

Original Research Article

<http://dx.doi.org/10.20546/ijcmas.2016.510.077>

Isolation of Tomato Fruits Mycoflora and Evaluation *in vitro* and *in vivo* by *Trichoderma harzianum*

Mokhtar Hamitou^{1,2*}, Laid Dehimat² and M. Mourad Senoussi¹

¹Laboratory of Bimolecular and Plant Amelioration, Larbi-Ben-M'hidi University, Oum El Bouaghi, BP 358, Constantine Road, 04000, Algeria

²Laboratory of Mycology, Biotechnology and Microbial activity, University of Mentouri, Constantine, 25000. Algeria

*Corresponding author

ABSTRACT

The present investigation aimed is to isolate and identify the mycetes accompanying the tomato fruits (*Lycopersicon esculentum*), and to evaluate *in vitro* and *in vivo* the ability of *T.harzianum* to control the isolated mycetes. Some infected tomato fruits by mycetes were brought from Oum-elbouaghi market. The results of isolation allowed the identification of *Stemphylium sp.* and *Aspergillus niger*. One isolate of *T.harzianum* / *Hypocrea lixii* was utilized in this study. The results of direct confrontation (*in vitro*) of *T.harzianum* against *Stemphylium sp.* and *A.niger* on PDA medium indicated the inhibition of mycelium growth in variable degrees; it was equal in the fourth day of the experiment to 54.54 % and 52.17% for *Stemphylium sp.* and *A.niger* respectively. However, it did not show any growth of the tested fungus when re-planting a disk from the interaction hyphal area between *T.harzianum* and *Stemphylium sp.* or *A.niger* from dual cultures, while *T.harzianum* grew alone in the plates. The microscopic observations of mycelia of dual culture in slide methods showed that the mycelia of *T.harzianum* induced degradation and aggregated the spores and analyzed the mycelia of *A.niger*, overgrowing the mycelia of *Stemphylium sp* and coiled around of them and degrading them. *In vivo* screening showed after 10 days of incubation an antagonistic activity of *T.harzianum* against the tested fungus on tomato fruits, with inhibition equal to 100 % and 95% in *Stemphylium sp.* and in *A.niger* respectively, compared with controls. Beside we found that the treated fruits with *T.harzianum* stayed saints as compared with control, when *Stemphylium* and *A.niger* soft rot infected all surface the test fruits. This strain of *T.harzianum* may offer potential for biological control of tomato *Stemphylium* and *A.niger* soft rot.

Keywords

Stemphylium sp.,
soft rot,
Aspergillus niger,
Lycopersicon
esculentum,
confrontation,
slide methods,
inhibition

Article Info

Accepted:
20 September 2016
Available Online:
10 October 2016

Introduction

Tomato plant (*L.esculentum* L.) originated South America belongs to Solanaceae family is a widely grown vegetable in the world. It is the most popular vegetable

world-wide. The leading producer of tomato in the world is USA followed by China, Italy, Turkey, Egypt, Spain, Romania, Brazil and Greece (Wani, 2011). In Algeria, the

tomato crop is grown over an area of 292 000 hectares, it accounts for 51% of the total vegetable production (Nechadi *et al.*, 2002). Tomato crop suffered every year from a number of pathogenic diseases (Wani, 2011). The fungus *Stemphylium solani* causes leaf blight of tomato in Brazil (Mehta, 1998). Most the *Stemphylium* species on record as plant pathogens in Japan (Daisuke *et al.*, 2015). *A.niger* caused a disease called black mold on certain fruits and vegetables such as grapes, tomato, onions and peanuts (Sharma, 2012). Mallek *et al.*, (1995) reported that the *A.niger* was a one of the most common pathogens and caused loss of 25% in tomato fruit in Egypt. *A.niger* was responsible for the post harvest rot of Tomato fruits in Nsukka (Nigeria) and the pathogenesis tests confirmed that, the fungal isolate is one of the causal agents of the rot (Jude and Nneka, 2012). Chemical compounds have been used to control plant diseases, this has no doubt increased crop production but with the attendant deterioration of the environment and human health. In addition to killing target pathogens, pesticides may also kill various beneficial organisms and their toxic effects can persist in the soil but abuse in their employment has favored the development of pathogens resistant (Kamala and Indira, 2012 ; Nneka and Uken, 2013). The Algerian farmer used the chemical fungicides to control the fungal diseases of tomato plants, but in more time the treatment traces are observed in tomato fruits, because they usually aren't rinsed fruits before marketing (fig.1.1). Biological control using potential microorganisms having strong antifungal activity is coming up as an alternative strategy for disease management, which is also ecology-conscious and environment friendly (Kamala and Indira, 2012). Several biocontrol strategies have been proposed for controlling the plant pathogens, but practical

applications are still limited (Hibar *et al.*, 2007). *Trichoderma* species are common soil-inhabiting fungi that have been developed as effective biocontrol agents against various phytopathogenic microorganisms (Bel Haj Khethr *et al.*, 2008).

