

Original Research Article

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The spectrum of Different Pathogens causing Ear Infections and its Antimicrobial Sensitivity Patterns in Patients from Gurgaon, Haryana state, India

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Ear infections can occur in all age groups. Otitis media is one of the most common illness in ENT practice and occurring all over the world. It may even cause hearing impairment especially neural deafness if it is not treated beneath expert care. The study was done to know the spectrum of different pathogens and its antimicrobial sensitivity causing ear infections among all age groups. A total of 295 ear samples were collected and processed according to standard microbiological procedures. Gram negative bacteria were isolated more frequently as correspond to gram positive (47.24 and 43.72% respectively). The frequency of gram positive bacterial isolates was *Staphylococcus aureus* (37.2%), *Streptococcus pyogenes* (3.0%), *Streptococcus pneumoniae* (2.5%) and *Enterococcus spp.* whereas the frequency of gram negative bacterial isolates was *Pseudomonas aeruginosa* (34.2%) followed by *Escherichia coli* (4.0%), *Proteus mirabilis* (4.0%), *Klebsiella pneumoniae* (2.5%) and *Enterobacter aerogenes* (2.0%). The most frequent fungal isolates were *Aspergillus spp.* (5.0%) followed by *Candida spp.* (4.0%). Continuous knowledge of the pathogens responsible for ear infections and choice of suitable antibiotics according to sensitivity test results will guide the clinician for the treatment thus reducing the complication of ear infections and curbing resistant to antibiotics.

Introduction

The Ear is a vital sensory organ which is dependable for hearing and also maintaining balance, it's separated into the outer, middle and inner ear with the outer and middle regions being most vulnerable to injury and infections. Ear infection (medically termed Otitis Media) generally refers to an infection of the middle part of the ear that lies behind the eardrum. It is commendable to note that ear infections are a very common problem worldwide (Richard *et al.*, 1998).

About 65-330 million people suffer from ear infection worldwide and 60% of them had significant hearing impairment. The discharging ear is a common problem in tropics. It is seen in all age groups but is more common in infants and children. It's falling incidence during and after an adolescent stage is the result of the growth and development of the pharynx. But still it remains one of the major problems of adults attending the Ear, Nose and Throat (ENT) clinics (Raakhee *et al.*, 2014).

Chronic Suppurative Otitis Media (CSOM) is chronic inflammation of the middle ear, which affects the tympanic membrane, middle ear mucosa and other middle ear structures. Clinically, CSOM presents with ear discharge and conductive deafness (Iqbal *et al.*, 2009).

CSOM is the one of the most frequent problems related to ear in developing and developed nations if left untreated causing more severe loss of hearing. It is characterized by persistent otorrhea for more than 6-12 weeks, through the perforated tympanic membrane, usually resulting from earlier acute infection (Kristo *et al.*, 2011).

The infection is due to the ear cleft, resultant in ear discharge (Srivastava *et al.*, 2011). The WHO defines CSOM as 'otorrhea through a perforated tympanic membrane present for at least two weeks (WHO, 1986).

Most commonly isolated aerobic bacteria in ear infections are *Staphylococcus aureus*, *Streptococcus pyogenes*, *Pseudomonas aeruginosa*, *Escherichia coli*, *Proteus spp.*, *Klebsiella spp.*

Fungal infection of the middle ear and external auditory meatus are common as fungi thrive well in moist pus and mostly isolated fungi are *Aspergillus species* and *Candida species*. But the type of organism isolated varies between the geographical area and other factors (Srivastava *et al.*, 2011). CSOM is a global problem and affects all ages but common in children younger than 7 years due to horizontal, wider and short Eustachian tube' (Haraldsson *et al.*, 2004). The normally occurring symptoms are ear discharge, deafness, itching, pain and every so often fever. If it is left untreated complications like loss of hearing, post aural swelling and post aural sinus may occur (Oguntibeju, 2003).

CSOM has received considerable attention, not only because of its high incidence and chronicity but also because of issues such as bacterial resistance and ototoxicity with both topical and systemic antibiotics (Haynes, 2011).

