Comparative Evaluation of Microscopy and ELISA in Diagnosis of Cryptosporidiosis in HIV and Non-HIV Patients

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**Abstract**

**Keywords**

-Cryptosporidium parvum
-Immunocompromised patients.

**Introduction**

Cryptosporidium species are coccidial protozoan parasites classified as emerging pathogens by the Centre for Disease and Prevention (CDC). These protozoan parasites inhabit the intestinal and respiratory surface of mammals, birds and reptiles and causes human clinical syndrome of cryptosporidiosis (Jayalakshmi et al., 2008; Parisi et al., 1995).

Cryptosporidiosis is a worldwide infection with prevalence rates being higher in developing (5 to 10%) than in developed (1 to 3%) countries. It is more common in young children (ages 1 to 4 years) than in adults, with the exception of patients with AIDS, and occurs more frequently in the warmer months of the year (Rosenblatt, 1993).

Organisms of the genus Cryptosporidium are small (2-4 μm) coccidian parasites which have only recently been recognized as important enteric pathogens of humans. Although at least 8 recognised species of Cryptosporidium are considered valid, human cryptosporidiosis is primarily attributed to C.parvum (Michael, 2002).

The common clinical manifestation of Cryptosporidiosis is profuse and watery diarrhoea often containing mucus but rarely blood and leucocytes. Other clinical
symptoms include abdominal cramps, low grade fever, nausea and vomiting. Infection in immune competent host is self limited with duration of approximately 2 weeks. In AIDS patients Cryptosporidiosis generally causes chronic, severe and debilitating voluminous watery diarrohea. It may be accompanied by severe abdominal cramps, weight loss, anorexia, malaise, low grade fever (Hellard et al., 2003).

_Cryptosporidium oocysts_ may be hard to detect except in laboratories with the repeated and continuous exposure to positive specimens that allows diagnostic proficiency. Alternatively, oocyst antigen capture methods such as enzyme immune assays (EIAs) or immunochromatographic-lateral flow (ICLF) assays may be used, and positive reactions must be confirmed by using a suitable confirmatory test. Due to tiny size of these oocysts, differential staining using the modified Ziehl-Neelsen technique & wet mount preparation method have limited value for the detection of _Cryptosporidium_ in faecal samples where oocysts can easily be confused with other material present in the samples (Beauty).

Recently, several groups have described the use of enzyme-linked immunosorbent assays (ELISA) for the diagnosis. Enzyme-linked immunosorbent assays (ELISAs) have been reported to be up to 10 times more sensitive than acid-fast staining, making the ELISA method currently the “gold standard” for antigen detection in infected stool samples.

**Materials and Methods**

The study was conducted in the department of microbiology at JSS hospital over a period of ten months from March to December 2013. A total of 100 subjects were included in the study which were divided into 4 groups

- **GROUP 1** - HIV positive patient with diarrohea - 25 subjects
- **GROUP 2** - Asymptomatic HIV positive patients - 25 subjects
- **GROUP 3** - HIV negative patient with diarrohea - 25 subjects
- **GROUP 4** - Healthy Individuals as controls - 25 subjects.

The cases of bacterial diarrohea were excluded. Patients were provided with a clean, wide mouthed container for collection of stool samples with the instruction to transfer them immediately to the microbiology laboratory. All the stool samples were processed for saline and wet mount and examined under the microscope for the presence of protozoal cysts and trophozoites. A second smear was made after the concentration with formalin ether method and stained by modified Kinyoun’s acid fast stain and examined under oil immersion lens to detect oocysts of _Cryptosporidium parvum_.

Same samples were processed further by ELISA as per manufacturers guidelines (Savyon Diagnostics Ltd. Israel)

**Results and Discussion**

Out of 100 stool samples examined _Cryptosporidium_ antigen was detected in 9 (9%) by ELISA, in which it was categorised as follows in Group 1 - 05, Group 2 - 01 and in Group 3 – 03 . No antigen was detected in the control group as shown in Table 1.

