

Original Research Article

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Effect of Cold Temperature Durations on the Emergence and Parasitization Efficiency of Laboratory Reared *Trichogramma chilonis* (Ishii)

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ABSTRACT

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The result over fitness life parameters of *Trichogramma* revealed that the percentage emergence of *Trichogramma* recorded after 5 days of storage at different temperature revealed that 10⁰C gave significantly highest emergence of parasitoid (96.20%) and was similar to control (97.4%) however; other temperatures, 12, 14 and 16⁰C showed 90.90%, 89.43% and 88.33% emergence of *Trichogramma*, respectively. At 10⁰C, the highest parasitism percentage (96.00%) was observed and further decreased to 53.66% at 10⁰C when stored for up to 30 days whereas, the lowest parasitism percentage assessed at 16⁰C (89.83%) and 6⁰C (90.03%). The maximum adult longevity measured, while it was stored at 6⁰C (4.50 days), whereas shortest adult longevity noted at 16⁰C (3.00 days). Further, adult longevity was decreased from 4.26 to 0.00 days when stored at 10⁰C from 5 to 50 days. At 6⁰C, *Trichogramma* gave considerable emergence (76.13-20.63%) and parasitization (91.46-42.83%) from 5 to 50 days storage, whereas at 12, 14 and 16⁰C, development was completed during storage. It was evident that storage at 10⁰C and 6⁰C were very conducive for life parameters of *Trichogramma* to get short term and long storage, respectively.

Introduction

Trichogramma species are widely used egg parasitoids for biological control of insect pests of different crops through augmentation and release. *T. chilonis* and *T. achaeae* have been promoted to use in controlling cabbage leaf eating caterpillars (Krishnamoorthy, 2012). Among the *Trichogramma* species the egg parasitoid, *T. chilonis* is the dominant species in India (Nagarkatti and Nagaraja, 1979). It is a very aggressive parasitoid and has the ability to increase their capability, sometimes gives near to 100% parasitism depending upon the availability of favorable condition. It is a natural enemy of many

harmful lepidopterous insect pests of crops and vegetables (Jalali and Singh, 1993). *Trichogramma* has been used against the lepidopterous pests of cotton, cabbage, apple and tomato (Smith, 1996). They parasitize the eggs of more than 400 lepidopteran pest species (Khan *et al.*, 2004 and Doyon and Boivin, 2005).

Mass rearing of bioagents is a prerequisite of biocontrol programme; this needs a regular and sufficient production of easily culturable factitious insect hosts for mass culturing of any bioagent (Wadaskar *et al.*,

2015). *Trichogramma* species is one of the most widely used biological control agent due to its easy rearing in insectaries and vigorous parasitism on eggs of target hosts. Thus, good quality egg parasitoid, *T. chilonis* could be utilized through inundative release for the management of many lepidopterous insect pest (Bhushan *et al.*, 2012 and Fand *et al.*, 2013).

Cold storage technique must ensure the availability of sufficient numbers (Tezze and Botto, 2004) and quality of egg parasitoids (Bigler, 1994) at the time of release. Therefore, the development of storage techniques for bio-control agents is considered of utmost importance to provide flexibility and efficiency in mass production, to synchronize a desired stage of development for peak release, and to make available standardized stocks for use in research (Greenberg *et al.*, 1996; Leopold, 1998 and Ravensberg, 1992). Besides, cold storage can permit a more cost-effective production schedule (Glenister and Hoffmann, 1998) providing a means to conserve biological control agents when not immediately needed (Pitcher *et al.*, 2002).

To get high rate of emergence in laboratory in hot summer, artificial manipulation in temperature is necessary for successful rearing (Rajendran, 1999). As stated above, there is a demand for information in relation to the mass rearing, emergence and parasitization ability of *T. chilonis* for successful implementation of bio control programmes. Hence, in this report, hypothesis can be drawn to evaluate the effect of a range of cold storage periods on the subsequent performance of *T. chilonis* to assess the effects of such storage on the emergence and parasitization efficiency of laboratory reared *T. chilonis* on eggs of its factitious host *Corcyra cephalonica*.

