
Original Research Article

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Studies on Occurrence of Invasive Salmonella spp. from Unorganised Poultry Farm to Retail Chicken Meat Shops in Mumbai City, India

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A B S T R A C T

The present study was planned to estimate occurrence of invasive Salmonella spp. in retail chicken supply chain of Mumbai and quantifying Salmonella at crucial stages of processing by the Most Probable Number (MPN) and confirmation by invA gene by PCR assay. A total of 18 (n = 108) farm samples were found to be positive for Salmonella with prevalence of 16.66% and statistical significance was observed amongst different sources at farm (p=0.027). Highest prevalence of Salmonella spp. was noticed in litter samples (50.00%) followed by cloacal swabs (25.00%), water utensil swabs (25.00%), faeces (16.66%), water (16.66%), wall dust (8.33%) and worker hand (8.33%). Over all prevalence of Salmonella spp. amongst various samples in retail shop was found to be 19.04 %. Out of 42 different chicken retail shop samples analysed 3 (7.14 %) swab samples of chopping board found positive while one sample each from water and swab samples of worker hand, platform, knife, and cloaca were found positive. Amongst 24 swab samples collected from chicken carcasses at various chicken processing stages, highest rate of contamination (50 %) was observed in post defeathering and post evisceration stages of processing with average count of 1.88 and 2.11, log MPN count/10cm² respectively. Out of 34 Salmonella isolates obtained in this study, 31 isolates showed positive amplification of 284 bp fragment specific for the invA gene with 91.17% detection level. Thus, study revealed that poultry litter at farm and post defeathering and post evisceration stages at retail chicken processing, are critical sources of cross contamination of invasive Salmonella spp.

Keywords
Poultry farms, retail chicken processing, Salmonella contamination, MPN quantification, invA.

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Introduction

Salmonella serotypes are significant zoonotic pathogens and cause a wide range of human diseases such as enteric fever, gastroenteritis and bacteremia in human and animals (Winokur et al., 2000 and Bennasar et al., 2000). Human salmonellosis is frequently associated with the consumption of poultry products (CDC, 2008; Hanning et al., 2009; Kang et al., 2009 and Pires et al., 2012). Contaminated poultry products are among the important sources for food-borne outbreaks in humans and Salmonella are isolated more often from poultry and poultry products (Habtamu et al., 2011; Kabir, 2010 and Linam and Gerber, 2007). As Salmonellosis is one of the most important foodborne diseases, few countries have a surveillance system that estimates the burden of salmonellosis in human populations (Flint et al., 2007 and WHO, 2005). Risk factors for colonization by Salmonella include season, hatchery of origin, feed mills, litter, water and...
various hygienic measures (Rose et al., 1999; Skov et al., 1999; Cardinale et al., 2004). Bryan and Doyle (1995) stated that commercially reared birds are in constant contact with litter and dust, both of which can be a source of contamination. Barnes, (1972) also mentioned that Salmonella contamination of birds may occur before, during or after the grow-out phase of production.

Indian broiler production has been growing, with an annual growth rate of 11.44 percent, production of 3.725 million tons (Index Mundi, 2015). In India, chicken is slaughtered at both industrial as well as at retail level but 95% chicken is slaughtered at retail level, while the remaining is slaughtered at industrial level (Badhe et al., 2013). Probability of cross contamination of raw chicken at retail level shops increases due to poor slaughtering practices, poor personal hygiene and poor cleaning. In India common sanitary problems that occur during the slaughtering and handling of poultry are hygienic condition and cleanliness of contact surfaces. Different genes like Inv, Spv, and Stn have been identified as major virulence genes responsible for pathogenic salmonellosis, the chromosomally located invasion gene invA being thought to trigger the invasion of Salmonellae into cultured epithelial cells (Asten and Dijk, 2005). Therefore it is important to evaluate the food safety risks because of pathogenic Salmonella along the production and retail processing and identification of effective control points or control strategies on the farm and at retail level chicken meat.

The present study was conducted to estimate occurrence of invasive Salmonella spp. in poultry farm environment of unorganized non-integrated broiler farms and six chicken retail shops in Mumbai using cultural isolation and enumeration by the Miniature Most Probable Number (MPN) method at stages of retail chicken meat production. Positive isolates were confirmed by amplifying invA gene which is unique to this genus and has been proved to be a suitable PCR target with a potential diagnostic application.