Since CSOM is a disease which can cause major morbidity, knowledge of pathogens responsible can assist in the selection of the most appropriate treatment regimen and can reduce complications that may require surgical treatment. This study would be the first study in this area that provides original data about the pathogen antibiotic sensitivity profile which is likely to assist physicians in empirical therapy and setting therapeutic protocols.

Materials and Methods

Study Design, period and area

This prospective study was conducted from discharging ears specimens submitted for aerobic culture and sensitivity during the period of 6 months (November 2015 - April 2016) referred to Department of Microbiology & Molecular Biology at Modern Diagnostic & Research Centre, Gurgaon, Haryana -India.

Collection and transport of specimen

Samples of patients having discharging ear were collected with the help of sterile cotton swab in screw capped polypropylene tubes (Hi-Media) from all age groups using aseptic techniques and transported immediately to the microbiology laboratory.

Microscopic Examination (Gram stain)

The smear was prepared as per standard procedures for gram staining and examined

for the presence of pus cells, bacteria and/or fungal elements. The result of the gram stain was informed to the ENT specialist within 2 hours of the collection of the swab in the department.

Culture and identification

Ear swabs were cultured on sheep blood agar (Biomerieux) & Mac Conkey agar (Hi-Media) with sterilized nichrome loop. All cases showing growth on culture media after 24 hours of incubation at 37⁰C were identified using colony morphology, gram staining and key biochemical battery of tests with standard procedures.

Quality Control

The bacterial suspension was prepared and was adjusted to a 0.5 McFarland solution (Hi-Media). American Type Culture Collection (ATCC) standard reference strains (*Pseudomonas aeruginosa* ATCC-27853, *Staphylococcus aureus* ATCC 25923, *Escherichia coli* ATCC-25922) were used as a quality control throughout the study for culture and antimicrobial susceptibility testing. Antibiotic and media quality control was performed on receiving of each lot and on a weekly basis. The antibiotic failed in new lot verification process was rejected.

Antibiotic Sensitivity testing

The organism was processed for antibiotic sensitivity testing by using Kirby-Bauer disc diffusion method on Mueller-Hinton agar using selected antibiotics panel according to Clinical and Laboratory Standard Institute (CLSI., 2015).

Antibiotic sensitivity pattern for bacterial isolates - Amoxicillin-clavulanate, piperacillin-tazobactam, cefepime, ceftriaxone, cefuroxime, ceftazidime,

cefexime, imipenem, gentamicin, amikacin, tetracycline, ciprofloxacin, levofloxacin, ofloxacin & cotrimoxazole were tested against *Enterobacteriaceae*. piperacillin-tazobactam, ticarcillin-clavulanate, ceftazidime, cefepime, aztreonam, imipenem, colistin, polymyxin B, gentamicin, amikacin, ciprofloxacin, levofloxacin & ofloxacin were tested against *Pseudomonas aeruginosa*. Penicillin, vancomycin (E-Test), teicoplanin, gentamicin, amikacin, erythromycin, clindamycin, tetracycline, ciprofloxacin, levofloxacin, ofloxacin, cotrimoxazole, chloramphenicol, linezolid & ceftazidime were tested against *Staphylococcus spp.* Ampicillin, penicillin, ofloxacin, vancomycin, teicoplanin, erythromycin, tetracycline, ciprofloxacin, levofloxacin, chloramphenicol, linezolid & high-level gentamicin were tested against *Enterococcus spp.* Penicillin, vancomycin, erythromycin, azithromycin, tetracycline, levofloxacin, ofloxacin, cotrimoxazole, chloramphenicol, clindamycin & linezolid were tested against *Streptococcus pneumoniae*. Penicillin, Ampicillin, cefepime, ceftriaxone, vancomycin, erythromycin, azithromycin, tetracycline, levofloxacin, ofloxacin, chloramphenicol, clindamycin and linezolid were tested against *Streptococcus pyogenes*. Antifungal sensitivity was not included in the study.