Materials and Methods

The present investigations on influence of storage on the emergence and parasitization efficiency of laboratory reared *Trichogramma* was carried out at Biocontrol Research Laboratory, JAU, Junagadh.

Experiment was conducted to find out the optimum storage temperature and duration for the parasitoid, *T. chilonis* at pupal stage in the BOD incubators. 200 eggs of *Corcyra* were pasted on the 17 × 11 cm paper card strips and exposed for 24 hrs to the one day age old parasitoids confined in test tubes. A drop of 50% pure honey was provided as adult feed through sterilized absorbent cotton which was placed inside the test tube. Host eggs on strips after exposure of 24 hrs to the parasitoids were taken out from test tube and kept under standard laboratory conditions *i.e.*, $28 \pm 1^{\circ} \text{C}$, and $65 \pm 5\% \text{RH}$ (Nadeem *et al.*, 2010).

These parasitized cards in the pupal stage were stored at six different temperature regimes *viz.*, 6, 8, 10, 12, 14 and 16°C each at 5, 10, 15, 20, 25, 30, 40 and 50 days in biological oxygen demand (BOD) incubators with complete darkness. After completing the respective storage duration, the parasitoid strips were taken out from the BOD incubator and placed at the standard conditions, where they were emerged after 1-3 days.

Observations recorded

The storage period was recorded precisely. At each temperature and storage period, per cent emergence, parasitism and longevity of adults were observed with meticulous care. Percentage of parasitism which was calculated by number of blackened eggs/number of total eggs × 100 and longevity was calculated as days from the day of emergence to day of death.

Results and Discussion

Per cent emergence of *T. chilonis*

The percentage emergence of *Trichogramma* recorded after 5 days of storage at different temperature revealed that 10⁰C gave significantly highest emergence of parasitoid (96.20%) and lowest (76.13%) from 6⁰C (Table 1). At ten days storage, the highest (94.10%) emergence was observed from *Trichogramma* parasitoids after held at 10⁰C storage. emergence percentage. At 10⁰C the highest (92.23%) emergence percentage after storage for fifteen days and at 6⁰C (55.30%) the least emergence was noted. At twenty days, pupal storage emphatically discloses the significant results on emergence percentage. The highest emergence percentage of *Trichogramma* obtained from 12⁰C (66.06%) and the lowest (43.63%) from 6⁰C.

The delineated results emphasize significant emergence percentage of *Trichogramma* after stored for twenty five days. The highest emergence percentage of *Trichogramma* obtained from 8⁰C (43.33%) was statistically at par with 10 (41.00%) and 12⁰C (41.70%). The lowest emergence percentage was observed at 14 and 16⁰C emergence was utterly occluded (Table 1). At 6⁰C emergence percentage seemed moderately (32.86%). The results revealed significant demarcation on emergence percentage of *Trichogramma* after thirty days storage. The highest emergence percentage of *Trichogramma* obtained from 10⁰C (36.66%). The second highest emergence percentage of *Trichogramma* evaluated at 12⁰C (32.93%) which was statistically at par with 6⁰C (30.60%) and 8⁰C (31.93%). Apparently the emergence percentage of *Trichogramma* after storage for forty days obviously divulged significant results. The highest emergence percentage recorded at 6⁰C (26.96%). Lowest emergence percentage recorded, at 8⁰C (18.76%). Adult

emergence was entirely thwarted at 10, 12, 14 and 16⁰C. After fifty days of storage the highest emergence percentage recorded at 6⁰C (20.63%). Lowest emergence percentage recorded, at 8⁰C (10.43%). Adult emergence was entirely thwarted at 10, 12, 14 and 16⁰C. The present result indicated that the parasitized *Corcyra* eggs could be stored for 50 days at low temperature (6⁰C) for long duration storage. It was evidenced from the above results that the emergence of *Trichogramma* was decreased with increasing storage temperatures and days intervals for storage.