The aim of the present investigation was to isolate and to identify the mycetes accompanying the tomato fruits (*Lycopersicon esculentum*), and to evaluate the *in vitro* and *in vivo* ability of *T.harzianum* to control that isolated mycetes.

Materials and Methods

Fungal strains

Stemphylium sp. and *Aspergillus niger* were isolated from infected tomato fruits, which were brought from Oum-elbouaghi market, and identified based on the microscopic observations of their reproductive and colony characteristics in laboratory of microbiology, university of Oum-elbouaghi (Algeria) (Botton *et al.*, 1990; Rémi, 1997; Robert *et al.*, 1981). A local strain of *T.harzianum* / *Hypocrea lixii* , was identified in the same laboratory and verified in Walloon Center of Biology Industrial, University of Liege, Belgium.

***In vitro* Evaluation of the antagonistic capability of *T.harzianum* against *Stemphylium sp.* and *A. niger*, on PDA medium (direct confrontation)**

To study the direct confrontation between *T.harzianum* and *Stemphylium sp.* or *A.niger* Two plugs of mycelium (8mm diameter) were cut from the margins of actively cultures growing on PDA medium, one carrying the stock of *T.harzianum* and the other of *Stemphylium sp.* or *A.niger*. Then they placed at the periphery of Petri plates

(9cm in diameter) at the same distance on PDA medium (dual cultures). One plug of *Stemphylium sp.* or *A.niger* were maintained as controls (alone cultures). Each replicate has three plates. Both the dual and alone cultures were incubated at 25°C for four days, and measurement of colony diameters (in millimeters) was taken every 24 hours. The percentage of inhibition growth (I) was calculated by using the formula given below: $[I (\%) = (1 - T / C) \times 100]$. Where: I=Percentage inhibition of pathogen growth by antagonists. C=Radial growth in control. T=Radial growth in the treatment (Berber *et al.*, 2009; Hamitou and Dehimat, 2015).

Evaluation of dual culture using slide method

For each pathogen (*Stemphylium* or *A.niger*)-*T.harzianum* interaction, a clean slide was placed in 9 cm diameter plates and sterilized. Following that, a small amount of PDA medium was spread over the slide to make a thin PDA film on the slide. The 5 mm discs of one week old of each pathogen and *T. harzianum* isolates were placed on the opposite sides of the slide 3 cm apart on the PDA surface.

Then 5ml of distilled water was added to the plate to prevent drying and then incubated at 25°C for a week. At the end of incubation period, region of contact between *T. harzianum*-Pathogen hyphae was stained with lacto phenol and cotton blue and examined under a light microscope (Al-Saeedi and Moqdad, 2014).

Preparation of tomato fruits

Intact red tomatoes (*L.esculentum* Mill.), uniform in size and color, were obtained from the market of Oum-Elbouaghi city. The fruits were surface-sterilized by soaking in 2% aqueous sodium hypochlorite for 5

min, they were thoroughly rinsed with sterile distilled water, dried using sterile filter papers, and then wounded by removing a rectangular area at the equator of each fruit, (3cmx4cm) in diam. and 3 mm in depth, from the surface, using a sterile scalpel (Berrada *et al.*, 2012).

In vivo. Evaluation of the antagonistic capability of *T.harzianum* against *Stemphylium sp.* and *A.niger* on tomato fruits

Fresh cultures of *Stemphylium sp.*, *A.niger* and *T.harzianum* were used for each experiment to evaluate the antagonistic activity. Two plugs of mycelium (8mm diameter) were cut from the margins of actively cultures growing on PDA medium, one carrying the stock of *T.harzianum* and the other of *Stemphylium sp.* or *A.niger* were then placed one beside of the other at the center of the rectangular area of tomato fruits.