Materials & Methods

Sample Collection

Sample collection from poultry farm: A total of 108 different samples were collected from randomly selected 12 unorganized non-integrated poultry farms with capacity of 1500 -2000 birds, aged between 35 to 42 days and supplying birds to retail chicken meat shops located in vicinity of Mumbai. Samples includes cloacal swab from the poultry birds, feed, drinking water, litter sample from poultry house, fresh feaces and pooled swab from hands of the personnel working in the houses, wall dust, feeder and drinker.

Sample collection from retail chicken shops: A total of 66 stage wise post processing breast swabs samples of 10cm² area (post bleeding, post scalding, post defeathering and post evisceration), neck skin of carcass before and after evisceration, environmental samples (washing water, scalding water and carcass contact surfaces) and cloacal swabs were collected from six chicken processing establishments identified as retails chicken shops

Swab samples from 10cm² area were collected aseptically as per the standard methods described by (Gill and Jones, 2005).

Cultivation and isolation of Salmonella spp.

Qualitative evaluation: Isolation of Salmonella spp. from various samples collected was carried out as per ISO 6579. In brief pre-enrichment of the collected samples
in Buffered Peptone Water as 1:10 dilution and then incubated aerobically at 37°C for 18 hours. 0.1 ml inoculum was transferred to a tube containing 10 ml of the Rappaport Vassiliadis Soy broth and then incubated at 41.5°C for 24 hours. From the enrichment culture, 10 µl inoculum was further inoculated onto the surface of Xylose Lysine Deoxycholate (XLD) and Brilliant Green Agar (BGA) plates then incubated at 37°C for 24 hours. The plates containing characteristic colonies of Salmonella appearing as smooth colonies with black centre on XLD and red to pink on BGA were selected and the gram staining was performed. Colonies showing typical Gram negative, non spore forming short rod shaped appearance were further subjected to biochemical characterization with biochemically negative for hydrolysis of urea, positive for TSI with alkaline slant (red), acid butt (yellow) with H2S gas production and positive citrate utilization considered as positive for Salmonella spp.

Quantitative evaluation by miniature MPN technique

Pre enriched swab samples and neck samples of poultry carcass collected at stages viz, before and after evisceration, were subjected for quantitative miniaturized most probable number described by Pavic et al., (2009), based on ISO 6579-2002. The swab sample suspension 1 ml of a 10^1 dilution was pipetted into a U-bottomed 96 deep well plates (Genexy scientific, India). Serial decimal dilutions (100: 900 µl) were performed in BPW using a micropipette to the previously described final dilutions of 10^-6 in a labelled 96 well U bottomed plates. All tubes were mixed by repeated aspiration. From each of the dilutions in the plasma tubes, 100 µl aliquots were transferred into each of three wells (i.e. A1 to A3) across a another U-bottomed 96 deep well plates with each dilution in a subsequent row (i.e. 10^-1 in row A1–A3, 10^-2 in row B1 to B3 to a theoretical maximum dilution of 10^-6 in row F1–F3), producing a 3-tube MPN. The plate was then covered with adhesive paraffin wax film and incubated (37°C for 24 h). From each post incubated well, the total volume was transferred to a corresponding U-bottomed 96 deep well plates containing 500 µl MSRV and then incubated (42°C for 24 h).

White colour change from blue to colourless in a tube was considered as a presumptive positive for the presence of Salmonella, with all tubes (regardless of colour development) being confirmed by subculturing onto XLD agar (37°C for 24 h). Following incubation, typical colonies were subcultured onto nutrient agar (37°C for 24 h) and confirmed by biochemical test and molecular characterization by PCR assay. The combination of positive and negative results yielded a MPN data set.

MPN values were calculated using MPN data by Thomas’ equation in MS EXCEL data sheet developed by Division of Mathematics in FDA/CFSAN (Blodgett, 2006).

Molecular characterization of isolated strains using Polymerase chain reaction (PCR) assay

Genomic DNA of Salmonella spp. was extracted as per the protocol of Rawool et al., (2007). Primers for Salmonella organism was used according Rahn et al., (1992) for invA gene. Sequence of forward primer (invA) was GTGAAATTATCGCCACGTTCGGGCA A) and reverse primer was TCATCGCACCGTCAAAGGAACC). DNA samples were amplified in a total of 25 µl as the following: 2.5µl of 10x PCR Buffer, 1.5 µlNTP Mix (10mM), 2µl MgCl2 (50mM) 1.25µl of forward primer, 1.25µl of reverse primer, 0.50µl Taq polymerase (500U) 14.0µl of PCR grade water and 2 µl of the template.
The PCR was performed under the using conditions of primary denaturation: 94°C / 2 min., secondary denaturation: 94°C / 30 sec., annealing: 65°C / 1 min., extension: 72°C / 2 min., No. of cycles: 30 and final extension: 72°C / 5 min. Aliquots of amplified PCR products were electrophoresed in 1.5% agarose gel. The samples and a 100 bp DNA ladder were loaded in the wells in amount of 7µl of sample. A current of 90 V for 1 hour was passed on the horizontal electrophoresis unit. Specific amplicons were observed under ultraviolet transillumination compared with the marker. The gel was photographed by a gel documentation system and the data were analyzed.

