclerotium rolfsii* Sacc. were isolated from the diseased

tuberose plants collected from the six different tuberose growing areas of Madurai, Dindigul, Dharmapuri and Sivagangai districts of Tamil Nadu. These isolates produced the typical wilt symptoms on the artificially inoculated tuberose plants in pot culture. The degree of virulence varied among the isolates of *S. rolfsii* to cause wilt of tuberose. The most virulent isolate was selected and mass multiplied in sand maize medium and used for this study.

### Plant materials and treatment

The present investigation was carried out during November 2014 at Agricultural College and Research Institute, Madurai. Three months old tuberose plants were used for this experiment. Fifteen treatments were set up with soil application of antagonists (Tv4, Bs1, Pf3), Mahua cake and hexaconazole (Table 1). Inoculation with *S. rolfsii* was done and the leaves were collected from the plants on the 0, 2, 4, 6 and 10 days after inoculation. Leaves were washed several times with sterile distilled water and used for enzyme assays and phenol estimation.

### Enzyme assay

Peroxidase activity was estimated by the method of Hammerschmidt *et al.*, (1982). One gram of fresh leaf tissue was homogenised with 5 ml of 0.1M phosphate buffer pH 7.0 in a pre-cooled pestle and mortar. The homogenate was centrifuged at 15,000 rpm at 4°C for 15 minutes.

The supernatant was used as enzyme source. The reaction mixture consisted of 1.5 ml of 0.05M pyrogallol, 0.1 ml of enzyme extract and 0.5 ml of 1% H<sub>2</sub>O<sub>2</sub>. The change in absorbance of the reaction mixture was recorded at 420 nm at 30 seconds interval for three minutes at room temperature. The boiled enzyme preparation served as blank. The

enzyme activity was expressed as change in absorbance  $\text{min}^{-1}\text{g}^{-1}\text{FW}$ .

Polyphenol oxidase was assayed by the method adopted by Mayer *et al.*, (1965). One g of fresh leaf sample was homogenised in 1 ml of 0.1 M sodium phosphate buffer (pH 6.5). The homogenate was centrifuged at 15,000 rpm for 15 minutes at 4°C and the supernatant was used as the enzyme source. The reaction mixture consisted of 1.5 ml of 0.1M sodium phosphate buffer pH 6.5 and 0.1 ml of the enzyme extract. The reaction was initiated by the addition of 0.2 ml of catechol (0.01M). The absorbance at 495 nm at 30 sec intervals for three minutes was recorded. The enzyme activity was expressed as change in absorbance  $\text{min}^{-1}\text{g}^{-1}\text{FW}$ .

The activity of phenylalanine ammonia lyase was estimated by the method of Dickerson *et al.*, (1984). Five hundred mg of leaf was homogenized in 5 ml of cold 25mM borate HCl buffer (pH 8.8) containing 5mM mercaptoethanol. The homogenate was centrifuged at 15,000 rpm for 15 minutes at 4°C and the supernatant was used as enzyme source. The assay mixture consisted of 0.2 ml of enzyme extract, 1.3 ml water and 0.5 ml borate buffer. The reaction was initiated by the addition of 1 ml of 12mM L-Phenylalanine. The reaction mixture was incubated for one hour at 32°C. The reaction was stopped by the addition of 0.5 ml of 2N HCl. A blank was run in which phenylalanine was added after adding 2N HCl. The absorbance was measured at 290 nm. The enzyme activity was expressed as  $\square$  mol trans cinnamic acid  $\text{min}^{-1}\text{g}^{-1}\text{FW}$ .

### Estimation of total phenols

One g of the leaf sample was ground well in a pestle and mortar after adding 10 ml of 80% methanol. The homogenate was centrifuged at 10,000 rpm for 20 min. The supernatant was collected and evaporated to dryness and the

residue was dissolved in 5 ml of distilled water. From this, 0.2 ml was taken and the volume was made up to 3 ml with distilled water and to that 0.25 ml of Folin-Ciocalteu reagent (1N) was added. After three minutes, one ml of 20% sodium carbonate was added and placed in boiling water bath for one min and cooled. The absorbance was measured at 725 nm against a reagent blank. The total phenol content was expressed as  $\mu\text{g}$  catechol  $\text{g}^{-1}\text{FW}$  (Zieslin and Ben Zaken, 1993).

