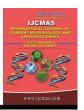


# International Journal of Current Microbiology and Applied Sciences ISSN: 2319-7706 Volume 14 Number 1 (2025)

Journal homepage: <a href="http://www.ijcmas.com">http://www.ijcmas.com</a>



## **Review Article**

https://doi.org/10.20546/ijcmas.2025.1401.007

## Microbiome Modulation-Based Therapeutic Interventions to Target Enhanced Human Health

Neena Kumar Dhiman \*\*

Department of Zoology, Gargi College, University of Delhi, Delhi-110 049, India

\*Corresponding author

#### ABSTRACT

Keywords

Probiotics, prebiotics, synbiotics, psychobiotics, intervention study, dysbiosis, immunomodulation, virome

#### **Article Info**

Received: 18 November 2024 Accepted: 25 December 2024 Available Online: 10 January 2025 The potential of gut microbiome in health and diseases has emerged as an area of profound scientific research and clinical exploration that elucidates the correlation of human immune system homeostasis, and metabolic functions with altered diversity of gut microbiota. This further validates the underlying causes and consequences of variability in gut microbiome profiles in the development of autoimmune and inflammatory diseases such as rheumatoid arthritis, cardiovascular and respiratory illnesses, neurological disorders such as neurodevelopment disorders, autism spectrum disorders, attention deficit hyperactivity disorder, stroke, Parkinson's disease, schizophrenia, Alzheimer's disease, depression, gastrointestinal inflammations including irritable bowel syndrome, inflammatory bowel disease, ulcerative colitis, Crohn's disease, Clostridium difficile infection, metabolic diseases including type 1 and type 2 diabetes mellitus, non-alcoholic fatty liver disease, liver cirrhosis, immunomodulation and certain types of cancers. Dietary interventions, fasting regimens, nutritional supplements, antibiotics, probiotics, prebiotics, synbiotics, postbiotics psychobiotics, bacteriophage, and fecal microbiota transplantation are the possible interventions that open avenues in the near future to exploit individual microbiota profiles in clinical practice as a biomarker for gut health of the patients who are at the risk of developing certain abnormalities and ailments attributed to dysbiosis. The knowledge of the present review on microbiome modulation-based therapeutic interventions to target enhanced human health comprehensively sheds light on how this upcoming field unveils the opportunities for improving human health and for preventing, treating and managing certain autoimmune diseases.

#### Introduction

The human body is home to trillions of microbes especially the intestinal compartment, where these are prevalent in maximum density to form the gut microbiota. Gut microbiota encompasses the cells of all

the microbes whereas microbiome includes their genetic materials too. Gut microbiota is a dynamic and everchanging organ that is the most forgotten part of the body including trillions of bacteria, viruses, parasites, archaea, and fungi that make humans a superorganism. It varies with sex, age, race, and lifestyle habits such as smoking, exercise, alcohol consumption, dietary habits, medications, geographical location, and temperature of the person. The microbial community starts to colonize at birth and is altered further during the life span of an individual based on the mode of delivery of the child, gestational age, breastfeeding, and infections. Exposure to xenobiotics and certain environmental factors such as level of oxygen, redox state, pH, temperature, and dietary nutrients in the gut additionally contribute to variations in gut microbiota (Spor et al., 2011; Milani et al., 2017). Besides these, sex hormones (Fransen et al., 2017), pharmaceuticals like proton pump inhibitors (PPIs) (Imhann et al., 2016) environmental intoxicants, treatment with antifungal and antibacterial agents (Wheeler et al., 2016) and consuming several prescribed drugs (Ticinesi et al., 2017) contribute to dysbiosis. The evolution of microbiota is through Proteobacteria, Actinobacteria and then to a stage where Bacteroides, Bifidobacterium, and Enterobacteriaceae are dominant to a group of Lachnospiraceae Ruminococcaceae families of phyla Firmicutes, Bacteroidetes. Actinobacteria proteobacteria. and Verrucomicrobis with family Akkermansia and eventually to a more personalized microbiota as found in adults (Heiman and Greenway, 2016; Wu et al., 2011).

pregnancy modulate These factors during the microbiome of offspring by regulating neurotransmitter pathways, signal transduction, and synaptic transmission (Lindsay et al., 2019). The study of microbiomes is important, concerning associated and correlated diseases, what are the causes, effects, and consequences of manipulations of gut microbiomes and how these interventions can be exploited for the possible development of novel, diagnostic, prognostic, and therapeutic strategies. Even the stage of the disease, risk assessment, and early diagnosis of the disease can be elucidated with the knowledge of these modulations (Fig.1).