Results and Discussion

Prevalence of Salmonella spp. from farm samples

Study revealed that 12 (16.66%) out of 108 samples were positive for Salmonella spp. in the environment of poultry farms located in vicinity of Mumbai city (Table: 01). There is significant difference (p=0.02) between sources in the farm and Salmonella occurrence. Results are comparable with study of Ahmed et al., (2014) who reported 11.1% of prevalence of Salmonella spp in the environment of broiler poultry farms of Khartoum, Sudan. Also Al-Zenki et al., (2007) who reported 5.4% prevalence from farm samples collected in Kuwait, this may be attributed hygienic measures applied. Kumar et al., (2014) in India reported 0%and 15.6% prevalence of Salmonella under intensive production system and free-range system, respectively.

This study showed that 6(50%) Salmonella spp. were isolated from litter. Salmonella from litter can lead to heavy contamination of the bird’s feathers and feet which increases the probability to recover the organism from carcasses in poultry processing plants due to fecal shedding onto the litter (Trampel et al., 2000). Results are in agreement with Scur et al., (2014) who observed 61.9 % prevalence of Salmonella spp. from litter samples. This study showed that there was a negative detection for Salmonella spp from feed and feeder swabs which confirm that use of heat treated feed material and proper storage conditions. Presence of Salmonella spp. in Drinker swabs (25.00%) and drinking water (16.66%) confirm that Salmonellae may originate either from faeces/litter or from water already contaminated by pathogenic organisms. The result pertaining to feed and water are opposite to the report of Alali et al., (2010) who has reported 27.5% and 0.00% prevalence of Salmonella from feed and water samples of conventional farms, respectively. El Hussein et al., (2010) who reported 7.23% prevalence from poultry drinking water which may be attributed to the variation in the numbers of collected samples.

Positive cloacal swabs (25.00%) and faeces (16.66%) indicate current infection in the flocks which is attributed to horizontal transmission from poultry environment. Dust in the poultry houses in large amount may also be a hazard, since dust has been recognized as a vehicle of transmission of Salmonella when large numbers of organisms are present (Harbaugh et al., 2006) a positive wall dust swab (8.33%) in our study confirms the same. The present result supports the report of Musa et al., (2014) and Corrique and Davies (2008) who reported that faeces/litter and dusts are the matrices of choice for Salmonella isolation and sources of cross contamination. This study also revealed that 01(8.3%) hand swabs was positive for Salmonella which confirms cross contamination. Similarly Ahmed et al., (2014) reported 01(5.6%) hand swab was positive for Salmonella.
The results are in agreement with Abunna et al., (2016), Marin et al., (2011) and AL-Iedani et al., (2014) who recovered Salmonella from various environmental samples mentioned in our study. Horizontal transmission can occur by direct bird-to-bird contact, ingestion of contaminated feces or litter, contaminated water, personnel, farm and personal equipment, and a variety of other sources (Nakamura et al., 1997; Nakamura et al., 1994 and Lahellec and Colin, 1985).

Variation in prevalence were reported by Agada et al., (2014) 10.9% in Nigeria, Al-Abadi and Al-Mayah, (2012) 9.2% in Iraq and Jahan et al., (2012) 45% in Bangladesh. Abbuna et al., (2016) stated that differences in prevalence might be due to the difference in study design, isolation technique, different in sample type and difference in geographical location.

**Prevalence of Salmonella spp. at chicken retail shop**

Over all 42 samples comprising of washing water, scalding water, swabs of worker hand, platform, chopping board, knife, and cloacal swab were analysed for Salmonella spp. Out of 42 samples 08 (19.04%) samples found positive. Surprisingly in used scalding water samples were negative, that might be because of high temperature of water. Out of individual 6 samples of all category 03 (7.14%) swab samples of chopping board found positive while one sample each from water and swab samples of worker hand, platform, knife, and cloaca were found positive. Olayinka, and Adeyanju (2014) reported 23.8, 11.90 and 0.00 per cent occurrence of Salmonella spp. from knives, weighing scales and wooden tables, respectively. Costerton et al., (1999) stated that Salmonella spp. noted as common contaminants of equipment used in processing of meat which are able to produce biofilm. Thiruppathi et al., (2004) observed Salmonella cross-contamination in retail chicken outlets in Chopping boards at (18.75%) and the butcher's hands (14.29%) followed by knives and the weighing balance tray. Study conducted by Ali et al., (2010) to find out microbial contamination of raw meat and its environment in retail shops in Karachi, Pakistan and reported 29% distribution of Salmonella in meat samples but zero detection of Salmonella from meat cutting surfaces (knives, wooden boards, weigh scales and meat mincers) and environmental surface swabs. Higher occurrence of Salmonella spp. in retail chicken shop is may be due lack of adherence to good hygienic practices and poor management practices on the farms.