## Results and Discussion

### Enzyme activities

Peroxidases (PO) has been implicated in a number of diverse phenomena observed in plants such as lignification, suberization, cell elongation, growth and regulation of cell wall biosynthesis and plasticity, which diversified during disease period (Chanda and Singh, 1997). This enzyme is also known to produce a toxic environment for the pathogens through the production of oxidative burst (Passardi *et al.*, 2005).

The activity of peroxidase was induced in the plants treated with biocontrol agents, organic amendments and challenge inoculation with the pathogen *S. rolfsii*. The results revealed that the activity of PO was significantly higher in tuberoses plants treated with consortial formulation of Tv4 @ 2.5 kg ha<sup>-1</sup> + Pf3 @ 2.5 kg ha<sup>-1</sup> + Bs1 @ 2.5 kg ha<sup>-1</sup> + Mahua cake @ 250 kg ha<sup>-1</sup> (0.849 change in absorbance  $\text{min}^{-1}\text{g}^{-1}\text{FW}$ ) at four days after challenge inoculation with *S. rolfsii* (Table 1). Similar result was reported by Karthikeyan *et al.*, (2008).

The accumulation of polyphenol oxidase (PPO) plays an important role in plants defense mechanism for inhibition of pathogens by the mechanism of cell wall reinforcements (Ngadze *et al.*, 2012). PPO is a nuclear encoded enzyme that catalyzes the

oxygen dependent oxidation of phenols to quinones. PPOs are considered to have broad antimicrobial properties. In the present study, an increasing trend of PPO activity was seen up to four days after inoculation with *S. rolfsii* in all treatments and then decreased (Table 2).

The treatment Tv4 @ 2.5 kg ha<sup>-1</sup> + Pf3 @ 2.5 kg ha<sup>-1</sup> + Bs1 @ 2.5 kg ha<sup>-1</sup> + Mahua cake @ 250 kg ha<sup>-1</sup> recorded the maximum (1.289 change in absorbance min<sup>-1</sup> g<sup>-1</sup> FW) PPO activity as compared to the other treatments in tuberose plants.

**Table.1** Changes in peroxidase (PO) activity in tuberose plants inoculated with *S. rolfsii* and treated with soil application of antagonists and organic fungicide