The present results enunciated the considerable emergence of adults up to fifty days at 6⁰C and it pertinent with findings of Gharbi (2014) who reported that the emergence rate was 87.56 percent at 15⁰C but significantly decreased to 45.32 percent at 35⁰C.

Per cent parasitism of *T. chilonis*

The result on per cent parasitism of *Trichogramma* revealed that after five days storage, the parasitism percentage was differed according to different low temperatures. At 10⁰C, the highest parasitism percentage (96.00%) was observed (Table 2). However, the lowest (90.03%) parasitism percentage of *Trichogramma* observed from 16⁰C. The empirical data showed significant difference in results on parasitization percentage of *T. chilonis*, after stored for ten days at different low temperatures. The highest parasitism percentage of *Trichogramma* obtained, when it was reared at 10⁰C (94.30%) which was statistically at par with 8⁰C (93.70%). The lowest parasitism percentage of *Trichogramma* assessed, when it was reared at 16 (89.83%) and 6⁰C (90.03%), which was statistically at par with each other.

Table.1 Emergence percentage of *T. chilonis* after stored at various low temperatures

Temp.	Emergence (%) at different storage duration (Days)							
	5	10	15	20	25	30	40	50
6 ^o C	60.77(76.13)	50.32(59.23)	48.04(55.30)	41.34(43.63)	34.97(32.86)	33.58(30.60)	31.28(26.96)	27.01(20.63)
8 ^o C	76.79 (94.53)	73.55 (91.93)	71.37 (89.70)	53.67 (64.86)	41.16 (43.33)	34.40 (31.93)	25.67 (18.76)	18.84 (10.43)
10 ^o C	79.08 (96.20)	76.18 (94.10)	73.90 (92.23)	53.98 (65.40)	39.81 (41.00)	37.26 (36.66)	4.05 (0.50)	4.05 (0.50)
12 ^o C	72.57 (90.90)	69.98 (88.20)	67.14 (84.90)	54.37 (66.06)	40.22 (41.70)	35.02 (32.93)	4.05 (0.50)	4.05 (0.50)
14 ^o C	71.16 (89.43)	70.19 (88.46)	66.43 (84.00)	53.43 (64.50)	4.05 (0.50)	4.05 (0.50)	4.05 (0.50)	4.05 (0.50)
16 ^o C	70.03 (88.33)	68.44 (86.50)	66.01 (83.46)	52.72 (63.30)	4.05 (0.50)	4.05 (0.50)	4.05 (0.50)	4.05 (0.50)
Control	80.98 (97.40)							
S.Em ±	2.20	1.67	1.31	1.40	0.62	0.50	0.16	0.10
C.D. at 5 %	4.72	3.64	2.85	3.04	1.35	1.08	0.35	0.21
C.V.%	3.69	3.01	2.45	3.32	2.79	2.48	1.64	1.15

Table.2 Parasitization percentage of *T. chilonis* after stored at various low temperatures