Results and Discussion

In this study, a total of 295 patients having ear discharge & suspected of otitis media were included in this study. Of them, 160 (54.2%) were males and 135 (45.8%) were female patients with the ratio of 1.19:1. Overall, 194 (65.8%) of patients with ear infection had culture positive. The proportion of ear infection with positive culture was 110 (56.70%) in males & 84 (43.30 %) in females with the ratio of 1.31:1. (Table 1)

The majority of the patients who had ear discharge were between <10 years of age. This was followed by the age group between 21-30, 11-20, 31-40, 41-50 and >50. Of them, the patients who had an ear infection with the positive culture were from age group was 31-40, 41-50, <10, 21-30, >50, 11-20. (Table 2).

Out of 194 patients, there were 199 isolates. One eighty-nine patients (64.1 %) had an infection from the single organism, five patients (1.7%) had an infection with two organisms and remaining one hundred one (34.2%) had a sterile culture with no organisms isolated. (Table 3).

Of 199 isolates, 181 (92.0%) were bacteria and 18 (9.0 %) were fungal isolates. Gram negative bacteria were isolated more frequently as compare to gram positive (47.24 and 43.72% respectively). The frequency of gram positive bacterial isolates was *Staphylococcus aureus* (37.2%), *Streptococcus pyogenes* (3.0%), *Streptococcus pneumoniae* (2.5%) and *Enterococcus spp.* whereas the frequency of gram negative bacterial isolates was *Pseudomonas aeruginosa* (34.2%) followed by *Escherichia coli* (4.0%), *Proteus mirabilis* (4.0%), *Klebsiella pneumoniae* (2.5%) and *Enterobacter aerogenes* (2.0%). The most frequent fungal isolates were *Aspergillus spp.* (5.0%) followed by *Candida spp.* (4.0%). (Table 4)

Resistance pattern of gram negative bacterial isolates was *Pseudomonas aeruginosa* (2.9-17.4%), *Escherichia coli* (12.5-37.5%), *Enterobacter aerogenes* (0-25.0), *Proteus mirabilis* (12.5-75.0%) and *Klebsiella pneumoniae* (20.0-60.0%) (Table 5).

Resistance pattern of gram positive isolates to the antibiotics tested was *Staphylococcus aureus* (1.4-93.2 %), *Streptococcus pyogenes* (16.7-50.0%), *Streptococcus*

pneumoniae (20.0-40.0 %), *Enterococcus spp* (0-50%). (Table 6)

The present study would be the first study that attempts to isolate and characterize the probable pathogens among ear infections from the patients of all age groups and their antibiotic sensitivity profile from Gurgaon, Haryana State.

Ear infection is the most frequent disease for patients to visit clinician and take antibiotics (Grevers., 2010). The most frequent symptoms of ear infections are ear discharge (otorrhea), ear pain (otalgia), hearing loss, vertigo, and tinnitus (Cheng *et al.*, 1997). In our study male (54.2%) predominance was higher than female (45.8%). This observation was parallel with the finding of the few authors (Kumar *et al.*, 2011; Lodhi *et al.*, 2010; Yousuf *et al.*, 2011).

In this study, out of 194 patients with positive culture report, there were 199 isolates. The analysis revealed that 189 (64.1%) had an infection from the single organism, 51 (1.7%) had an infection with two organisms and remaining 101 (34.2%) had a sterile culture with no organism isolated. Matching figures reported by other authors vary significantly. Aslam *et al.*, 2004 from Pakistan, in their study on 142 samples revealed that 76% of them were pure and 23.9% were mixed culture and only 2.1% fungi whereas Poorey *et al.*, 2002 from India, in their study on 100 samples found pure growth from 82, mixed growth from 10 and no growth in 8 samples. The difference in results of a variety of authors could have been due to a difference in the patient population studies and geographical variations.

The clinical findings of patients with otitis media occurrence and peak age prevalence (mostly in children younger than 10 years of age) in our study are similar to findings of

previous studies done in Ethiopia and other developing countries (Melaku *et al.*, 1999; Tobhi *et al.*, 2006).