As control, fruits were either inoculated with *Stemphylium sp.* or *A.niger* alone. The fruits were then stored at 20°C ± 2. for 10 days in autoclaved glass jars with hermetic covers. The percentage of disease reduction of *Stemphylium* or *A.niger* rot on tomato fruits, was calculated using the following formula:

$$(\%) = (A-B)/A \times 100$$

where A is the lesion diameter recorded in tomato fruit inoculated with the *Stemphylium sp.* or *A.niger* alone

B is the lesion diameter recorded in infected tomato fruits treated with *T.harzianum*. All *in vivo* antagonism assays were made in triplicate (Berrada *et al.*, 2012; Hamitou and Dehimat, 2015).

Results and Discussion

***In vitro* Evaluation of the antagonistic capability of *T.harzianum* against *Stemphylium sp.* and *A.niger* on PDA medium (direct confrontation)**

The results of the direct confrontation between *T.harzianum* against *Stemphylium sp.* and *A.niger* on PDA medium, showed that when the mycelium of the both cultures came due to the contact together, the hyphal growth of *Stemphylium sp.* and *A.niger* were found to be inhibited by hyphae of *T.harzianum* fig.(2.1 and 2.4). That inhibition in the third day of the experiment was: 41.18 % and 29.41% and in the fourth day the amounts were: 54.54 % and 52.17% for *Stemphylium sp.* and *A.niger* respectively (table1) and (fig.3). Besides, showed no growth of mycelia of *Stemphylium sp.* or *A.niger* when re-planting the disks from the interaction hyphal area between *T.harzianum* and *Stemphylium sp.* or *A.niger* from dual cultures, while *T.harzianum* grew alone in the plates Fig (2.3 and 2.6).

Evaluation of dual culture using slide methods

The microscopic observations of mycelia of

dual culture in slide methods showed that the mycelia of *T.harzianum* overgrowing the mycelia of *Stemphylium sp.* and coiled around of them and degrading them fig.(4.2, 4.3); induced degradation and aggregated the spores and analyzed the mycelia of *A.niger* fig (4.5,4.6), compared with controls fig(4.1 and 4.4).

***In vivo* Evaluation of the antagonistic capability of *T.harzianum* against *Stemphylium sp.* and *A.niger* on tomato fruits**

After 10 days of incubation the *T.harzianum* showed an inhibition activity with a different ratios against *Stemphylium sp.* and *A.niger* on tomato fruits. The latter was equal to: 75 % and 91.66% in the seventh day and the amount in the tenth day reached 100 % and 95% for *Stemphylium sp.* and *A.niger* respectively (fig.5).

Beside we found that the treated fruits with *T.harzianum* stayed saints (fig. 6.d) and (fig.7.d) compared with controls when *Stemphylium* and *A.niger* soft rot infected all surface the fruits (fig.6.c) and (fig.7.c).

Table.1 *In vitro*. Effect of *T.harzianum* on the mycelia growth of *Stemphylium sp.* and *A.niger*, on PDA medium.

| Fungus species | Radial growth rate (mm) after: | | | |
|---|--------------------------------|---------|---------|---------|
| | 24 hour | 48 hour | 72 hour | 96 hour |
| Dual culture <i>T.harzianum</i> | 30 | 78 | 130 | 130 |
| Dual culture <i>Stemphylium sp.</i> | 6 | 16 | 20 | 20 |
| Alone culture <i>Stemphylium sp.</i> | 10 | 20 | 34 | 44 |
| % inhibition of mycelia growth <i>Stemphylium sp.</i> | 40 | 20 | 41.18 | 54.54 |
| Dual culture <i>T.harzianum</i> | 34 | 80 | 130 | 130 |
| Dual culture <i>A.niger</i> | 10 | 20 | 24 | 22 |
| Alone culture <i>A.niger</i> | 12 | 22 | 34 | 46 |
| % inhibition of mycelia growth <i>A.niger</i> | 16.16 | 9.1 | 29.41 | 52.17 |



Fig.1 Treated tomato fruits with fungicide, 1. Infected tomato fruits with *A.niger* rot,2., Infected tomato fruits with *Stemphylium* rot,3., Some tomato fruits utilized in the *in vivo* test, 4.