Temp.	Parasitism(%) at different storage duration (Days)							
	5	10	15	20	25	30	40	50
6 ⁰ C	73.07*(91.46)	71.64(90.03)	65.27(82.46)	63.87(80.60)	63.02(79.40)	60.51(75.76)	55.53(67.96)	40.87(42.83)
8 ⁰ C	76.41 (94.36)	75.48 (93.70)	68.39 (86.40)	60.42 (75.63)	54.15 (65.70)	51.21(60.76)	40.99 (43.03)	33.12 (29.86)
10 ⁰ C	78.59 (96.00)	76.21 (94.30)	70.23 (88.53)	57.57 (71.23)	51.06 (60.50)	47.10 (53.66)	4.05 (0.50)	4.05 (0.50)
12 ⁰ C	73.85 (92.20)	73.17 (91.60)	72.00 (90.36)	54.07 (65.56)	45.32 (50.56)	41.90 (44.60)	4.05 (0.50)	4.05 (0.50)
14 ⁰ C	73.97 (92.30)	73.26 (91.70)	67.56 (85.40)	51.14 (60.63)	4.05 (0.50)	4.05 (0.50)	4.05 (0.50)	4.05 (0.50)
16 ⁰ C	71.68 (90.03)	71.43 (89.83)	63.71 (80.36)	48.15 (55.50)	4.05 (0.50)	4.05 (0.50)	4.05 (0.50)	4.05 (0.50)
Control	80.71 (97.23)							
S.Em ±	1.89	0.95	1.38	0.58	0.56	0.44	0.28	0.12
C.D. at 5 %	4.04	2.05	3.01	1.27	1.22	0.96	0.61	0.26
C.V.%	3.06	1.57	2.50	1.28	1.87	1.56	1.85	1.00

Table.3 Adult longevity (days) of *T. chilonis* after stored at various low temperatures

Temp.	Adult longevity (%) at different storage duration (Days)							
	5	10	15	20	25	30	40	50
6 ⁰ C	5.26	5.10	4.50	4.00	3.70	3.60	3.00	2.03
8 ⁰ C	5.60	4.63	4.10	3.70	3.00	2.80	2.00	1.03
10 ⁰ C	4.26	3.83	3.50	3.53	3.20	3.00	0.50	0.50
12 ⁰ C	4.40	3.83	3.20	3.10	2.20	2.00	0.50	0.50
14 ⁰ C	4.06	3.60	3.10	2.50	0.50	0.50	0.50	0.50
16 ⁰ C	3.76	3.33	3.00	2.00	0.50	0.50	0.50	0.50
Control	6.10							
S.Em ±	0.13	0.11	0.11	0.10	0.07	0.07	0.05	0.03
C.D. at 5%	0.27	0.24	0.24	0.22	0.14	0.14	0.10	0.05
C.V.%	3.29	3.39	3.80	3.97	3.74	3.95	4.95	3.95

The pragmatic data of *Trichogramma* revealed significant difference in results, when it was stored for fifteen days at different low temperatures. The utmost level of parasitization percentage assessed, while it was stored at 12⁰C (90.36%). Similar trend of parasitism was observed at 10⁰C (88.53%). The least parasitization percentage of *Trichogramma* noted from 16⁰C (80.36%), while it was statistically at par with 6⁰C (82.46%). At twenty five and thirty days, *Trichogramma* pupal storage fetched significant results on parasitization by parasitoids (Table 2). The uppermost parasitization percentage of *Trichogramma* recorded at 6⁰C (79.40%, 75.76%) followed by at 8⁰C (65.70%, 60.76%), respectively. The highest parasitization percentage recorded after forty days storage at 6⁰C (67.96%), while it was found lowest at 8⁰C (43.03%). Emergence was totally thwarted at 10, 12, 14 and 16⁰C so, there was no parasitization occurred. *Trichogramma* pupal storage up to fifty days brought significant results on parasitization of adults. The greatest level of parasitization recorded at 6⁰C (42.83%) followed by at 8⁰C (29.86%). The present result indicated that the parasitized *C. cephalonica* eggs could be stored for 50 days at low temperature (6⁰C) for long duration storage. It was evidenced from the above results that the parasitism of *Trichogramma* was decreased with increasing storage temperatures and days intervals for storage. The present findings indicated that the highest parasitism was obtained from the *T. chilonis* held at 10⁰C after five days storage, which was close to control values. This statement utterly supported the Nadeem *et al.*, (2010) who reported 97.4% parasitism after five days storage at 10⁰C. Kosha and Brar (2000) elucidated the *Trichogramma* could be stored in the refrigerator and successfully utilized for 23 days without adversely affecting their parasitization efficiency.