The high incidence rate in children is may be due to numerous reasons as young children and infants may have low resistance and also because of the relatively short Eustachian tube. Due to the short Eustachian tube, infected material from the nose, adenoids and sinuses passes more readily along the Eustachian tube to the tympanic cavity, particularly during coughing, sneezing, vomiting and forced feeding commonly practiced in our environment with the child's nose blocked, while being held head down and half prone (Nwabuisi *et al.*, 2002). It may be attributed to the fact that they are more prone to upper respiratory tract infection. Poor hygiene and unorthodox approach to the treatment like unconventional ear drops and concoctions such as oil and honey into the middle ear may initiate the proliferation of opportunistic pathogens leading to the blockage of the Eustachian tube (Parkash *et al.*, 2004).

In the present study *Staphylococcus aureus* was found to be most predominant organism 37.2 % followed by *Pseudomonas aeruginosa* (34.7%), *Proteus mirabilis* (4.0%) and *Escherichia coli* (4.0%). Higher prevalence of *Staphylococcus aureus* in this is also same as other author having 54% of *Staphylococcus aureus* (Park *et al.*, 2008). Similar findings have been observed in Ireland (Mubassabi., 2007), Pakistan (Arshad *et al.*, 2004), and Greece (Bardanis *al.*, 2003), which reported that *P. aeruginosa* and *S. aureus* are the most common organism isolated from the patients. The reason behind *S.aureus* to be most prevalent organism might be because it is an opportunistic pathogen and normal flora of the skin, but when it enters the

human body it causes infection to tissue and mucous membrane (Alo *et al.*, 2012).

The prevalence of *K. pneumoniae* and *E. coli* in this study was 2.5 and 4.0 % respectively. Likewise, 3.4 and 2.4 % of *K. pneumoniae* and *E. coli* was, respectively reported in studies from Ethiopia (Wasihun *et al.*, 2015). Moreover, a study in India reported 8 and 4 % of *K. pneumoniae* and *E. coli*, respectively through the proportion varies (Prakash *et al.*, 2013). Isolation of fecal bacteria like *K. pneumoniae* and *E. coli* might indicate that individuals were at risk of infection due to poor hygiene conditions.

P. aeruginosa showed sensitivity to Imipenem (S-100%), Piperacillin/tazobactam (S- 100%), Ceftazidime (S-97.1%, R- 2.9%), Cefepime (97.1%, R-2.9%), Amikacin (S-88.4%, R-11.6%), Gentamicin (S- 85.5%, I- 2.9%, R- 11.6%), Aztreonam (S-85.5%, I-10.1%, R-4.3%), Ciprofloxacin (S- 82.6%, R- 17.4%), Levofloxacin (S- 82.6%, R- 17.4%), Ofloxacin (S- 81.2%, I-2.9%, R-15.9%) & Ticarcillin/Clavulanate (S-78.3%, I-15.9%, R-5.8%). The antimicrobial pattern of Imipenem, Ciprofloxacin, Ceftazidime, and Levofloxacin is in agreement with the study conducted in by Moorthy *et al.*, 2013 Whereas the pattern of Amikacin & Gentamicin is in agreement with the study conducted by AHC Loy *et al.*, 2002.

S. aureus showed sensitivity to Vancomycin (S-100%), Teicoplanin(S-100%), Linezolid (S-100%), Chloramphenicol (S-97.3%, R-2.7%), Amikacin (S-97.3%, I-1.4%, R-1.4%), Tetracycline (S-91.9%, R-8.1%), Gentamicin (S-85.1%, I-12.2%, R-2.7%), Clindamycin (S-78.4%, R-21.6%), Cefoxitin (S-75.7%, R-24.3%), Erythromycin (S-68.9%,I-9.5%,R-21.6%), Levofloxacin (S-40.5%,I-33.8%,R-25.7%), Cotrimoxazole (S-40.5%,R-59.5%), Ciprofloxacin (S-

25.7%, I-14.9%, R-59.5%), Ofloxacin (S-25.7%, I-14.9%, R-59.5%), & penicillin (S-6.8%, R-93.2%). The antimicrobial pattern of vancomycin, Cefoxitin, Amikacin & Gentamicin is in agreement with the study conducted in by Nazir *et al.*, 2014.