The gut microbiome not only contributes to the well-being and diseases of the gut but also regulates the health of extra enteric organs such as the liver, pancreas, heart, brain, skin, bones, muscles, etc. It leads to the construction of the gut epithelium, its maintenance, and metabolism affecting energy balance, digestion of food, and development of the immune system and protecting against physiologic stress (Visconti *et al.*, 2019; Adak and Khan, 2019). The gut microbiome plays a role in angiotensin II-induced vascular dysfunction and hypertension (Karbach *et al.*, 2016).

Gut microbiota plays a key role in the nutritional and metabolic functions of the host and determines immune homeostasis. It modulates the level of various neurotransmitters and neuromodulators, which induce intestinal epithelial cells to release molecules such as cytokines and hormones for modulating signaling pathways within the enteric nervous system thereby controlling cognitive abilities, brain function, and behavior of the person. The brain in turn can alter the composition and functioning of microbiota via the release of hormones and neurotransmitters to influence gut physiology and environment, where certain types of microbial population can thrive. This is mediated through the gut-brain axis. Some strains of these microbial populations mediate their impact on the brain via the vagus nerve. For example, gut microbiota controls the metabolism of precursors of the kynurenine pathway which along with serotonin can be derived from tryptophan. The metabolites of the pathway can modulate neurotransmission serving themselves as neuroactive molecules (Cryan and Dinan, 2012). Signals from the gut microbiome to the brain are mediated by several bacterial neuroactive metabolites like short-chain fatty acids (SCFAs), acetate, butyrate, and propionate (MacFabe et al., 2011). Non-digestible carbohydrates serve as a source of carbon and energy for SCFA production to influence body homeostasis (Cani and Jordan, 2018; Chambers et al., 2018).

Dysbiosis is also correlated with irritable bowel syndrome (IBS), inflammatory bowel disease (IBD), Crohn's disease, ulcerative colitis, colon cancer, nonalcoholic fatty liver disease (NAFLD), liver cirrhosis, cardiovascular disease (CVD), neurological disorders, neurodegeneration, metabolic disorders such as diabetes type1 and type 2 (T1DM, T2DM), obesity and more chances of hepatitis B virus infections complications related to liver cirrhosis, hepatic encephalopathy, bacterial peritonitis, and renal failure (Cohen, 2016; Ling et al., 2016; Lakshmi et al., 2010). Gutdysbiosis is also associated with chronic liver failure, hepatocarcinoma, fibrosis and mortalities due to specific inflammatory cytokines from specific bacterial families which lead to reduced pancreaticobiliary secretions, reduced intestinal motility, impaired intestinal barrier, increased intestinal permeability, decreased gastric acidity and activation of Toll-like receptors (TLRs) (Dapito et al., 2012). Gut dysbiosis can lead to endotoxemia that may induce immune dysfunctions leading to cell necrosis and hepatic failure (Boursier et al., 2016).

Besides playing a central role in immunomodulation, gut microbiota also regulates oxidative stress which brings about homeostasis of inflammations, and macrophage cell population in the central nervous system (Angoorani et al., 2021; Li et al., 2017; Baek et al., 2013; Sampson Even increased abundance al..2016). Enterobacteriaceae, Fusibacteriaceae. Pasteurellacea. Veillonellaceae and decreased levels and Bacteroidales, Clostridiales, and Erysipelotrichales during the analysis of IBD patient samples show a correlation with the status of the disease which paves the way for easy diagnosis and offers an opportunity for early treatment (Gevers et al., 2014).

Microbiome-based therapies are still in the early stages of development due to the challenge of an undefined healthy microbiome, standardized microbiome sampling and analysis, safety and efficiency, undefined pharmacodynamics and pharmacokinetics, variability from individual to individual, less understood long-term side effects and less explored and less understood microbiome—host-environment cross-talk for health and disease management (Fig 2).

The present review aims to elucidate the data correlated with possible interventions at one place to provide deep insight into strategies attributing to the modulation of gut microbiota composition for the health of the host and to understand the existing gaps in knowledge and to explore the possibilities of future therapeutics based upon personalized plans.