_Cryptosporidium_ was found in all age group, the highest number of patients reported during the study belonged to the age group between 21-30 years (30%) . Table 2

_Cryptosporidium_ were detected in 9 out of 100 stool samples by ELISA, while 1 was detected by wet mount. 5 were detected by modified acid fast staining. Out of 9 sample which were positive by ELISA only 5 were positive for microscopy and only one sample was found to be positive by all the three methods. We found detection of cryptosporidial antigen was better with
ELISA in comparison with other methods (Table 3) The other parasitoligical profile showed *Isopora belli* as the second most common pathogen in HIV patients (4) and (2), in non HIV patients. An equal distribution of Cysts *Blastocystis hominis* was found in both HIV and non-HIV. Eggs of hook worm, *Entamoeba histolytica* cysts were detected. Cysts of Giardia were also found both in HIV(2) and non HIV patients(4). Coinfection with *Cryptosporidium* and *Isospora belli* were seen in 10 symptomatic HIV patients (GROUP 1). 5 had coinfection in HIV negative patients with diahorrea (GROUP 3) as shown in Table 4.

Cryptosporidiosis is a common gastrointestinal disease, and it has been recognized worldwide as a common cause of diarrhea. The disease is widespread in many developed and developing countries (Chalmers et al., 2003). According to the World Health Organization, more than 33% of global deaths are due to parasitic diseases. Intestinal parasitic infections are among the most common infections in the world responsible for mortality and morbidity.

The diagnosis of Cryptosporidiosis is essential in immunocompromised patients, as it may interfere with therapeutical procedures. Because of the difficulty and time-consuming nature of conventional microscopy examination for the detection of *Cryptosporidium* oocysts, there is a need for a simple and rapid test for the coprodiagnosis of *Cryptosporidium* infection (Joseph et al., 2001).

In the present study, stool samples from each subject were collected and screened for the presence of *Cryptosporidium* by wet mount, modified acid fast staining and ELISA.

Our study showed significant number of *Cryptosporidium* infection in HIV patients (28%) than in non HIV patients (6%). The study conducted by Jayalakshmi et al., reported 12.4% in HIV patients. Satheesh kumar et al., & Singh et al., reported 10.8 to 43% of *Cryptosporidium* infection in various parts of the country. Kumar et al. has reported *Cryptosporidium* as the commonest parasite which was associated with HIV infections in Southern parts of India (Kumar et al., 2002). Our study also projects *Cryptosporidium* as the most common coccidian parasite which was associated with HIV positive subjects. Studies conducted by Basak et al., and Mohandas et al., have also reported similar results (2010; 2002).

The ELISA test is simple and rapid to use and offers a less subjective method than microscopy for detecting this protozoan in faecal samples submitted to a busy diagnostic laboratory. This is a highly sensitive and specific technique, and it is useful for screening large numbers of specimens in a short time period. Also, it does not rely on microscopy skills (Cavasini et al., 2009).

The present study also evaluated different methods of detection of *Cryptosporidium* in stool samples by microscopy - wet mount, modified acid fast staining, and ELISA. In our study *Cryptosporidium* was found to be positive in 1 (11.1%) sample by all the 3 methods, 5 samples by ELISA & modified acid fast staining (55.5%) & 3 samples found to be positive only by ELISA (33.3%). All the positive samples were detected by ELISA (100%)

Hence detection of *Cryptosporidium* antigen by ELISA is simple and rapid. Our results show that the ELISA method, in addition to having excellent specificity, exhibits vastly improved sensitivity over those of the routine methods used. ELISA proved to be the most sensitive (100%) technique indicating the presence of *Cryptosporidium*.
Table. 1 Total Cryptosporidium Positive among HIV & non-HIV

<table>
<thead>
<tr>
<th>Patient Criteria</th>
<th>Total Positive</th>
</tr>
</thead>
<tbody>
<tr>
<td>HIV positive patient with diarhoea</td>
<td>5</td>
</tr>
<tr>
<td>Asymptomatic HIV positive patients.</td>
<td>1</td>
</tr>
<tr>
<td>HIV negative patient with diarhoea.</td>
<td>3</td>
</tr>
</tbody>
</table>

Table. 2 Age Distribution: Association of age with rate of Cryptosporidium infection