When the results of various workers were compared, it became obvious that the profile of pathogens and antibiotic sensitivity

patterns causing ear infections has been changing from time to time. The probable reasons for this variation could be contributed to antimicrobial resistance profile of bacteria varies among the population because of difference in geography, local antimicrobial prescribing practices and prevalence of resistant bacterial strains.

Table.1 Distribution of ear infection in relation to Gender

Gender	No of samples received (n= 295)	Percentage	Number of positive Cases (n= 194)	Percentage
Male	160	54.2	110	68.80
Female	135	45.8	84	62.20

Table.2 Age and Sex wise distribution of positive cases

Age (Years)	Male		Female		Total		%
	Total Received	Positive (%)	Total Received	Positive (%)	Total	Positive	
<10	50	34 (68.0)	34	22 (64.7)	84	56	66.7
11-20	27	16 (59.3)	20	9 (45.0)	47	25	53.2
21-30	47	32 (68.1)	30	19 (63.3)	77	51	66.2
31-40	21	17 (81.0)	23	17 (73.9)	44	34	77.3
41-50	8	6 (75.0)	15	10 (66.7)	23	16	69.6
> 50	7	5 (71.4)	13	7 (53.8)	20	12	60.0
Total	160	110 (68.8)	135	84 (62.2)	295	194	65.7

Table.3 Number of Isolates

Number of Isolates	Number of patients	Percentage
0	101	34.2
1	189	64.1
2	05	1.7

Table.4 Microbiological profile of pathogens isolated from ear swabs (n= 199)

Type of Isolate	Type of Organism	Frequency of Isolates	% of Patients Infected
Gram positive bacteria (n= 87) 43.72%	<i>Staphylococcus aureus</i>	74	37.2
	<i>Streptococcus pyogenes</i>	6	3.0
	<i>Streptococcus pneumonia</i>	5	2.5
	<i>Enterococcus spp.</i>	2	1.0
Gram negative bacteria (n= 94) 47.24%	<i>Pseudomonas aeruginosa</i>	69	34.7
	<i>Escherichia coli</i>	8	4.0
	<i>Proteus mirabilis</i>	8	4.0
	<i>Klebsiella pneumonia</i>	5	2.5
	<i>Enterobacter aerogenes</i>	4	2.0
Fungal Isolates (n=18) 9.05%	<i>Aspergillus spp.</i>	10	5.0
	<i>Candida spp.</i>	8	4.0
Total		199	---

Table.5 Antibiotic sensitivity patterns of Gram-negative bacteria from ear swabs

Antibiotic	<i>P.aeruginosa</i> (n= 69)			<i>E.coli</i> (n=7)			<i>E. aerogenes</i> (n=4)			<i>P.mirabilis</i> (n=6)			<i>K.pneumoniae</i> (n=4)		
	S%	I%	R%	S%	I%	R%	S%	I%	R%	S%	I%	R%	S%	I%	R%
CXM (O)	-	-	-	50.0	12.5	37.5	-	-	-	50.0	0.0	50.0	40.0	0.0	60.0
CXM (P)	-	-	-	62.5	0.0	37.5	-	-	-	50.0	0.0	50.0	40.0	0.0	60.0
CFM	-	-	-	62.5	12.5	25.0	-	-	-	25.0	0.0	75.0	40.0	0.0	60.0
CAZ	97.1	0.0	2.9	75.0	0.0	25.0	100	0.0	0.0	75.0	0.0	25.0	60.0	0.0	40.0
CTR	-	-	-	75.0	0.0	25.0	100	0.0	0.0	75.0	0.0	25.0	80.0	0.0	20.0
CPM	97.1	0.0	2.9	75.0	0.0	25.0	100	0.0	0.0	75.0	0.0	25.0	80.0	0.0	20.0
OF	81.2	2.9	15.9	87.5	0.0	12.5	100	0.0	0.0	75.0	12.5	12.5	100	0.0	0.0
CIP	82.6	0.0	17.4	87.5	0.0	12.5	100	0.0	0.0	75.0	12.5	12.5	100	0.0	0.0
LE	82.6	0.0	17.4	87.5	0.0	12.5	100	0.0	0.0	87.5	0.0	12.5	100	0.0	0.0
AMC	-	-	-	75.0	0.0	25.0	-	-	-	100	0.0	0.0	80.0	0.0	20.0
PT	100	0.0	0.0	100	0.0	0.0	100	0.0	0.0	100	0.0	0.0	100	0.0	0.0
GEN	85.5	2.9	11.6	100	0.0	0.0	100	0.0	0.0	87.5	0.0	12.5	100	0.0	0.0
AK	88.4	0.0	11.6	100	0.0	0.0	100	0.0	0.0	87.5	0.0	12.5	100	0.0	0.0
COT	-	-	-	75.0	0.0	25.0	75.0	0.0	25.0	0.0	0.0	100	60.0	0.0	40.0
TE	-	-	-	75.0	0.0	25.0	75.0	25.0	0.0	-	-	-	60.0	0.0	40.0
IPM	100	0.0	0.0	100	0.0	0.0	100	0.0	0.0	100	0.0	0.0	100	0.0	0.0
AT	85.5	10.1	4.3	-	-	-	-	-	-	-	-	-	-	-	-
TCC	78.3	15.9	5.8	-	-	-	-	-	-	-	-	-	-	-	-