## Mechanism of Action of Gut Microbiota

Bifidobacterium has been known as a key modulator of G1T helper 17 (Th17) cells besides maintaining barrier function to check the entry of pathogenic microbes. The role of gut microbiota in tumorigenesis is in addition to genetic factors. The possible mechanism includes the promotion of inflammations due to more expression of pro-inflammatory genes such as COX2, IL-6, IL-8, TNF-α, and MMP3. However, *Helicobacter pylori* are known to play a protective role in the development of esophageal adenocarcinoma (Hamada *et al.*, 2000).

Enterotoxigenic *Bacteroides fragilis* leads to activation of Wnt/β-catenin signaling pathway and NF-κB producing inflammatory mediators via the release of a toxin known as fragilysin which causes cells to proliferate excessively (Sokol, 1999; Shiryaev *et al.*, 2013). *Enterococcus faecalis* and *E.coli* induce DNA

damage by producing superoxides (Huycke *et al.*, 2002). Gut microbiota also influences the circadian clock (Leone *et al.*, 2015). *Firmicutes* and *Clostridia* abundance can resolve the issues of cow milk allergy if made present at the age of 3 to 6 months old. *Clostridium sensu stricto* is responsible for IgE levels in serum (Ling *et al.*, 2014; Atarashi *et al.*, 2011).

Clostridium species that belong to clusters IV, XIVa, and XVIII produce SCFAs and thereby induce Foxp3<sup>+</sup>, CD4<sup>+</sup>, and Treg cells (Atarashi et al., 2013) while Bacteroides fragilis binds to TLR2 (Toll-like receptor 2) of dendritic cells (DCs) to enhance immuno-tolerance through the production of cytokine IL-10 by regulatory T cells (Das Gupta et al., 2014). Short-chain Fatty Acids (SCFAs) produced by beneficial microbiota mediate their functions by activation of G-protein coupled receptors (GPCRs) such as GPR41 and GPR43. These also block the action of histone deacetylase (HDAC) (Zaibi et al., 2010). SCFAs produce chemokines such as CINC-2 and MCP-1, cytokines like TNF-α, IL-10, IL-2, and IL-6 and help leucocytes to migrate to inflammation sites and destroy pathogens (Vinolo, 2011; Cox et al., 2009; Luster et al., 2005). SCFAs also cause apoptosis of lymphocytes, neutrophils, and macrophages (Aoyama et al., 2010; Bailón et al., 2010; Fei and Zhao, 2013) (Fig. 3).

## **Strategies**

Microbiota therefore is an essential organ of the human body and its total absence impairs social behavior leading to anxiety and stress response (Desbonnet *et al.*, 2014; Foster *et al.*, 2013). Various microbiome modulation-based therapeutic strategies are to be followed for modulating gut microbiota towards a healthy one and form the body of the review (Fig.4).

## **Dietary Interventions**

The dynamic ecosystem of gut microbiota depends upon various factors such as age, lifestyle, environment, genetic predisposition and mainly diet. Diet along with other factors impacts the gut microbiota and brain-gut axis, which in early stages help in protection against the onset and progression of disorders linked with development and mental health. Certain dietary interventions serve as alternative or adjuvant strategies for the management of neurodegenerative disorders (NDDs) emphasizing long-term consequences on health and aiming at their prevention and treatment.

#### Fat-rich diet

Maternal high-fat diet-induced obesity during pregnancy can be managed by modifying the gut microbiota composition of the offspring thereby determining the socio-emotional behavior and cognitive abilities of the child (Rivera et al., 2015). Supplementation with Bifidobacterium breve strain from birth promotes the growth of the resident Bifidobacteria to mitigate negative consequences associated with C-section progeny such as social behavior and anxieties. Infants with longer duration of breastfeeding have increased levels of Bacteroides fragilis and Lactobacillus to restore healthpromoting microbiome in Caeserian-born progeny (Coker et al., 2021). Dietary supplementation of omega-3-polyunsaturated fatty acids (PUFAs) in the early life period provides protection against neurological impairments (Lei et al., 2013; Gow et al., 2013).