T. No.	Treatment details	PO activity (Change in absorbance min <sup>-1</sup> g <sup>-1</sup> FW)*					
		Days after inoculation					
		0	2	4	6	10	
T <sub>1</sub>	Tv <sub>4</sub> (2.5 kg ha <sup>-1</sup> )	Inoculated	0.372	0.589	0.693	0.659	0.485
		Healthy	0.201	0.331	0.289	0.221	0.223
T <sub>2</sub>	Pf <sub>3</sub> (2.5 kg ha <sup>-1</sup> )	Inoculated	0.361	0.487	0.678	0.653	0.466
		Healthy	0.187	0.329	0.388	0.344	0.234
T <sub>3</sub>	Bs <sub>1</sub> (2.5 kg ha <sup>-1</sup> )	Inoculated	0.296	0.478	0.589	0.538	0.418
		Healthy	0.129	0.278	0.372	0.279	0.179
T <sub>4</sub>	Mahua cake (250 kg ha <sup>-1</sup> )	Inoculated	0.384	0.546	0.741	0.696	0.482
		Healthy	0.198	0.352	0.456	0.424	0.346
T <sub>5</sub>	Tv <sub>4</sub> (2.5 kg ha <sup>-1</sup> ) + Pf <sub>3</sub> (2.5 kg ha <sup>-1</sup> )	Inoculated	0.351	0.523	0.710	0.672	0.559
		Healthy	0.232	0.364	0.453	0.371	0.262
T <sub>6</sub>	Pf <sub>3</sub> (2.5 kg ha <sup>-1</sup> ) + Bs <sub>1</sub> (2.5 kg ha <sup>-1</sup> )	Inoculated	0.378	0.586	0.724	0.702	0.495
		Healthy	0.182	0.229	0.347	0.253	0.197
T <sub>7</sub>	Tv <sub>4</sub> (2.5 kg ha <sup>-1</sup> ) + Bs <sub>1</sub> (2.5 kg ha <sup>-1</sup> )	Inoculated	0.389	0.568	0.733	0.706	0.563
		Healthy	0.238	0.252	0.327	0.289	0.154
T <sub>8</sub>	Tv <sub>4</sub> (2.5 kg ha <sup>-1</sup> ) + Pf <sub>3</sub> (2.5 kg ha <sup>-1</sup> ) + Bs <sub>1</sub> (2.5 kg ha <sup>-1</sup> )	Inoculated	0.331	0.521	0.702	0.684	0.423
		Healthy	0.288	0.381	0.456	0.327	0.232
T <sub>9</sub>	Tv <sub>4</sub> (2.5 kg ha <sup>-1</sup> ) + Pf <sub>3</sub> (2.5 kg ha <sup>-1</sup> ) + Mahua cake (250 kg ha <sup>-1</sup> )	Inoculated	0.322	0.467	0.798	0.665	0.479
		Healthy	0.203	0.284	0.347	0.264	0.197
T <sub>10</sub>	Pf <sub>3</sub> (2.5 kg ha <sup>-1</sup> ) + Bs <sub>1</sub> (2.5 kg ha <sup>-1</sup> ) + Mahua cake (250 kg ha <sup>-1</sup> )	Inoculated	0.357	0.427	0.678	0.636	0.346
		Healthy	0.212	0.288	0.393	0.343	0.212
T <sub>11</sub>	Tv <sub>4</sub> (2.5 kg ha <sup>-1</sup> ) + Bs <sub>1</sub> (2.5 kg ha <sup>-1</sup> ) + Mahua cake (250 kg ha <sup>-1</sup> )	Inoculated	0.322	0.418	0.587	0.539	0.398
		Healthy	0.232	0.257	0.293	0.221	0.122
T <sub>12</sub>	Tv <sub>4</sub> (2.5 kg ha <sup>-1</sup> ) + Pf <sub>3</sub> (2.5 kg ha <sup>-1</sup> ) + Bs <sub>1</sub> (2.5 kg ha <sup>-1</sup> ) + Mahua cake (250 kg ha <sup>-1</sup> )	Inoculated	0.418	0.647	0.849	0.789	0.523
		Healthy	0.245	0.319	0.452	0.363	0.245
T <sub>13</sub>	Hexaconazole (0.1%)	Inoculated	0.373	0.591	0.826	0.796	0.528
		Healthy	0.219	0.322	0.668	0.443	0.294
T <sub>14</sub>	Control (Inoculated)		0.329	0.366	0.501	0.479	0.276
T <sub>15</sub>	Healthy control		0.336	0.387	0.519	0.491	0.288
	CD (p=0.05) Treatments 0.007 Days 0.004 Treatment x Days 0.017						

\*All values are means of three replications

**Table.2** Changes in polyphenol oxidase (PPO) activity in tuberose plants inoculated with *S.rolfsii* and treated with soil application of antagonists and organic fungicide