A total of 24 swab samples were collected from chicken carcass at various chicken processing stages at retail shop along with 6 neck samples of eviscerated carcasses. Statistically non-significant difference was observed amongst different processing stages (p=0.43). Highest rate of contamination (50%) was observed in post defeathering and post evisceration stages of processing. While only one swab samples post bleeding and post scalding stages were positive. Out 06 post eviscerated carcass neck skin samples two samples were found positive, being at the lowest point in terms of gravity, neck skin may accumulate bacterial particles from run-off from washing (Table No: 2).The handling and processing of retail chicken needsto be improved to reduce the Salmonella incidence level in these stages along with washing of carcasses before and after evisceration. In similar study conducted by Morris and Wells (1970) at processing plants noticed 13.2% and 7% level of contamination after picking and after evisceration, respectively. Difference in the occurrence might be due to mechanical and non-mechanical processing operations. The level of Salmonella in live birds brought
for slaughter in retail markets might be very low but during processing under unhygienic stages carcasses were contamination by environmental sources. The previous studies as reported by other researchers only focused on the prevalence of *Salmonella* in chicken carcasses or chicken cuts and environment. This could explain the new way of detecting cross contamination of *Salmonella* which focused on the occurrence of *Salmonella* at different stages of retail chicken processing.

**Quantification of *Salmonella* spp. by miniature MPN technique**

As defeathering and evisceration are the major site of cross-contamination in poultry processing (Notermans *et al*., 1980; Clouser *et al*., 1995), each of 6 Samples at post defeathering and post evisceration stages of chicken processing were collected and subjected for quantification of *Salmonella* spp. by miniature MPN technique. Three samples each from post defeathering and post evisceration stages were positive with average log MPN count/10cm² of 1.88 and 2.11, respectively (Table :03). Shashidhar *et al*., (2011) observed *Salmonella* load in the range of 1.30 to 120 MPN/g in the retail chicken sample similarly Straver *et al*., (2007) have reported that the number of *Salmonella* on chicken filets varied from 1 to 3.81 log MPN per filets. Very little work has been done in India on the amount of this organism present on the carcasses during the processing stages. This is first attempt in India to quantify the *Salmonella* during chicken processing stages using miniature MPN method.

**Detection of invasive gene of virulent *Salmonella* (invA) using polymerase chain reaction (PCR)**

Out of 34 *Salmonella* isolates obtained from in this study 31 isolates showed positive amplification of 284 bp fragment specific for the invA gene (common gene) from examined samples with 91.17% detection level (fig. 1).

<table>
<thead>
<tr>
<th>Sr. No</th>
<th>Type of Sample</th>
<th>No of Samples Collected</th>
<th>No of Positive Samples</th>
<th>Per cent Prevalence</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Cloacal Swab</td>
<td>12</td>
<td>3</td>
<td>25.00</td>
</tr>
<tr>
<td>2</td>
<td>Feed</td>
<td>12</td>
<td>0</td>
<td>0.00</td>
</tr>
<tr>
<td>3</td>
<td>Drinking Water</td>
<td>12</td>
<td>2</td>
<td>16.66</td>
</tr>
<tr>
<td>4</td>
<td>Litter</td>
<td>12</td>
<td>6</td>
<td>50.00</td>
</tr>
<tr>
<td>5</td>
<td>Faeces</td>
<td>12</td>
<td>2</td>
<td>16.66</td>
</tr>
<tr>
<td>6</td>
<td>Worker Hand Swab</td>
<td>12</td>
<td>1</td>
<td>8.33</td>
</tr>
<tr>
<td>7</td>
<td>Wall Dust Swab</td>
<td>12</td>
<td>1</td>
<td>8.33</td>
</tr>
<tr>
<td>8</td>
<td>FeederSwab</td>
<td>12</td>
<td>0</td>
<td>0.00</td>
</tr>
<tr>
<td>9</td>
<td>DrinkerSwab</td>
<td>12</td>
<td>3</td>
<td>25.00</td>
</tr>
<tr>
<td>Total</td>
<td>108</td>
<td></td>
<td>18</td>
<td>16.66</td>
</tr>
</tbody>
</table>