A high-fat diet is known to alter the gut microbial composition and is associated with cognitive decline and ADHD (attention deficit hyperactivity disorder), yet pieces of evidence for ADHD management based upon gut microbiota-directed interventions need further confirmation (Fernandez-Real *et al.*, 2015). However, prenatal and early infancy supplementation with *Lactobacillus rhamnosus* GG (LGG) shows a decreased risk of developing ADHD (Rianda *et al.*, 2019). A high saturated fat dietalso leads to an increase in Proteobacteria and Firmicutes and a decrease in Bacteriodetes, increasing gut permeability, insulin resistance, and adipose tissue inflammation (Malesza *et al.*, 2021).

Milk fat intake causes gut inflammation due to the thriving of sulfate-reducing bacteria (Devkota et al., 2012). Monounsaturated fatty acids (MUFAs) enhance the richness of Prevotella, Enterobacteriaceae family members, Parabacteroides, and Turicibacter genera showing positive health effects and increasing diversity of gut microbiota (Wolters et al., 2019). A diet rich in coconut oil increases Lactobacillus, Allobaculum, Clostridium, Staphylococcus, and Firmicutes hence adipose tissue inflammation (de Moura e Dias et al., 2018). Polyunsaturated fatty acids (PUFAs) have the potential to produce butyrate and Lachnospiraceae taxa (Noriega et al., 2016). Prolonged high saturated fat intake causes high production of bacterial amyloids which is associated with misfoldings of proteins and enhanced neuroinflammation. High-fat diet intake leads to an abundance of Enterococcaceae.

Roseburia, Staphylococcus, Dorea, and Coprobacillus with reduced cognition abilities and slow brain metabolism (Sanguinetti et al., 2018).

# Gluten-free, casein-free, and food additives-rich diet

Foods rich in salicylates and artificial additives contribute to ADHD development. Gluten-free and casein-free diets (GFCF) are found to be effective for spectrum disorder (ASD) treatment (Paniwowarczyk et al., 2018). GFCF diets do not produce opioid peptides BCM7 (β-caso morphin7) when subjected to partial digestion of gluten and casein peptides which otherwise correlates with negative effects on mental health. These food-derived opioid peptides tend to cross the blood-brain barrier to reach and bind with their respective receptors in the brain and act as neuromodulators to affect neurotransmission and unfold the pathogenesis of ASD (Jarmolowska et al., 2019). Gluten tolerance can be assumed by a gluten-free diet when taken with prebiotics and probiotics with concomitant increase in Clostridiaceae, Victivallaceae and Coriobacteriaceae families and reduced Roseburia feces and Ruminicoccus bromii causing a possibility to cure celiac disease (Bonder, 2016). Certain food additives such as aspartame, saccharin, and sucralose increase Bacteroides spp and decrease Bifidobacterium and Clostridium spp leading to glucose intolerance (Suez et al., 2014).

## **Ketogenic Diet**

Ketogenic diet has been found to have a high therapeutic potential for pediatric epilepsy and many other mental disorders (Castro et al., 2015). The Keto diet (KD) leads to reduced levels of tumor necrosis factor (TNF-α) in plasma and Interleukin-1β (IL-1β) in the brain with favorable effects in Schizophrenia (SZ) patients (Dupuis et al., 2015). KD intake is known to trigger loss of microbial diversity with decreased Bifidobacteria and increased E.coli, Shigella, Akkermansia muciniphila, and Parabacteroides merdae resulting in protection against epileptic seizures (Olson et al., 2018). Oral intake of Lcarnitine and phosphatidylcholine is reported to elevate levels of trimethylamine (TMA) and its proatherogenic metabolite trimethyl amine-N-oxide (TMAO) with increased risk of atherosclerosis and associated cardiovascular diseases (CVDs) (Koeth et al., 2013). TMAO contributes to atherosclerotic plaques, increased coronary plagues, plague-rupture possibilities, and

enhanced risk of the coronary syndrome and myocardial infarction (Zhu *et al.*, 2016; Tang *et al.*, 2013; Tang *et al.*, 2015) (Fig. 5).

The onset of depression induced by an inflammatory cytokine interferon- $\alpha$  (IFN- $\alpha$ ) can be avoided by pretreating with eicosapentaenoic acid (EPA) (Su *et al.*, 2014). Intake of the ketogenic diet leads to an increase in *Desulfovibrio spp.* and *E. coli* with a corresponding rise in gut inflammations and a decrease in *Bifidobacterium* and *Dialister* (Lindefeldt *et al.*, 2019).