T.No.	Treatment details	PPO activity (Change in absorbance min <sup>-1</sup> g <sup>-1</sup> FW)*					
		Days after inoculation					
		0	2	4	6	10	
T <sub>1</sub>	Tv <sub>4</sub> (2.5 kg ha <sup>-1</sup> )	Inoculated	0.443	0.589	0.788	0.643	0.603
		Healthy	0.418	0.478	0.701	0.598	0.475
T <sub>2</sub>	Pf <sub>3</sub> (2.5 kg ha <sup>-1</sup> )	Inoculated	0.438	0.583	0.788	0.649	0.607
		Healthy	0.366	0.487	0.701	0.608	0.575
T <sub>3</sub>	Bs <sub>1</sub> (2.5 kg ha <sup>-1</sup> )	Inoculated	0.419	0.432	0.674	0.610	0.573
		Healthy	0.389	0.407	0.546	0.512	0.464
T <sub>4</sub>	Mahua cake (250 kg ha <sup>-1</sup> )	Inoculated	0.464	0.632	0.806	0.735	0.536
		Healthy	0.412	0.578	0.739	0.662	0.484
T <sub>5</sub>	Tv <sub>4</sub> (2.5 kg ha <sup>-1</sup> ) + Pf <sub>3</sub> (2.5 kg ha <sup>-1</sup> )	Inoculated	0.467	0.532	0.798	0.694	0.634
		Healthy	0.398	0.486	0.679	0.601	0.559
T <sub>6</sub>	Pf <sub>3</sub> (2.5 kg ha <sup>-1</sup> ) + Bs <sub>1</sub> (2.5 kg ha <sup>-1</sup> )	Inoculated	0.474	0.989	1.018	0.752	0.549
		Healthy	0.423	0.910	0.932	0.634	0.488
T <sub>7</sub>	Tv <sub>4</sub> (2.5 kg ha <sup>-1</sup> ) + Bs <sub>1</sub> (2.5 kg ha <sup>-1</sup> )	Inoculated	0.446	0.785	0.943	0.715	0.517
		Healthy	0.412	0.702	0.821	0.669	0.436
T <sub>8</sub>	Tv <sub>4</sub> (2.5 kg ha <sup>-1</sup> ) + Pf <sub>3</sub> (2.5 kg ha <sup>-1</sup> ) + Bs <sub>1</sub> (2.5 kg ha <sup>-1</sup> )	Inoculated	0.423	0.656	0.832	0.698	0.626
		Healthy	0.388	0.553	0.797	0.558	0.516
T <sub>9</sub>	Tv <sub>4</sub> (2.5 kg ha <sup>-1</sup> ) + Pf <sub>3</sub> (2.5 kg ha <sup>-1</sup> ) + Mahua cake (250 kg ha <sup>-1</sup> )	Inoculated	0.476	0.772	0.859	0.683	0.615
		Healthy	0.422	0.537	0.798	0.542	0.521
T <sub>10</sub>	Pf <sub>3</sub> (2.5 kg ha <sup>-1</sup> ) + Bs <sub>1</sub> (2.5 kg ha <sup>-1</sup> ) + Mahua cake (250 kg ha <sup>-1</sup> )	Inoculated	0.467	0.679	0.812	0.648	0.632
		Healthy	0.416	0.523	0.735	0.463	0.412
T <sub>11</sub>	Tv <sub>4</sub> (2.5 kg ha <sup>-1</sup> ) + Bs <sub>1</sub> (2.5 kg ha <sup>-1</sup> ) + Mahua cake (250 kg ha <sup>-1</sup> )	Inoculated	0.425	0.498	0.791	0.647	0.472
		Healthy	0.386	0.399	0.637	0.598	0.313
T <sub>12</sub>	Tv <sub>4</sub> (2.5 kg ha <sup>-1</sup> ) + Pf <sub>3</sub> (2.5 kg ha <sup>-1</sup> ) + Bs <sub>1</sub> (2.5 kg ha <sup>-1</sup> ) + Mahua cake (250 kg ha <sup>-1</sup> )	Inoculated	0.994	1.177	1.289	0.920	0.618
		Healthy	0.883	0.948	1.022	0.757	0.485
T <sub>13</sub>	Hexaconazole(0.1%)	Inoculated	0.493	0.735	0.957	0.884	0.816
		Healthy	0.423	0.676	0.848	0.788	0.456
T <sub>14</sub>	Control (Inoculated)		0.402	0.425	0.605	0.479	0.308
T <sub>15</sub>	Healthy control		0.414	0.423	0.658	0.506	0.347
	CD (p=0.05) Treatments 0.011 Days 0.006 Treatment x Days 0.026						

\*All values are means of three replications

**Table.3** Changes in phenylalanine ammonia lyase (PAL) activity in tuberose plants inoculated with *S. rolfsii* and treated with soil application of antagonists and organic fungicide

