Studies on the Effect of Time of Wedge Grafting and Growing Conditions on Growth and Leaf Characteristics of Guava (Psidium guajava L.) Grafts

L. Vanaja*, D.V. Swami, B. Prasanna Kumar and P. Subbaramamma

College of Horticulture, Dr. Y.S.R.Horticultural University, Venkataramannagudem, West Godavari District-534101, A.P., India

*Corresponding author

Abstract

An experiment was conducted on wedge grafting time, growing conditions on subsequent growth of grafted plants in guava (Psidium guajava L.) during 2014-2015. Wedge grafting was done during August (M1), September (M2), October (M3), November (M4), December (M5), January (M6) and February (M7) months under poly house and 50% shade net conditions. Wedge grafting in the month of August under poly house condition gave better response with respect to number of leaves per sprout, leaf area per sprout and leaf chlorophyll content.

Keywords
Guava, Grafting time, poly house and shade net conditions, Leaf characters, Wedge grafting.

Introduction

Guava (Psidium guajava L.), the poor man’s apple and “apple of tropics” belongs to the family Myrtaceae, is considered to be one of the exquisite, nutritionally valuable and remunerative crops. Besides its high nutritional value, it bears heavy crop every year and gives good economic returns. This has prompted several farmers to take up guava orcharding on a commercial scale. In recent years, guava is getting popularity in the international trade due to its nutritional value and processed products. Guava is highly cross pollinated crop and the seed being a heterozygous in nature, it is not possible to get true-to-type plants through seeds. However, the greatest handicap in guava plantation is discriminate multiplication of plants from unreliable sources by nurserymen (Singh et al., 2005). Non-availability of quality planting materials and consequent substitution of poor quality seedlings have adversely affected the guava production and productivity. The planting material is the best requirement for any fruit crop because it influences the ultimate yield both in terms of quality and quantity (Singh et al., 2005). The scenario is changing from traditional propagation with incorporation of science and technology to nursery management and trade. In view of the high return and the potential for processing, there is a tremendous scope for bringing substantial additional area under guava crop in India. So, a rapid and successful propagation technique is required.
as the area under crop is expanding and there is a demand to prepare the guava sampling throughout the year. While choosing a particular technique for propagation of guava, the time of wedge grafting and climatic conditions should be taken into consideration.

In the present context, use of rapid multiplication methods become very important when planting material was limited for improved varieties or clones. Though, guava is propagated through air-layering, stooling, grafting and inarching and still not commercially viable due to varying rates of success due to absence of tap root system and involving cumbersome process for its multiplication.

Keeping in view of these facts, the investigation was carried out on the response of wedge grafting under poly house and shade net conditions to ensure success of grafts growth and large scale multiplication of superior planting material in guava.

**Materials and Methods**

An experiment was carried during 2014-2015 at the Horticultural College and Research Institute, Venkataramannagudem, West Godavari District, Andhra Pradesh. Wedge grafting was performed during seven months, viz., August (M₁), September (M₂), October (M₃), November (M₄), December (M₅), January (M₆) and February (M₇) and grown under poly house as well as in 50% shade net conditions. There were fourteen treatment combinations replicated thrice in a Factorial Randomized Block Design. The grafting was done on 6 to 8-month old rootstock, which has attained a stem diameter of 0.5-1.0 cm. The scion shoots of 15-18 cm long of pencil thickness (0.5 to 1.0 cm) with 3 to 4 healthy buds were selected for grafting. Selected scions were defoliated on the mother plant, about one week prior to grafting. At the same time apical growing portion of selected shoots was also beheaded, which helped in forcing the dormant grafts to swell. In this way, the grafts on the scion were ready to start sprouting at the time of grafting. The scion stick was inserted into the split of the stock and pressed properly so that cambium tissue of rootstock and scion could come into contact with each other. The observations were recorded on leaf characters such as number of leaves per sprout, leaf area per sprout and leaf chlorophyll content.

**Number of leaves per sprout**

The number of leaves per successful graft sprout was recorded at 15 days interval starting from 30\(^{th}\) DAG to 120\(^{th}\) DAG for five randomly selected grafts in each replication. The average number of leaves per successful graft sprout were calculated and expressed in number.

**Leaf area per sprout (cm\(^2\))**

The leaf area per successful graft sprout was recorded at 15 days interval starting from 30\(^{th}\) DAG to 120\(^{th}\) DAG by using scale to measure the length and breadth of leaf for five randomly selected grafts in each replication.