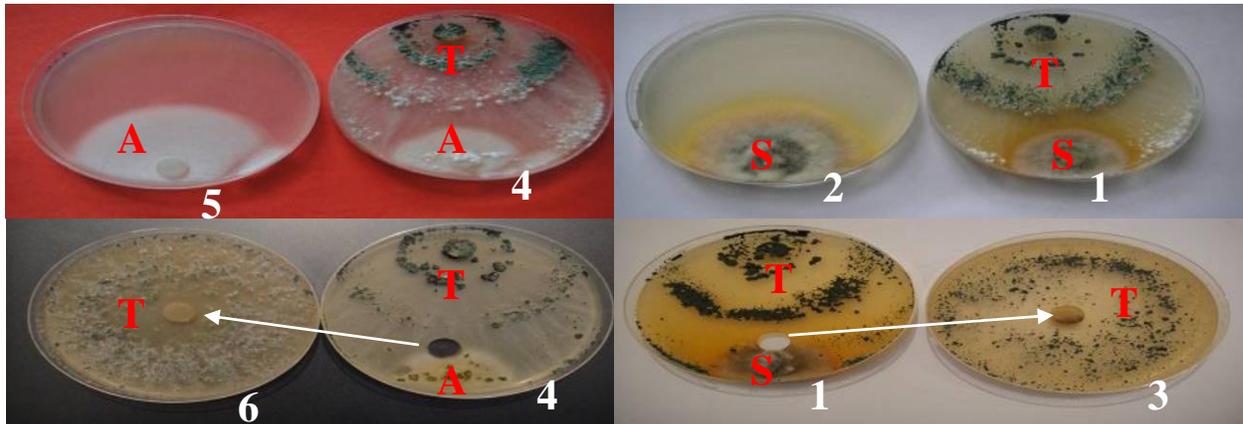


Fig.2 *In vitro* effect of *T.harzianum* against *Stemphylium* sp. and *A.niger*. dual cultures(1)and (4)., controls (2) and (5)., re-planting plates (3) and (6). S= *Stemphylium* , A= *Aspergillus*, T=*Trichoderma*.

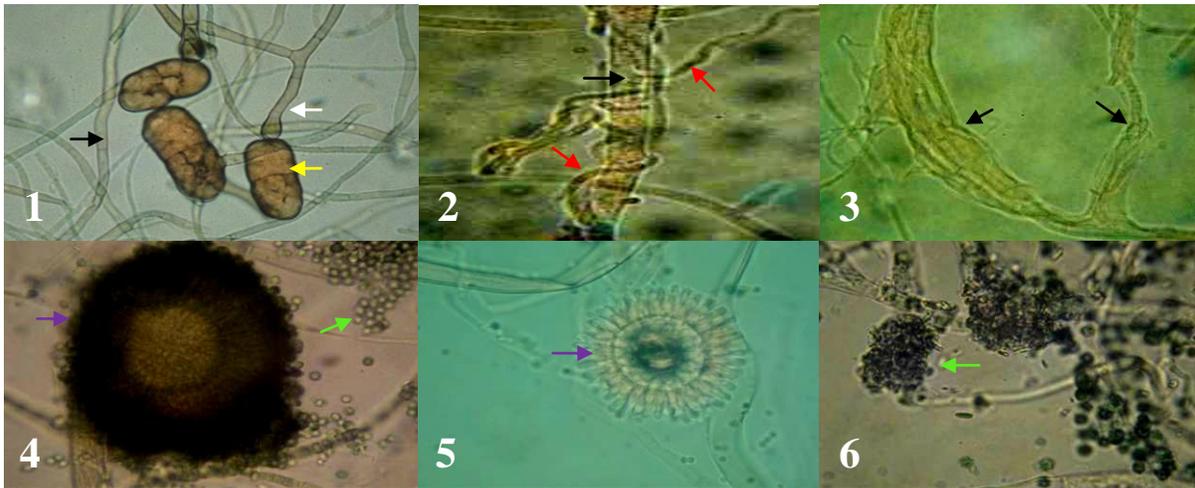
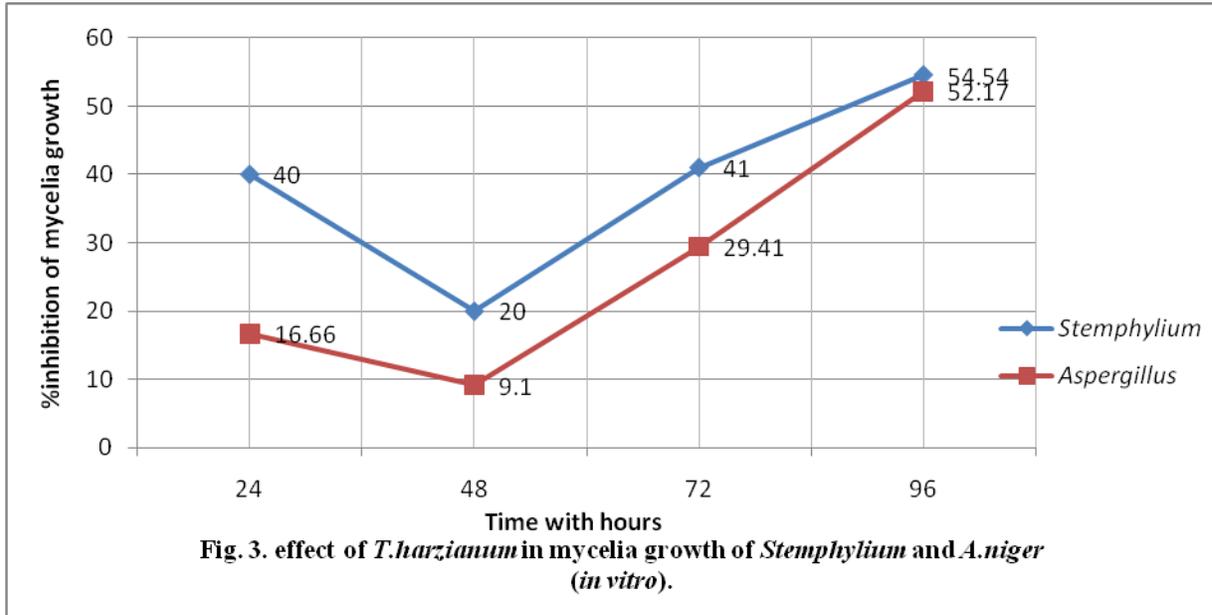