T.No	Treatment details		PAL activity ( $\mu\text{mol trans-cinnamic acid min}^{-1}\text{g}^{-1}\text{FW}$ )*				
			Days after inoculation				
			0	2	4	6	10
T <sub>1</sub>	Tv <sub>4</sub> (2.5 kg ha <sup>-1</sup> )	Inoculated	0.462	0.588	0.697	0.686	0.604
		Healthy	0.323	0.546	0.589	0.538	0.424
T <sub>2</sub>	Pf <sub>3</sub> (2.5 kg ha <sup>-1</sup> )	Inoculated	0.412	0.517	0.663	0.645	0.593
		Healthy	0.378	0.486	0.545	0.328	0.234
T <sub>3</sub>	Bs <sub>1</sub> (2.5 kg ha <sup>-1</sup> )	Inoculated	0.436	0.486	0.583	0.541	0.487
		Healthy	0.369	0.384	0.589	0.532	0.424
T <sub>4</sub>	Mahua cake (250 kg ha <sup>-1</sup> )	Inoculated	0.478	0.523	0.721	0.702	0.676
		Healthy	0.388	0.472	0.681	0.568	0.538
T <sub>5</sub>	Tv <sub>4</sub> (2.5 kg ha <sup>-1</sup> ) + Pf <sub>3</sub> (2.5 kg ha <sup>-1</sup> )	Inoculated	0.463	0.498	0.707	0.626	0.578
		Healthy	0.425	0.436	0.688	0.596	0.482
T <sub>6</sub>	Pf <sub>3</sub> (2.5 kg ha <sup>-1</sup> ) + Bs <sub>1</sub> (2.5 kg ha <sup>-1</sup> )	Inoculated	0.481	0.578	0.745	0.686	0.648
		Healthy	0.416	0.526	0.692	0.578	0.498
T <sub>7</sub>	Tv <sub>4</sub> (2.5 kg ha <sup>-1</sup> ) + Bs <sub>1</sub> (2.5 kg ha <sup>-1</sup> )	Inoculated	0.462	0.524	0.737	0.678	0.536
		Healthy	0.428	0.469	0.688	0.567	0.436
T <sub>8</sub>	Tv <sub>4</sub> (2.5 kg ha <sup>-1</sup> ) + Pf <sub>3</sub> (2.5 kg ha <sup>-1</sup> ) + Bs <sub>1</sub> (2.5 kg ha <sup>-1</sup> )	Inoculated	0.434	0.512	0.726	0.632	0.512
		Healthy	0.386	0.464	0.679	0.588	0.448
T <sub>9</sub>	Tv <sub>4</sub> (2.5 kg ha <sup>-1</sup> ) + Pf <sub>3</sub> (2.5 kg ha <sup>-1</sup> ) + Mahua cake (250 kg ha <sup>-1</sup> )	Inoculated	0.416	0.507	0.714	0.608	0.538
		Healthy	0.389	0.462	0.684	0.546	0.462
T <sub>10</sub>	Pf <sub>3</sub> (2.5 kg ha <sup>-1</sup> ) + Bs <sub>1</sub> (2.5 kg ha <sup>-1</sup> ) + Mahua cake (250 kg ha <sup>-1</sup> )	Inoculated	0.407	0.473	0.711	0.567	0.523
		Healthy	0.326	0.387	0.674	0.558	0.477
T <sub>11</sub>	Tv <sub>4</sub> (2.5 kg ha <sup>-1</sup> ) + Bs <sub>1</sub> (2.5 kg ha <sup>-1</sup> ) + Mahua cake (250 kg ha <sup>-1</sup> )	Inoculated	0.392	0.442	0.708	0.519	0.473
		Healthy	0.323	0.344	0.689	0.483	0.425
T <sub>12</sub>	Tv <sub>4</sub> (2.5 kg ha <sup>-1</sup> ) + Pf <sub>3</sub> (2.5 kg ha <sup>-1</sup> ) + Bs <sub>1</sub> (2.5 kg ha <sup>-1</sup> ) + Mahua cake (250 kg ha <sup>-1</sup> )	Inoculated	0.507	0.686	0.872	0.732	0.687
		Healthy	0.428	0.567	0.736	0.687	0.632
T <sub>13</sub>	Hexaconazole (0.1%)	Inoculated	0.491	0.533	0.732	0.712	0.646
		Healthy	0.367	0.437	0.658	0.646	0.548
T <sub>14</sub>	Control (Inoculated)		0.398	0.516	0.704	0.579	0.426
T <sub>15</sub>	Healthy control		0.384	0.426	0.628	0.433	0.403
	CD (p=0.05) Treatments 0.010 Days 0.005 Treatment x Days 0.022						