<table>
<thead>
<tr>
<th>Age in years</th>
<th>Cryptosporidium in HIV &amp; non HIV patients</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Negative</td>
</tr>
<tr>
<td>≤ 20</td>
<td>7(7.7%)</td>
</tr>
<tr>
<td>21-30</td>
<td>27(29.7%)</td>
</tr>
<tr>
<td>31-40</td>
<td>26(28.6%)</td>
</tr>
<tr>
<td>41-50</td>
<td>18(19.8%)</td>
</tr>
<tr>
<td>51-60</td>
<td>8(8.8%)</td>
</tr>
<tr>
<td>&gt;60</td>
<td>5(5.5%)</td>
</tr>
<tr>
<td>Total</td>
<td>91(100%)</td>
</tr>
</tbody>
</table>

Table. 3 Comparison of methods of detecting Cryptosporidium in HIV and non-HIV cases

<table>
<thead>
<tr>
<th>Methods</th>
<th>Number (n=100)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Wet mount</td>
<td>01(1%)</td>
</tr>
<tr>
<td>Modified ZN staining</td>
<td>06(6%)</td>
</tr>
<tr>
<td>ELISA</td>
<td>09(9%)</td>
</tr>
</tbody>
</table>

Table. 4 Various parasites detected:

<table>
<thead>
<tr>
<th>Parasites detected</th>
<th>HIV</th>
<th>Non-HIV</th>
</tr>
</thead>
<tbody>
<tr>
<td>Eggs of Hookworm.</td>
<td>1</td>
<td>2</td>
</tr>
<tr>
<td>Giardia.</td>
<td>2</td>
<td>4</td>
</tr>
<tr>
<td>Trichuris trichiura.</td>
<td>0</td>
<td>1</td>
</tr>
<tr>
<td>Blastocystis Hominis</td>
<td>1</td>
<td>1</td>
</tr>
<tr>
<td>Entamoeba histolytica</td>
<td>0</td>
<td>1</td>
</tr>
<tr>
<td>Isospora belli</td>
<td>4</td>
<td>2</td>
</tr>
<tr>
<td>Cryptosporidium</td>
<td>6</td>
<td>3</td>
</tr>
</tbody>
</table>
**Fig.1** Modified ZN Staining Showing *Cryptosporidium* oocyst

**Fig.2** ELISA Test strip for *Cryptosporidium* Antigen in Stool Samples

A 1 – POSITIVE CONTROL
B 1 – NEGATIVE CONTROL
E 1
A 4 \} POSITIVE TEST SAMPLE
Fabiana Rangel Marques et al conducted a study and reported that, among ninety-four paired samples analyzed the overall sensitivity of the immunoenzymatic assay was 100%, with a specificity of 96%; positive and negative predictive values were 89% and 100%, respectively (Cavasini et al., 2009). Jayalakshmi et al., found ELISA to be a simple, reliable and less subjective test which could be very useful in routine diagnosis and for screening a large number of specimens in short time, particularly in large-scale epidemiological surveys. Barua et al., found Modified acid fast technique to be efficacious method for routine screening of fecal specimens for Cryptosporidium spp in comparison with ELISA (Barua et al., 2008). But in our study ELISA was found to be more efficacious in the detection of Cryptosporidium antigen, as only 05 were detected by modified acid fast staining out of 09 positive which were detected by ELISA.
Along with Cryptosporidium spp other parasites like Isospora belli, Giradia, Cysts of Blastocystis, Eggs of Hookworm, Entamoeba histolytica were also detected

However ELISA is the most preferred method in the laboratories of the developed countries due to its high specificity and sensitivity, easy usage, fast application and scoring, and easy standardization for determination of Cryptosporidium antigens in stool samples (Hasan, 2008).

In conclusion, cryptosporidiosis is an important water borne disease. The study highlights the importance of evaluation of HIV infected as well as non HIV patients with diarrhoea. For intestinal protozal infection which may help in better management. Based on our data ELISA was found to be more sensitive and specific hence, ELISA is an useful assay for ruling out Cryptosporidiosis in immunocompromised individuals.

References


Fabiana Rangel Marques, Luciana Ventura Cardoso, Carlos Eugênio Cavasini, Magali Carmem de Almeida, Nair Aparecida Bassi, Margarete Teresa Gottardo de Almeida, Andréa Regina Baptista Rossitl and Ricardo Luiz Dantas Machado; Performance of an Immunoenzymatic Assay for Cryptosporidium Diagnosis of Fecal Samples; The Brazilian J. Infectious Dis., 9(1): 3-5.


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