Fig.4 Microscopic observations of the *in vitro* effect of *T.harzianum* against *Stemphylium sp.* and *A.niger*. Decomposition phenomenon(3), (5) and(6); Mycoparasitism phenomenon (*Trichoderma* hyphal coiling around of *Stemphylium* hyphal),2. Controls,1 and 4. *Stemphylium*(hyphe=black arrow, sporophore = white arrow, dictyospore= yellow arrow)., *A.niger* (conidia= green arrow, vesicle=purple arrow., *Trichoderma* hyphe = red arrow.

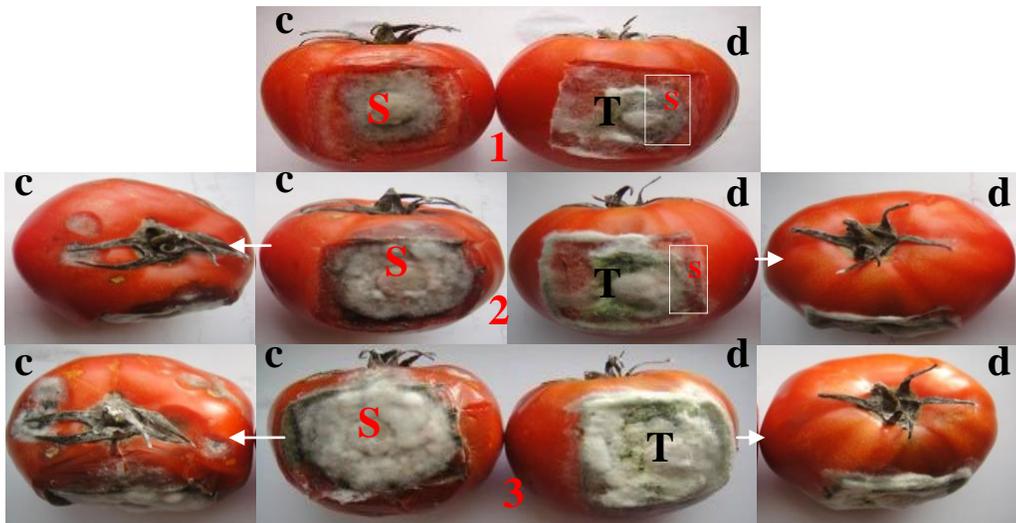
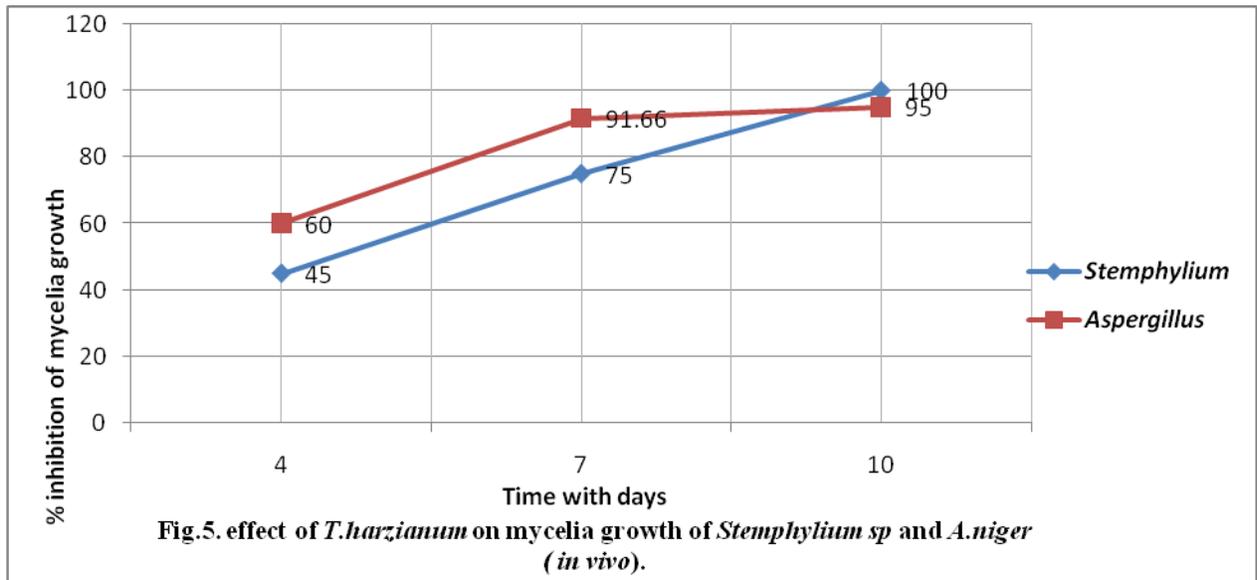


Fig. 6 In vivo effect of *T.harzianum* against *Stemphylium sp*. Test(after 4 days, 1. ; after 7 days,2. ; after ten days,3.) ; c= control., d= dual culture ; S=*Stemphylium*, T=*Trichoderma*.

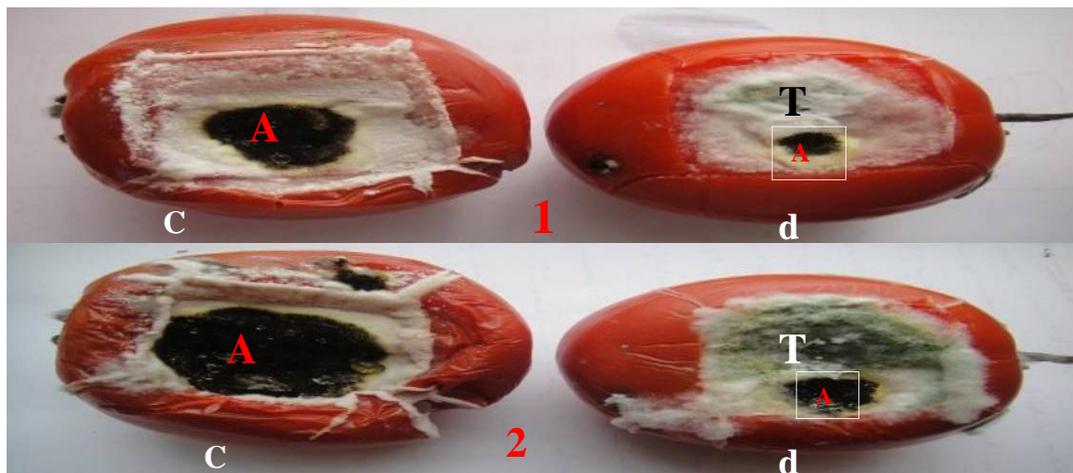


Fig.7 *In vivo* effect of *T.harzianum* against *A.niger*. test(after 4 days,1.; after 7 days,2.); c= control., d= dual culture. A= *A.niger*, T=*Trichoderma*.