(Poultry environmental samples *p* = 0.02)
Table 2 Occurrence of *Salmonella* Spp. isolated from retail chicken processing shop

<table>
<thead>
<tr>
<th>Sr. No.</th>
<th>Sample Source</th>
<th>Number of Samples Collected</th>
<th><em>Salmonella</em> Positive Samples</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td><strong>A) Environmental and other samples</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>1</td>
<td>Washing Water</td>
<td>6</td>
<td>1 (2.3)</td>
</tr>
<tr>
<td>2</td>
<td>Scalding water</td>
<td>6</td>
<td>0</td>
</tr>
<tr>
<td>3</td>
<td>Worker Hand</td>
<td>6</td>
<td>1 (2.3)</td>
</tr>
<tr>
<td>4</td>
<td>Carcass Contact Platform</td>
<td>6</td>
<td>1 (2.3)</td>
</tr>
<tr>
<td>5</td>
<td>Chopping Board</td>
<td>6</td>
<td>3 (7.14)</td>
</tr>
<tr>
<td>6</td>
<td>Knife Swab</td>
<td>6</td>
<td>1 (2.3)</td>
</tr>
<tr>
<td>7</td>
<td>Cloacal Swabs</td>
<td>6</td>
<td>1 (2.3)</td>
</tr>
<tr>
<td></td>
<td><strong>Total</strong></td>
<td><strong>42</strong></td>
<td><strong>08 (19.04)</strong></td>
</tr>
<tr>
<td></td>
<td><strong>B) Sampling at different Processing Stages</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>8</td>
<td>Post Bleeding</td>
<td>6</td>
<td>1 (16.66)</td>
</tr>
<tr>
<td>9</td>
<td>Post Scalding</td>
<td>6</td>
<td>1 (16.66)</td>
</tr>
<tr>
<td>10</td>
<td>Post Defeathering</td>
<td>6</td>
<td>3 (50.00)</td>
</tr>
<tr>
<td>11</td>
<td>Post Evisceration</td>
<td>6</td>
<td>3 (50.00)</td>
</tr>
<tr>
<td>12</td>
<td>Neck Sample of Post eviscerated carcass</td>
<td>6</td>
<td>2 (33.33)</td>
</tr>
<tr>
<td></td>
<td><strong>Total</strong></td>
<td><strong>30</strong></td>
<td><strong>10 (33.33)</strong></td>
</tr>
</tbody>
</table>

(Processing stages p = 0.43)

Fig.1 PCR products of 284 bp DNA fragment of *Salmonella* isolates

Lane 1 and 2: 284 bp PCR products of *Salmonella* isolates recovered from poultry farm and chicken retail shop samples; Lane 3: Standard *Salmonella* Typhimurium (MTCC 3224); Lane 4 (M): 100bp DNA Ladder
Table 3 Quantification of *Salmonella* spp. in pre and post evisceration swab samples in chicken processing

<table>
<thead>
<tr>
<th>Sr. No</th>
<th>Processing Stage</th>
<th>Samples</th>
<th>Number of Positive samples</th>
<th>Average log MPN Count/10cm²</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Post Defeathering</td>
<td>6</td>
<td>3</td>
<td>1.88</td>
</tr>
<tr>
<td>2</td>
<td>Post Evisceration</td>
<td>6</td>
<td>3</td>
<td>2.11</td>
</tr>
<tr>
<td>Total</td>
<td></td>
<td>12</td>
<td>06</td>
<td></td>
</tr>
</tbody>
</table>

Results are in agreement with Ohtsuka et al., (2005) who reported 90% detection of *Salmonella* by PCR, whereas less than Salehi et al., (2005), Ozbev and Ertas (2006) and Samaxa et al., (2012) who reported 100% detection of *Salmonella* spp. by PCR. This may be attributed to variation due to targeting different genes and the concentration of DNA template added to the PCR reactions.

In conclusion, this study revealed that the prevalence rate of *Salmonella* spp. in farms and chicken retail shops in Mumbai were 16.66% and 19.04% respectively. Highest prevalence of *Salmonella* spp. was noticed in litter samples followed by cloacal swabs and drinker swabs which would be considered as risk factors for cross contamination at farm level. Post defeathering and Post evisceration stages of processing are found critical stages of retail processing. Higher prevalence rate could be attributed to lack of adherence to good hygienic practices and poor management practices on the farms and retail chicken shop. Application of hygienic measures during farm management and processing stages may reduce the risk of contamination of carcasses.

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