\*All values are means of three replications

**Table.4** Changes in total phenolic content in tuberose plants inoculated with *S. rolfsii* and treated with soil application of antagonists and organic fungicide

T.No.	Treatment details	Total phenols ( $\mu\text{g catechol g}^{-1}\text{ FW}$ )*					
		Days after inoculation					
		0	2	4	6	10	
T <sub>1</sub>	Tv <sub>4</sub> (2.5 kg ha <sup>-1</sup> )	Inoculated	0.453	0.588	0.789	0.646	0.606
		Healthy	0.412	0.482	0.711	0.596	0.473
T <sub>2</sub>	Pf <sub>3</sub> (2.5 kg ha <sup>-1</sup> )	Inoculated	0.435	0.582	0.786	0.648	0.602
		Healthy	0.368	0.486	0.703	0.608	0.576
T <sub>3</sub>	Bs <sub>1</sub> (2.5 kg ha <sup>-1</sup> )	Inoculated	0.418	0.431	0.676	0.615	0.573
		Healthy	0.389	0.408	0.544	0.516	0.466
T <sub>4</sub>	Mahua cake (250 kg ha <sup>-1</sup> )	Inoculated	0.476	0.636	0.804	0.738	0.536
		Healthy	0.412	0.578	0.739	0.666	0.482
T <sub>5</sub>	Tv <sub>4</sub> (2.5 kg ha <sup>-1</sup> ) + Pf <sub>3</sub> (2.5 kg ha <sup>-1</sup> )	Inoculated	0.463	0.536	0.796	0.694	0.644
		Healthy	0.398	0.486	0.689	0.621	0.569
T <sub>6</sub>	Pf <sub>3</sub> (2.5 kg ha <sup>-1</sup> ) + Bs <sub>1</sub> (2.5 kg ha <sup>-1</sup> )	Inoculated	0.486	0.689	0.818	0.752	0.546
		Healthy	0.423	0.536	0.732	0.635	0.484
T <sub>7</sub>	Tv <sub>4</sub> (2.5 kg ha <sup>-1</sup> ) + Bs <sub>1</sub> (2.5 kg ha <sup>-1</sup> )	Inoculated	0.442	0.783	0.643	0.718	0.516
		Healthy	0.426	0.702	0.826	0.669	0.436
T <sub>8</sub>	Tv <sub>4</sub> (2.5 kg ha <sup>-1</sup> ) + Pf <sub>3</sub> (2.5 kg ha <sup>-1</sup> ) + Bs <sub>1</sub> (2.5 kg ha <sup>-1</sup> )	Inoculated	0.438	0.662	0.834	0.696	0.628
		Healthy	0.386	0.552	0.794	0.568	0.521
T <sub>9</sub>	Tv <sub>4</sub> (2.5 kg ha <sup>-1</sup> ) + Pf <sub>3</sub> (2.5 kg ha <sup>-1</sup> ) + Mahua cake (250 kg ha <sup>-1</sup> )	Inoculated	0.476	0.772	0.859	0.683	0.615
		Healthy	0.432	0.548	0.785	0.552	0.526
T <sub>10</sub>	Pf <sub>3</sub> (2.5 kg ha <sup>-1</sup> ) + Bs <sub>1</sub> (2.5 kg ha <sup>-1</sup> ) + Mahua cake (250 kg ha <sup>-1</sup> )	Inoculated	0.468	0.683	0.822	0.658	0.622
		Healthy	0.414	0.538	0.757	0.473	0.432
T <sub>11</sub>	Tv <sub>4</sub> (2.5 kg ha <sup>-1</sup> ) + Bs <sub>1</sub> (2.5 kg ha <sup>-1</sup> ) + Mahua cake (250 kg ha <sup>-1</sup> )	Inoculated	0.425	0.498	0.791	0.647	0.472
		Healthy	0.383	0.389	0.635	0.588	0.323
T <sub>12</sub>	Tv <sub>4</sub> (2.5 kg ha <sup>-1</sup> ) + Pf <sub>3</sub> (2.5 kg ha <sup>-1</sup> ) + Bs <sub>1</sub> (2.5 kg ha <sup>-1</sup> ) + Mahua cake (250 kg ha <sup>-1</sup> )	Inoculated	0.984	0.998	1.179	0.890	0.756
		Healthy	0.873	0.858	1.012	0.798	0.685
T <sub>13</sub>	Hexaconazole (0.1%)	Inoculated	0.495	0.734	0.967	0.882	0.815
		Healthy	0.423	0.676	0.846	0.786	0.589
T <sub>14</sub>	Control (Inoculated)		0.403	0.425	0.606	0.478	0.310
T <sub>15</sub>	Healthy control		0.416	0.434	0.668	0.536	0.378
CD (p=0.05) Treatments 0.011 Days 0.006 Treatment x Days 0.025							