Adult longevity (days) of *T. chilonis*

The result on adult longevity revealed that the highest longevity of *Trichogramma* observed from 8⁰C (5.60 days) whereas, 5.26 days of adult longevity was perceived when the parasitized card stored at 6⁰C which was found next in order (Table 3). At 10 and 12⁰C, moderate longevity (4.26 and 4.40 days) obtained, which was statistically at par with each other. The shortest longevity period assessed at 16⁰C (3.76 days) and 14⁰C (4.06 days). Ten days storage of *Trichogramma* on adult longevity emphatically gave significant results. After stored for ten days, the lengthiest longevity recorded, when *Trichogramma* stored at 6⁰C (5.10 days) followed by 8⁰C (4.63 days). The shortest longevity of parasitoids evaluated from 16⁰C (3.33 days). The moderate longevity and statistically similar results were obtained at 10, 12 and 14⁰C such as 3.83, 3.83 and 3.60 days, respectively.

The maximum adult longevity measured, while it was stored at 6⁰C (4.50 days) followed by 8⁰C (4.10 days), whereas shortest adult longevity noted at 16⁰C (3.00 days). The 16⁰C was statistically at par with 12 (3.20 days) and 14⁰C (3.1 days). The moderate longevity seemed at 10⁰C (3.50 days). The present data revealed the significant results on adult longevity of *T. chilonis* after twenty days storage (Table 3). The utmost level of adult longevity obtained from 6⁰C (4.00 days) followed by at 8⁰C (3.70 days) which was statistically at par with 10⁰C (3.53 days). The shortest longevity assessed from 16⁰C (2.00 days). Perspicuous data expressed significant results on adult longevity of *Trichogramma* after twenty five and thirty days of storage. At 6⁰C, the maximum adult longevity of 3.70 and 3.60 days was assessed, respectively. At 14 and 16⁰C, adult emergence was completely occluded in twenty five and thirty days of storage. Longevity of *Trichogramma* after

storage of forty and fifty days provided significant results. The adult longevity ranged from 3.00 to 1.03 at 6⁰C and 8⁰C.

Our findings on decreased adult longevity at 8⁰C for prolonging storage are pertinent with the study reported by Ozder (2004), where adult longevity of egg parasitoid *T. cacoeciae* was decreased after 31 days storage at 8⁰C. Rundel *et al.*, (2004) exemplified the storage temperatures lower than 10⁰C and storage times 3 week or longer had a negative impact on longevity.