CXM(O)- Cefuroxime (Oral), CXM (P)- Cefuroxime (Parenteral), CFM- Cefixime, CAZ- Ceftazidime, CTR- Ceftriaxone, CPM-Cefepime, OF- Ofloxacin, CIP- Ciprofloxacin, LE- Levofloxacin, AMC- Amoxycylav-clavulanate, PT- Piperacillin-tazobactam, GEN- Gentamycin, AK- Amikacin, COT- Cotrimoxazole, TE- Tetracycline, IPM- Imipenem, AT- Aztreonam, TCC- Ticarcillin/Clavulanate.

S= Sensitive, R- Resistant, I= Intermediate

Table.6 Antibiotic sensitivity patterns of Gram-positive bacteria from ear swabs

Antibiotic	<i>S.aureus</i> (n= 74)			<i>S.pyogenes</i> (n= 6)			<i>S.pneumoniae</i> (n= 5)			<i>Enterococcus spp.</i> (n= 2)		
	S%	I%	R%	S%	I%	R%	S%	I%	R%	S%	I%	R%
P	6.8	0.0	93.2	100	0.0	0.0	100	0.0	0.0	50.0	0.0	50.0
AMP	-	-	-	100	0.0	0.0	-	-	-	50.0	0.0	50.0
CTR	-	-	-	100	0.0	0.0	-	-	-	-	-	-
CPM	-	-	-	100	0.0	0.0	-	-	-	-	-	-
CIP	25.7	14.9	59.5	-	-	-	-	-	-	50.0	0.0	50.0
OF	25.7	14.9	59.5	66.7	0.0	33.3	100	0.0	0.0	50.0	0.0	50.0
LE	40.5	33.8	25.7	66.7	0.0	33.3	100	0.0	0.0	50.0	0.0	50.0
GEN	85.1	12.2	2.7	-	-	-	-	-	-	-	-	-
AK	97.3	1.4	1.4	-	-	-	-	-	-	-	-	-
AZM	-	-	-	50.0	0.0	50.0	80.0	0.0	20.0			
E	68.9	9.5	21.6	50.0	0.0	50.0	80.0	0.0	20.0	50.0	0.0	50.0
CD	78.4	0.0	21.6	66.7	0.0	33.3	80.0	0.0	20.0	-	-	-
C	97.3	0.0	2.7	83.3	0.0	16.7	100	0.0	0.0	100	0.0	0.0
TE	91.9	0.0	8.1	83.3	16.7	0.0	80.0	0.0	20.0	100	0.0	0.0
COT	40.5	0.0	59.5	-	-	-	60.0	0.0	40.0	-	-	-
LZ	100	0.0	0.0	100	0.0	0.0	-	-	-	100	0.0	0.0
TEI	100	0.0	0.0	-	-	-	-	-	-	100	0.0	0.0
VA	100	0.0	0.0	100	0.0	0.0	100	0.0	0.0	100	0.0	0.0
CX	75.7	0.0	24.3	-	-	-	-	-	-	-	-	-
HLG	-	-	-	-	-	-	-	-	-	100	0.0	0.0