\*All values are means of three replications

Phenylalanine ammonia lyase (PAL) is the primary enzyme in the phenylpropanoid pathway, which leads to the conversion of L-phenylalanine to trans-cinnamic acid with the elimination of ammonia. It is the key enzyme in the synthesis of several defense related secondary compounds such as phenols and lignin (Anjum, 2012). Activity of phenylalanine ammonia lyase was induced in tuberose plants treated with effective

biocontrol agents, organic fungicide and challenge inoculation with the pathogen *S. rolfsii*. The enzyme activity reached the maximum at four days after inoculation and maintained at higher level up to six days after challenge inoculation with respective pathogen and declined thereafter in all the treatments. Whereas, in healthy and pathogen inoculated control plants, the activity was less than that in the other treatments. Consortial

formulation of biocontrol agents Tv4 @ 2.5 kg ha<sup>-1</sup> + Pf3 @ 2.5 kg ha<sup>-1</sup> + Bs1 @ 2.5 kg ha<sup>-1</sup> + Mahua cake @ 250 kg ha<sup>-1</sup> recorded higher activity of PAL enzyme in plants. The maximum PAL activity (0.872  $\mu$  mol of transcinamic acid min<sup>-1</sup> g<sup>-1</sup> FW) was recorded at four days after challenge inoculation. Similar results were reported by Kannan and Karthik (2009) and Nandhini *et al.*, (2010) (Table 3).

### Total phenols

Phenolics are known to be involved in plant-pathogen interactions. Some of the oxidized products of phenols are toxic to microorganisms. Phenols also act as antioxidants in scavenging reactive oxygen species (Demidchik, 2012).

The present study showed that total phenol content had an increasing trend up to fourth day. Consortial formulation of biocontrol agents Tv4 @ 2.5 kg ha<sup>-1</sup> + Pf3 @ 2.5 kg ha<sup>-1</sup> + Bs1 @ 2.5 kg ha<sup>-1</sup> + Mahua cake @ 250 kg ha<sup>-1</sup> recorded higher phenol content in plants. The maximum amount of phenol (1.179  $\mu$ g of catechol g<sup>-1</sup> FW) was recorded at four days after challenge inoculation and then declined slowly and reached 0.756  $\mu$ g of catechol g<sup>-1</sup> FW at ten days after inoculation (Table 4). Previous studies also reported similar trends (Angayarkanni, 2014).

It is concluded that defense related enzyme activity and phenol content increased when subjected to pathogen attack along with treatment with antagonists and fungicides.

The activity of defense related enzymes was higher in the treatment combination of Tv4 @ 2.5 kg ha<sup>-1</sup> + Pf3 @ 2.5 kg ha<sup>-1</sup> + Bs1 @ 2.5 kg ha<sup>-1</sup> + Mahua cake @ 250 kg ha<sup>-1</sup> which in turn resisted the growth of pathogen (*S. rolfisii* Sacc.), obviously controlling the wilt in tuberose.