The apparent leaf area obtained by multiplying length and breadth is reduced to actual leaf area by correcting with constant k. The average leaf area per sprout was calculated by employing the following formula as suggested by Watson (1952) and expressed in cm\(^2\).

\[ \text{Leaf area (A)} = k \times L \times B \]

Where

A = leaf area
L = Length of leaf
B = Breadth of leaf
k = 0.635 constant value

L and B are maximum length and breadth at right angle to each other respectively and k is
constant determined for species under investigation.

**Leaf chlorophyll content (SPAD units)**

The leaf chlorophyll content of successful grafts were estimated at 15 days interval starting from 30th DAG to 120 days by using SPAD chlorophyll meter for five randomly selected grafts in each replication. The average leaf chlorophyll content was calculated and expressed in SPAD units.

**Results and Discussion**

The number of leaves per sprout was more in the grafts prepared under shade net condition (1.85, 3.04, 5.62, 8.22, 10.72, 13.60, and 15.69 at 30, 45, 60, 75, 90, 105 and 120 DAG respectively) than under poly house condition. Significant differences were observed among the time of wedge grafting for number of leaves per sprout at all days after grafting. The number of leaves per sprout was gradually increased from 30 DAG to 120 DAG. The number of leaves per sprout was more in the plants grafted during January month (3.33, 5.18, 7.71, 10.88 and 14.23 at 30, 45, 60, 75, 90, 105 and 120 DAG respectively) followed by August month. The number of leaves per sprout was less in the plants grafted during September month (Table 1).

The interaction between growing conditions and time of wedge grafting was found significant for number of leaves per sprout at all days after grafting. The maximum number of leaves per sprout (3.40, 4.33, 7.46, 11.63, 16.70, 19.23 and 24.13 at 30, 45, 60, 75, 90, 105 and 120 DAG respectively) was observed in the grafts prepared during August month under poly house conditions. The minimum number of leaves per sprout was observed in the grafts prepared during September month under poly house conditions.

It could be due to auxins and prevailing favourable environmental conditions helps in the formation of more number of leaves per sprout through proper development of intercalary meristem at nodal regions (Taiz and Zeiger, 2012). These results are in agreement with the earlier findings of Singh and Singh, (2006) in guava; Syamal et al., (2012); Beer et al., (2013); Joshi et al., (2014) and Mahendra et al., (2015) in guava.

The leaf area per sprout (15.20, 17.77, 22.99, 25.77, 28.86, 32.73 and 36.01 cm² at 30, 45, 60, 75, 90, 105 and 120 DAG respectively) was high in plants grafted under shade net condition than under poly house condition.

Significant differences were observed among the time of wedge grafting for leaf area per sprout at all days after grafting. The leaf area per sprout (25.63, 31.94, 37.09, 44.47, 49.05 and 56.19 cm² at 45, 60, 75, 90, 105 and 120 DAG respectively expect 30 DAG) was high in the grafts prepared during August month followed by December month. The leaf area per sprout (2.75, 3.14, 3.31, 3.45, 3.53, 3.96 and 4.31 cm² at 30, 45, 60, 75, 90, 105 and 120 DAG respectively) was low in the plants grafted during September month (Table 2).

The interaction between growing condition and time of wedge grafting was found significant for leaf area per sprout at all days after grafting.

The maximum leaf area per sprout (26.45, 32.42, 39.55, 47.74, 58.51, 63.42 and 74.26 cm² at 30, 45, 60, 75, 90,105 and 120 DAG respectively) was observed in the grafts prepared during August month under poly house conditions. The minimum leaf area per sprout (2.55, 3.07, 3.18, 3.44, 3.52, 3.81 and 4.15 cm² at 30, 45, 60, 75, 90,105 and 120 DAG respectively) was observed in the grafts prepared during September month under poly house condition.
### Table 1: Effect of growing conditions, time of wedge grafting and their interaction on number of leaves per sprout in guava