In this investigation, this local strain of *T. harzianum* showed a high efficiency both *in vitro* and *in vivo* against *Stemphylium sp.* and *A.niger*. This results was confirmed by many paper studies, where found that the *T.harzianum* could restrict growth of *A.niger in vitro* (dual culture) with 75% and amounted to reach 78.77% (Agrwal *et al.*, 2011; Lone *et al.*, 2012). The *T.harzianum* can inhibited the growth of *Bipolaris sp.*, *F.oxysporium*, *Fusarium sp.* and *R.solani* with a different ratios, and inhibited there spore's formation, with recording a different degrees of parasitism (Azza and Allam, 2004; Berber *et al.*, 2009; Comporota, 1985; Hibar *et al.*, 2005). *T.harzianum* strains produced an inhibitor metabolites as 1, 3- b- glucanase and chitinase which were inhibited the growth of *G.graminis* var. *tritici*, *F.culmorum* and *F.moniliforme* on PDA medium (Cigdem and Merih, 2004). *T.harzianum* can produced nonanoic acid into a liquid culture medium. The latter has a strongly affected both mycelial growth and spore germination of the cacao pathogen (*Crinipellis perniciosa* and *Moniliophthora*

roreri) (Anejaa *et al.*, 2005). *T.harzianum* reduced disease incidence significantly against *P.ultimum* and *R.solani* on both cucumber and tomato on greenhouse (Johanne *et al.*, 2002). In the similar study, Yacoub, (1999), found that the *Trichoderma sp* reduced the lesion development and number of conidiophores of *Botrytis sp.* in foliar discs of strawberry test, compared with the non-treated (control). This local strain of *T.harzianum* may offer potential for biological control of tomato *Stemphylium* and *A.niger* soft rot.

References

- Agarwal, T., A. Malhotra., M. Biyani., and Trivedi, P.C. 2011. *In vitro* interaction of *Trichoderma* isolates against *Aspergillus niger*, *Chaetomium sp.* and *Penicillium sp.* *Indian J. Fundamental and Appl. Life Sci.*, 1: 125- 128.
- AL-Saeedi, S.S., and Moqdad AL-Ani, B. 2014. Study of antagonistic capability of *Trichoderma harzianum* isolates against some pathogenic soil borne

- fungi. *Agric. Biol. J.N. Am.*, 5: 15-23.
- Anejaa, M., J.T. Gianfagnaa., and Prakash K.H. 2005. *Trichoderma harzianum* produces nonanoic acid, an inhibitor of spore germination and mycelial growth of two cacao pathogens. *Physiol. Mol. Plant Pathol.*, 67: 304-307.
- Azza, A.T., and Allam, D.A. 2004. Improving cumin production under soil infestation with *Fusarium* Pathogen 1- screening of biocontrol agents. *Ass. Univ. Bull. Environ. Res.*, 2: 35- 45.
- Bel Haj Khethr, F., S. Ammar., D. Saïdana., M. Daami., J. Chriaa., K. Liouane., M. A. Mahjoub., A. N. Helal., and Mighri, Z. 2008. Chemical composition, antibacterial and antifungal activities of *Trichoderma* sp. growing in Tunisia. *Annals of Microbiol.*, 58: 303- 308.
- Berber, F., A.O. Touhami., A. Badoc., et Douira, A. 2009. Antagonisme *in vitro* et *in vivo* de deux *Trichoderma* à l'égard de quatre espèces de Bipolaris pathogens sur le sorgho. *Bull, Soc. Pharm.*, Bordeaux. 148:93-114.
- Berrada, I., Benkhemmar, O., Swings, J., Bendaou, N. and Amar, M. 2012. Selection of halophilic bacteria for biological control of tomato gray mould caused by *Botrytis cinerea*. *Phytopathologia Mediterranea*, 51: 625–630.
- Botton, B., A. Breton., M. Fevre., S. Gauthir., J. P. Larpent., P. H. Gay., P. Reymond., J. J. Sanglier., Y. Vayssier and Veau, P. 1990. Moisissures utiles et nuisible importance industrielle. . Masson., Paris, Milan, Barcelone, Mexico.
- Camporota, P. 1985. Antagonisme *in vitro* de *Trichoderma* spp. vis-à-vis de *Rhizoctonia solani* Kuhn. *Agronomie*. 5:111- 115.
- Cigdem, K., and Merih, K 2004. *In vitro* antifungal activity of strains of *Trichoderma harzianum*. *Türk. J. Biol.*, 28: 111-115.
- Daisuke, K., M. Tomoo., I. Kazunori., K.Gan., H. Hong, Long., F. Naruto., T. Kenichi., and Seiya, T. 2015. Taxonomic re-examination of several Japanese *Stemphylium* strains based on morphological and molecular phylogenetic analyses. *J. Gen. Plant Pathol.*, 81: 358- 367.
- Hamitou, M., and Dehimat, L. 2015. *In vitro* and *in vivo* efficiency of *Trichoderma harzianum* against *Phoma* and *Glocladium* soft rot occurred on tomato fruits (*Lycopersicon esculentum*). *Int. J. Curr. Microbiol. App. Sci.*, 4: 141-147.
- Hibar, K., D.R. Mejda., K. Haifa., and Mohamed, E. 2005. Effet inhibiteur *in vitro* et *in vivo* du *Trichoderma harzianum* sur *Fusarium oxysporium* f. sp. *Radicis lycopersici*. *Biotechnol. Agron. Soc. Environ.*, 9: 163- 171.
- Hibar, K., M. Daami-Remadi., and El Mahjoub, M. 2007. Induction of resistance in tomato plants against *Fusarium oxysporum* f. sp. *radicis-lycopersici* by *Trichoderma* spp. *Tunisian J. Plant Protection*, 2: 47-58.
- Johanne, C., L. Lucie., O. Pierre. et Richard, R. B. 2002. Utilisation d'une souche indigène de *Trichoderma harzianum* contre cinq agents pathogènes chez le concombre et la tomate de serre au Québec. *Phytoprotection*, 83:73- 87.
- Jude, A.U., and Nneka, V.C. 2012. Preliminary Investigations of the Cause of Post-harvest Fungal Rot of Tomato. *J. Pharm. Biol. Sci.*, 4: 36-39.
- Kamala, T., and Indira D.S. 2012. Biocontrol properties of indigenous *Trichoderma* isolates from North-east