P- Penicillin, AMP- Ampicillin, CTR- Ceftriaxone, CPM- Cefepime, CIP-Ciprofloxacin, OF- Ofloxacin, LE- Levofloxacin, GEN- Gentamicin, AK- Amikacin, AZM- Azithromycin, E-Erythromycin, CD- Clindamycin, C- Chloramphenicol, TE- Tetracycline, COT- Cotrimoxazole, LZ- Linezolid, TEI- Teicoplanin, VA- Vancomycin, CX- Cefoxitin, HLG- High Level Gentamicin.

S= Sensitive, R- Resistant, I= Intermediate

In conclusion, the Bacterial ear infection is a foremost health trouble in the study area. *S.aureus*, *P. aeruginosa*, *P. mirabilis* and *E. coli* were the dominant isolates. In gram-positive bacteria, high resistance was found in *S. aureus* isolates, penicillin (93.2%) was found to be most resistant followed by ciprofloxacin, Ofloxacin, cotrimoxazole (59.5%), and cefoxitin (24.3%). Moreover, a considerable level of oxacillin-resistant *S.aureus* suggests the diffusion of Methicillin-resistant *S.aureus* in the community.

As the symptoms collapse, many patients stop taking antibiotics before the completion of therapy and let the partly resistant microbes to flourish. Patients should be instructed to evade such practice. Continuous and periodic assessment of microbiological pattern and antibiotic sensitivity of isolates is essential to decrease the potential risk of complications by early institution of appropriate treatment under expert supervision. I believe that data from this study may contribute to an effective management of ear infections.

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References

- Alo, M.N., Anyimm, C. *et al.*, 2012. Prevalence, antibiogram of bacterial pathogens associated with otitis media among primary school children in Ebonyi State. *J. Pharm. Bio. Sci.*, 1(78): 17-20.
- Arshad, M., Khan, N.U. *et al.*, 2004. Sensitivity and spectrum of bacterial isolates in infectious otitis externa. *J. Coll. Phy. Surg. Pak.*, 14: 146-149.
- Aslam, M.A., Ahmed, Z. *et al.*, 2004. Microbiology and drug sensitivity patterns of chronic suppurative otitis media. *J. Coll. Physicians Surg. Pak.*, 14: 459-461.
- Iqbal, B., J., Khan, W. *et al.*, 2009. Frequency of chronic suppurative otitis media in the junior ranks of pak army. *Pak. Armed forces Med. J.*, 59: 367-370.
- Bardanis, J., Batzakakis, D., *et al.*, 2003. Types and causes of otorrhea. *Auris Naus Larynx*, 30: 253-257.
- Cheng, A.T., Youg, N.M. 1997. Inflammatory diseases of the ear. *Indian J Pediatr* 64: 747-753.
- CLSI, Performance standards for antimicrobial susceptibility testing; Twenty-fifth International supplement., 2015. CLSI document M100-S24. Wayne, PA: Clinical Laboratory and Standard Institute.
- Grevers, G. 2010. Challenges in reducing the burden of otitis media disease: an ENT perspective on improving management and prospects for prevention. *Int. J. Pediatr. Otorhinolaryngol.*, 74: 572-577.
- Haraldsson, G., Holbrook, W.P. *et al.*, 2004. Clonal similarity of salivary and nasopharyngeal *Fusobacterium nucleatum* in infants with acute otitis media experience. *J. medical microbiol.*, 532: 165-5.
- Haynes, D.S. 2002. Preoperative antibiotics in chronic supportive otitis media. *Ear Nose Throat J.*, 81: 13-15.
- Kristo, B., Buljan M. 2011. Microbiology of the chronic suppurative otitis media. *Medicinski Glasnik*, 8: 2.
- Kumar, H., Seth., S. 2011. Bacterial and fungal study of 100 cases of chronic suppurative otitis media. *J. Clin. Diagn. Res.*, 5(6); 1224-7.
- Lodhi, M., Munir, T. *et al.*, 2010. Chronic suppurative otitis media; Empiric quinolones in children. *Professional Med. J.*, 17(3): 420-4.
- Loy, A.H.C., Tan, A.L., *et al.*, Microbiology of chronic suppurative otitis media in Singapore. *Singapore Med. J.*, 43(6): 296-299.
- Melaku, A., Lulseged, S., 1999. Chronic suppurative otitis media in childrens hospital in Addis, Ababa, Ethiopia. *Ethiop Med. J.*, 37: 237-45.
- Moorthy, P.N.S., Jadi, L. *et al.*, 2013. Clinical application of microbiological study of chronic suppurative otitis media. *Indian J. Otolaryngol. Head Neck Surg.*, 2: 290-94.
- Mubassabi, K. 2007. Bacteriology of discharging ears. *Ir. Med. J.*, 100: 379-380.
- Nazir, A., Kadri, S.M. 2014. Aerobic bacteriology of chronic suppurative