<table>
<thead>
<tr>
<th>Treatments</th>
<th>30 DAG</th>
<th>45 DAG</th>
<th>60 DAG</th>
<th>75 DAG</th>
<th>90 DAG</th>
<th>105 DAG</th>
<th>120 DAG</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>C1</td>
<td>C2</td>
<td>Mean</td>
<td>C1</td>
<td>C2</td>
<td>Mean</td>
<td>C1</td>
</tr>
<tr>
<td>M_1</td>
<td>1.13</td>
<td>3.40</td>
<td>2.26</td>
<td>2.70</td>
<td>4.33</td>
<td>3.51</td>
<td>5.90</td>
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<tr>
<td>M_2</td>
<td>0.17</td>
<td>0.10</td>
<td>0.13</td>
<td>0.30</td>
<td>0.17</td>
<td>0.23</td>
<td>0.47</td>
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<tr>
<td>M_3</td>
<td>1.00</td>
<td>0.80</td>
<td>0.90</td>
<td>2.30</td>
<td>2.23</td>
<td>2.26</td>
<td>5.16</td>
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<tr>
<td>M_4</td>
<td>1.30</td>
<td>0.53</td>
<td>0.91</td>
<td>2.96</td>
<td>0.90</td>
<td>1.93</td>
<td>6.13</td>
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<tr>
<td>M_5</td>
<td>2.10</td>
<td>2.70</td>
<td>2.40</td>
<td>2.50</td>
<td>3.63</td>
<td>3.06</td>
<td>6.53</td>
</tr>
<tr>
<td>M_6</td>
<td>5.16</td>
<td>1.50</td>
<td>3.33</td>
<td>7.16</td>
<td>3.20</td>
<td>5.18</td>
<td>8.76</td>
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<tr>
<td>M_7</td>
<td>2.10</td>
<td>0.73</td>
<td>1.41</td>
<td>3.43</td>
<td>1.76</td>
<td>2.60</td>
<td>6.56</td>
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<tr>
<td>Mean</td>
<td>1.85</td>
<td>1.42</td>
<td>3.04</td>
<td>2.34</td>
<td>5.62</td>
<td>4.29</td>
<td>8.22</td>
</tr>
<tr>
<td>C.D (5%)</td>
<td>0.22</td>
<td>0.15</td>
<td>0.17</td>
<td>0.08</td>
<td>0.23</td>
<td>0.11</td>
<td>0.14</td>
</tr>
<tr>
<td>SE(m)</td>
<td>0.05</td>
<td>0.05</td>
<td>0.15</td>
<td>0.05</td>
<td>0.17</td>
<td>0.08</td>
<td>0.23</td>
</tr>
</tbody>
</table>

**Factor –1**

C_1 = Shade net  
C_2 = Polyhouse

**Factor –2**

M_1 = August  
M_2 = September  
M_3 = October  
M_4 = November

**DAG**

Days After Grafting

### Table 2: Effect of growing conditions, time of wedge grafting and their interaction on leaf area per sprout in guava

<table>
<thead>
<tr>
<th>Treatments</th>
<th>30 DAG</th>
<th>45 DAG</th>
<th>60 DAG</th>
<th>75 DAG</th>
<th>90 DAG</th>
<th>105 DAG</th>
<th>120 DAG</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>C1</td>
<td>C2</td>
<td>Mean</td>
<td>C1</td>
<td>C2</td>
<td>Mean</td>
<td>C1</td>
</tr>
<tr>
<td>M_1</td>
<td>14.24</td>
<td>26.45</td>
<td>20.34</td>
<td>18.85</td>
<td>32.42</td>
<td>25.63</td>
<td>24.33</td>
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<tr>
<td>M_2</td>
<td>2.96</td>
<td>2.55</td>
<td>2.75</td>
<td>3.22</td>
<td>3.07</td>
<td>3.14</td>
<td>3.44</td>
</tr>
<tr>
<td>M_5</td>
<td>19.45</td>
<td>24.45</td>
<td>21.95</td>
<td>21.33</td>
<td>26.43</td>
<td>23.88</td>
<td>28.68</td>
</tr>
<tr>
<td>M_7</td>
<td>18.24</td>
<td>8.98</td>
<td>13.61</td>
<td>20.83</td>
<td>10.45</td>
<td>15.64</td>
<td>26.94</td>
</tr>
<tr>
<td>Mean</td>
<td>15.20</td>
<td>13.52</td>
<td>17.77</td>
<td>16.13</td>
<td>22.99</td>
<td>19.00</td>
<td>25.77</td>
</tr>
<tr>
<td>C.D (5%)</td>
<td>0.22</td>
<td>0.15</td>
<td>0.17</td>
<td>0.08</td>
<td>0.23</td>
<td>0.11</td>
<td>0.14</td>
</tr>
<tr>
<td>SE(m)</td>
<td>0.05</td>
<td>0.05</td>
<td>0.15</td>
<td>0.05</td>
<td>0.17</td>
<td>0.08</td>
<td>0.23</td>
</tr>
</tbody>
</table>