- India against *Fusarium oxysporum* and *Rhizoctonia solani*. *African J. Biotechnol.*, 11: 8491- 8499.
- Lone, A.M., M.R. Wani., S.A. Sheikh., S. Sahay., and M. Suliman Dar, M.S. 2012. Antagonistic Potentiality of *Trichoderma harzianum* Against *Cladosporium sphaerospermum*, *Aspergillus niger* and *Fusarium oxysporum*. *J. Biol. Agri. Healthcare*, 2: 72- 76.
- Mallek, A.Y., S.K. Hemida., and Bagy, M.K. 1995. Studies associated with tomato fruit and effectiveness of some commercial fungicides against three pathogen. *Mycopathologica*, 130: 109-116.
- Nechadi, S., F. Benddine., A. Moumen., and M. Kheddami. 2002. Etat des maladies virales de la tomate et stratégie de lutte en Algérie. *EPPO Bull.*, 32: 21-24.
- Nneka, C.V., and Ukeh, J.A. 2013. Efficacy of *Aframomum melegueta* and *Zingiber officinale* extracts on fungal pathogens of tomato fruit. *J. Pharm. Biol. Sci.*, 4: 13- 16.
- Rémi, C. 1997. Identifier les champignons transmis par les semences. INRA, France.
- Robert, A.S., S.H. Ellen and Connie, A.N.V. 1981. Introduction to –Food-borne Fungi C.B.S. Institute of the Royal Netherlands, Academy Arts and Science.
- Sharma, R. 2012. Pathogenicity of *Aspergillus niger* in plants. *Cibtech J. Microbiol.*, 1: 47- 51.
- Wani, A.H. 2011. An overview of the fungal rot of tomato. *Mycopath.*, 9: 33- 38.
- Yacoub, B. 1999. Biological effect of two strains of microorganisms antagonistic to *Botrytis Cinerea* causal organism of gray mold on strawberry. *An- najah Univ. J. Res.*, 13: 67- 83.

How to cite this article:

Mokhtar Hamitou, Laid Dehimat and M. Mourad Senoussi. 2016. Isolation of Tomato Fruits Mycoflora and Evaluation *in vitro* and *in vivo* by *Trichoderma harzianum*. *Int.J.Curr.Microbiol.App.Sci*. 5(10): 708-717. doi: <http://dx.doi.org/10.20546/ijcmas.2016.510.077>