- otitis media: a hospital based study. *Int. J. Res. Med. Sci.*, 2(4): 521-524.
- Nwabuisi, C., Ologe F.E. 2002. Pathogenic agents of chronic suppurative otitis media in Ilorin, Nigeria. *East Afr. Med. J.*, 79(4): 202-5.
- Oguntibeju, O. 2003. Bacterial isolates from patients with ear infection. *Indian J. Med. Microbiol.*, 21(4): 214-294.
- Park, D.C., Lee, S.K. *et al.*, 2008. Antimicrobial resistance of Staphylococcus from otorrhea in chronic suppurative otitis media and comparison with results of all isolated Staphylococci. *Eur. J. Clin. Microbiol. Infec. Dis.*, 27(7); 571-7.
- Poorey, V.K., Iyer, A. 2002. Study of bacterial flora in CSOM and its clinical significance. *Indian J. Otolaryngol. Head Neck. Surg.*, 59: 91-95.
- Prakash, M., Lakshmi, K. *et al.*, 2013. Bacteriological profile and their antibiotic susceptibility pattern of cases of chronic suppurative otitis media. *Asian J. Pharm. Clin. rRes.*, 6(3): 210-212.
- Prakash, R., Juyal D. *et al.*, 2013. Microbiology of chronic suppurative otitis media in a tertiary care setup of Uttarkhand State. *India N. Am. J. Med. Sci.*, 5(4) :282-7.
- Raakhee, T., and Unguturu, S.R. 2014. Bacteriological study of discharging ear in patients attending a tertiary care hospital. *Int. J. Res. Med. Sci.*, 2(2): 602-606.
- Richard, E.B., Roberts, M.K.1996. Otitis Media and its complications in Nelson's Textbook of pediatrics. 1814-1824.
- Srivastava, A., Singh, R *et al.*, 2011. Microbiological evaluation of an active tubotympanic type of chronic suppurative otitis media. *Nepalese J. Ent, Head and Neck Surg.*, 12: 14-16.
- Tobih, J.E., Taiwo, S.S. *et al.*, 2006. Clinical and Microbiological profile of EAR INFECTIONJS IN Osogbo, Nigeria. *Trop .Doct.*, 36: 165-166.
- Wasihun, A.G., Zemene, Y. 2015. Bacterial profile and antimicrobial susceptibility patterns of otitis media in Ayder teaching and referral Hospital, Mekelle University, Northern Ethiopia. *Springerplus*, 4: 701.
- World Health Organization. 1986. Prevention of deafness and hearing impairment. Geneva: report by the Director General. Document a29/14.
- Yousuf, M., Majumder, K.A. 2011. Clinical study on chronic suppurative otitis media with cholesteatoma, Bangladesh. *J. Otorhinolaryngol.*, 17(1): 42-7.

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