### References

- Anjum, T, Fatima, S. and Amjad, S. 2012. Physiological changes in wheat during development of loose smut. *Tropical Plant Pathol.* 37: 102-107.
- Angayarkanni, T. 2014. Biomangement of root rot and leaf spot disease of *Stevia rebaudiana* using plant growth promoting rhizobacteria. *Shodhganga.* pp 1-80.
- Chanda, S.V., Singh, Y.D. 1997. Changes in peroxidase and IAA oxidase activities during wheat grain development. *Plant Physiol. Biochem.* 35: 245-250.
- Demidchik, V. 2012. Reactive oxygen species and oxidative stress in plants In: *Plant stress physiology*, Shabala. S (Ed). CAB International, UK. pp 24-58.
- Dickerson, D.P., Pascholati, S.F., Hagerman, A.E., Butler, L.G. and Niholson, R.L. 1984. Phenylalanine ammonia lyase and hydroxyl cinnamate: CoA ligase in maize mesocotyls inoculated with *Helminthosporium maydis* or *Helminthosporium carbonum*. *Physiol. Plant Pathol.* 25: 111-123.
- Hammerschmidt, R., Nuckles, E.M. and Kuc, J. 1982. Association of enhanced peroxidase activity with induced systemic resistance of cucumber to *Colletotrichum lagenarium*. *Physiol. Plant Pathol.* 20: 73-82.
- Kannan, C. and Karthik, M. 2009. Systemic induction of defense enzymes by rhizosphere microbes in cocoa seedlings. *J. Biol. Control.* 23: 427-431.
- Karthikeyan, M., Radhika, K., Bhaskaran, R., Mathiyazhagan, S., Sandoskumar, R., Velazhahan, R. and Alice, D. 2008. Biological control of onion leaf blight disease by bulb and foliar application of powder formulation of antagonist mixture. *Archives of Phytopathol. And Plant Protection.* 41: 407 – 417.



- Kim, Y.H., Kim, C.Y., Song, W.K., Park, D.S., Kwon, S.Y., Lee, H.S., Bang, J.W. and Kwak, S.S. 2008. Over expression of sweet potato swpa4 peroxidase results in increased hydrogen peroxide production and enhances stress tolerance in tobacco. *Planta*. 227: 867-881.
- Lavania, M., Chauhan, P.S., Chauhan, S.V.S., Singh, H.B. and Nautiyal, C.S. 2006. Induction of plant defence enzymes and phenolics by treatment with plant growth-promoting rhizobacteria *Serratia marcescens* NBRI1213. *Current Microbiol.* 52: 363-368.
- Mayer, A.M., Harel, E. and Shaul, R.B. 1965. Assay of catechol oxidase a critical comparison of methods. *Phytochemistry*. 5: 783-789.
- Nandhini, D., Mohan, J.S. and Singh, G. 2010. Induction of Systemic acquired resistance in *Arachis hypogea* L. by *Sclerotium rolfsii* derived elicitors. *J. Pytopathol.* 158: 594-600.
- Ngadze, E., Icishahayo, D., Coutinho, T.A. and van der Waals, J.E. 2012. Role of polyphenol oxidase, peroxidase, phenylalanine ammonia lyase, chlorogenic acid, and total soluble phenols in resistance of potatoes to soft rot. *Plant Disease*. 96: 186-192.
- Passardi, F., Cosio, C., Penel, C. and Dunand, C. 2005. Peroxidases have more functions than a Swiss army knife. *Plant Cell Reports*. 24: 255-265.
- Sivasakthi, S., Usharani, G. and Saranraj, P. 2014. Biocontrol potentiality of plant growth promoting bacteria (PGPR)-*Pseudomonas fluorescens* and *Bacillus subtilis*; a review. *Afr. J. Agricultural Research*. 9: 1265-1277.
- Virupakshaprabhu, H. and Hiremath, P.C. 2003. Biological control of collar rot of cotton caused by *Sclerotium rolfsii* Sacc. *Karnataka J. Agricultural Sciences*. 16: 576-579.
- Zape, A.S., Gade, R.M., Singh, R. and Deshmukh, V.A. 2014. Efficacy of different antagonist against the *Sclerotium rolfsii*, *Rhizoctonia solani* and *Fusarium solani*. *The Bioscan*. 9: 1431-1434.

**How to cite this article:**

Ragavi, G., M.L. Mini, I. Yesuraja and Sethuraman, K. 2017. Induction of Enzyme Activities in Tuberoses Plants Treated with Antagonists and Organic Fungicide under Artificial Inoculation of *Sclerotium rolfsii* Sacc.. *Int.J.Curr.Microbiol.App.Sci.* 6(6): 595-603.  
doi: <https://doi.org/10.20546/ijcmas.2017.606.070>