**Factor –1**

C_1 = Shade net  
C_2 = Polyhouse

**Factor –2**

M_1 = August  
M_2 = September  
M_3 = October  
M_4 = November

**DAG**

Days After Grafting
Table 3: Effect of growing conditions, time of wedge grafting and their interaction on leaf chlorophyll content in guava

<table>
<thead>
<tr>
<th>Treatments</th>
<th>Leaf chlorophyll content (SPAD Units)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>30 DAG</td>
</tr>
<tr>
<td></td>
<td>C₁</td>
</tr>
<tr>
<td>M₁</td>
<td>9.40</td>
</tr>
<tr>
<td>M₂</td>
<td>1.77</td>
</tr>
<tr>
<td>M₆</td>
<td>14.20</td>
</tr>
</tbody>
</table>

SE(m) C.D (5%) SE(m) C.D (5%) SE(m) C.D (5%) SE(m) C.D (5%) SE(m) C.D (5%) SE(m) C.D (5%) SE(m) C.D (5%) SE(m) C.D (5%) SE(m) C.D (5%) SE(m) C.D (5%)

Factor –1 0.25 0.74 0.29 0.84 0.33 0.97 0.37 1.09 0.41 1.20 0.42 1.24 0.44 1.29
Factor –2 0.47 1.38 0.54 1.56 0.62 1.81 0.70 2.03 0.77 2.24 0.79 2.31 0.82 2.41
Interaction (1×2) 0.67 1.95 0.77 2.21 0.88 2.56 0.98 2.87 1.08 3.17 1.12 3.27 1.17 3.41

Factor -1 C₁ = Shade net C₂ = Polyhouse
Factor –2 M₁ = August M₂ = September M₃ = October M₄ = November
M₅ = December M₆ = January M₇ = February
DAG Days After Grafting

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The leaf chlorophyll content (9.71, 11.40, 13.61, 15.22, 18.77, 21.03, 22.65 SPAD units at 30, 45, 60, 75, 90, 105 and 120 DAG respectively) was more in the grafts prepared under shade net condition than under poly house condition (7.96, 9.35, 10.98, 12.58, 15.44, 17.03 and 18.63 SPAD units at 30, 45, 60, 75, 90, 105 and 120 DAG respectively).

The differences among the time of wedge grafting were found significant for leaf chlorophyll content. The maximum leaf chlorophyll content (13.28, 15.35, 18.57, 21.40, 26.48, 31.10 SPAD units at 30, 45, 60, 75, 90, 105 and 120 DAG respectively) was noticed in the grafts prepared during August month followed by December month. The minimum leaf chlorophyll content (1.82, 2.08, 2.43, 2.72, 2.98, 3.08 and 3.20 SPAD units at 30, 45, 60, 75, 90, 105 and 120 DAG respectively) was observed in the grafts made during September month (Table 3).

The interaction between growing conditions and time of grafting was found significant for leaf chlorophyll content. The leaf chlorophyll content (17.17, 20.10, 24.43, 28.40, 34.70, 38.40 and 40.00 SPAD units at 30, 45, 60, 75, 90,105 and 120 DAG respectively) was more in the grafts prepared during August month under poly house conditions. The leaf chlorophyll content (1.77, 2.03, 2.40, 2.67, 2.87, 2.93 and 3.07 SPAD units at 30, 45, 60, 75, 90,105 and 120 DAG respectively) was less in the grafts made during September month under shade net condition.

A perusal of the results revealed that, grafts prepared in the month of August under poly house condition recorded the maximum number of leaves per sprout, leaf area per sprout and leaf chlorophyll content. This might be due to the fact that warmer and humid air inside the polyhouse induces the soil to warm up. Plants grafted in the month of August showed better performance which might be due to favourable conditions (moderate humidity i.e. from 25% to75% and optimum temperature i.e. from 24ºC to 35ºC) adequate supply of soil moisture and availability of dormant and swollen terminal buds of scion in bulging condition, which encouraged the maximum growth. These results are in agreement with the results of Singh and Singh (2006) in guava; Lal et al., (2007) in kiwi fruit; Mir and Kumar (2011) in walnut; Raghavendra et al., (2011) in wood apple; Beer et al., (2013) in guava; Syamal et al., (2012) in guava; Anushma et al., (2014) in jamun; Sivudu et al., (2014) in mango; Joshi et al., (2014) and Mahendra et al., (2015) in guava. On the basis of results obtained from the present investigation, it can be concluded that the best results were obtained in the plants grafted during August month under poly house condition is more favorable for maximum number of leaves per sprout, leaf area per sprout and leaf chlorophyll content.

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