References

- Bhushan, S., Singh, R.P. and Shankar, R. 2012. Biopesticidal management of yellow stem borer (*Scirpophaga incertulas* Walker) in rice. *The Bioscan*, 7(2): 317-319.
- Bigler, F. 1994. Quality control in *Trichogramma* production. In: Biological Control with Egg Parasitoids, Wajnberg, E., Hassan, S.A. (Eds.), CAB International, Wallingford, UK. pp. 93-111.
- Doyon, J. and Boivin, G. 2005. The effect of development time on the fitness of female *Trichogramma evanscens*. *J. Insect Sci.*, 5(4): 5.
- Fand, B.B., Suroshe, S.S. and Gautam, R.D. 2013. Fortuitous biological control of insect pests and weeds: a critical review. *The Bioscan*, 8(1): 1-10.
- Gharbi, N. 2014. Influences of cold storage period and rearing temperature on the biological traits of *Trichogramma oleae*. *Tunisian J. Plant Prot.*, 9: 143-153.
- Glenister, C.S. and Hoffmann, M.P. 1998. Mass-reared natural enemies: scientific, technological, and informational needs and considerations. In: Mass Reared Natural Enemies: Application, Regulation, and Needs, Ridgway, R. L., Hoffmann, M. P., Inscoc, M. N., Glenister, C. S. (Eds.). Thomas Say Publications in Entomology, Entomological Society of America, Lanham, M. D. pp. 242-267.
- Greenberg, S.M., Nordlund, D.A. and King, E.G. 1996. Mass production of *Trichogramma* spp. *J. Biocontrol News and Information*, 17: 51-60.
- Jalali, S.K. and Singh, S.P. 1993. Superior strain selection of the egg parasitoid *Trichogramma chilonis* (Ishii), Biological parameters. *J. Biol. Control*, 7: 57-60.
- Khan, M.S., Farid, A., Ullah, F. and Badshah, H. 2004. Effect of host and parasitoid density on parasitism efficiency of *Trichogramma chilonis* Ishii. *Asian J. Plant Sci.*, 3: 647-650.
- Kosha, S.S. and Brar, S.K. 2000. Effect of storage on the emergence and parasitization efficiency of laboratory reared and field collected populations of *Trichogramma chilonis* (Ishii). *J. Biol. Control*, 14(2): 71-74.
- Krishnamoorthy, A. 2012. Exploitation of egg parasitoids for control of potential pests in vegetable ecosystems in India. *Comunicata Scientiae*, 3(1): 1 - 15.
- Leopold, R.A. 1998. Cold storage of insects for integrated pest management. In: Temper Temperature Sensitivity in Insects and Application in Integrated Pest Management, Hallman, G.J., Denlinger, D. L. (Eds.), Westview Press, Boulder, CO. pp. 235-267.
- Nadeem, S., Ashfaq, M., Hamed, M. and Ahmed, S. 2010. Optimization of short and long term storage duration for *Trichogramma chilonis* (Ishii) (Hymenoptera: Trichogrammatidae) at low temperatures. *Pak. J. Zool.*, 42: 63-67.
- Nagarkatti, S. and Nagaraja, H. 1979. The status of *Trichogramma chilonis* Ishii. *J. Oriental Insects*, 13: 115-117.

- Ozder, N. 2004. Effect of different cold storage periods on parasitization performance of *Trichogramma cacoeciae* (Hymenoptera: Trichogrammatidae) on eggs of *Ephestia kuehniella* (Lepidoptera: Pyralidae). *J. Biocontrol Sci. and Tech.*, 14: 441-7.
- Pitcher, S.A., Hoffmann, M.P., Gardner, J., Wright, M.G., and Kuhar, T.P. 2002. Cold storage of *Trichogramma ostriniae* reared on *Sitotroga cerealella* eggs. *J. Biocontrol*, 47: 525-535.
- Rajendran, B. 1999. Emergence of *Trichogramma chilonis* from the parasitoid cards under laboratory conditions during 1996-1998. *Coop. Sugar*, 31: 331.
- Ravensberg, W.J. 1992. Production and utilization of natural enemies in western European glasshouse crops. In: *Advances in Insect Rearing for Research and Pest Management*, Anderson, T.E., Leppla, N.C. (Eds.), Westview, Boulder, CO. pp. 465-487.
- Rundel, B.J., Thomson, L.J. and Hoffmann, A.A. 2004. Effects of cold storage on field and laboratory performance of *Trichogramma carverae* (Hymenoptera: Trichogrammatidae) and the response of three *Trichogramma* spp. (*T. carverae*, *T. brassicae*, and *T. funiculatum*) to cold. *Econ. J. Entomol.*, 97(2): 213-221.
- Smith, S.M. 1996. Biological control with *Trichogramma*: advances, successes, and potential of their use. *Ann. Rev. Entomol.*, 41: 375-406.
- Tezze, A.A. and Botto, E.N. 2004. Effect of cold storage on the quality of *Trichogramma nerudai* (Hymenoptera: Trichogrammatidae). *J. Biol. Control*, 30: 11-16.
- Wadaskar, P.S., Jethva, D.M., Vigneswaran S. and Rode, N.S. 2015. Studies on effect of temperature and relative humidity on biology of rice moth *Corcyra cephalonica* (Stainton) under laboratory condition. *The Ecoscan*, 9(1&2): 201-204.

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