Early introduction of the ketogenic diet modulates the gut microbiome to enhance brain vascular functions and improve cognition and memory via the production of SCFAs which reduces leakiness of the gut and limits the diffusion of lipopolysaccharides (LPS), thereby reducing the risks for Alzheimer's disease (AD) and improving the overall health of the brain.

Release of gamma amino butyric acid (GABA) for the overall health of the brain is attributed to increased levels of Akermansia muciniphila, Bacteroides fragilis, Dialister invisus, and Bifidobacterium adolescentis upon intake of Mediterranian keto diet. Western diet on the other hand leads to an abundance of Helicobacter pylori, Prevotella, Campylobacter, and Porphyromonas with subsequent release of LPS, the later enters the bloodstream to activate TLR-4 dependent CD14 and causes damage to the intestinal barrier to enhance permeability of gut and release of pro-inflammatory cytokines (Kim et al., 2021; Qin et al., 2007). With the keto diet, the neurotransmitter epinephrine, GABA release is enhanced with decreased production of reactive oxygen species (ROS), biogenesis of mitochondria, hyperpolarization of neurons, and energy metabolism gene upregulation (Bough and Rho, 2007).

These begin a cascade of synaptic disruption and neuroinflammation that is the root cause of neurodegenerative diseases.

## **High and Low-Calorie Diets**

A very low-calorie diet fed to obese patients for 6 weeks is known to alter gut microbiota (Simões *et al.*, 2014). Obesity and non-alcoholic fatty liver disease (NAFLD) can be prevented by making alterations in gut microbe composition based on diet (Henao-Mejia *et al.*, 2012). Reduction in dysbiosis occurs with dietary intervention to alter the composition of microbiota to a favorable and

stable state such as a decrease of *Acinetobacter spp* and increase of *Corynebacterium*. Decreased *Actinobacter* is correlated with enhanced cytokine response in patients with chronic mucocutaneous candidiasis (CMC) and hyper immunoglobulin E syndrome (HIES) with fungal pathogens *Candida albicans* and *Staphylococcus aureus* for fighting against these two immunodeficiencies (Smeekens *et al.*, 2014).

The prevalence of Akkermansia in the gut largely determines the basal metabolic rate in humans (Xu et al., 2022). For ulcerative colitis (UC) remission, an antiinflammatory diet is highly recommended (Kedia et al., 2022). High sugar intake induces colon inflammations by increasing the levels of Akkermansia muciniphala, enhancing gut permeability, and reducing the production of SCFAs. This is mediated by the production of an enzyme that degrades the mucus layer (Khan et al., 2020; Laffin et al., 2019). Probiotic intervention declines recovery of endotoxin from the gut of obese, diabetic, and hypertensive subjects from 35% to non-detectable resolved issues of traces with hypertension, hyperinsulinemia, and hyperglycemia (Fei and Zhao, 2013).

## **Animal and Plant-Based Proteins**

Dietary intake of animal protein-based and dairy products leads to an abundance of *Bacteroides, Bilophila,* and *Alistipes* which further increase TMAO, nitrosamines, and ammonia. TMAO with pro-atherogenic potential leads to an increased risk of CVDs (David *et al.*, 2014; Barrea *et al.*, 2019; Zhang *et al.*, 2022).

The animal protein-based diet also exacerbates the growth of *Desulfovibrio spp* that produces H<sub>2</sub>S and increases gut inflammation. On the contrary, plant-based proteins increase the abundance of beneficial bacteria such as *Eubacterium faecalibacterium*, *Clostridium*, *Roseburia*, *Bifidobacterium*, and *Lactobacillus* while decreasing *Bacteroides* and *Clostridium* (Swiątecka *et al.*, 2011; Graf *et al.*, 2019).

## Polyols and Polysorbate-rich diet

Polyols induce laxative effects and also an increased *Bifidobacterium* in humans. Polysorbate 80 and carboxymethyl cellulose can promote gut inflammation with an increase of *Akkermansia muciniphila* and proteobacteria (Ruiz-Ojeda *et al.*, 2019; Chassaing *et al.*, 2015; Chassaing *et al.*, 2017).

## Vegetarian and Vegan diets

Vegetarian and vegan diets lead to an abundance of *Bacteroides, Clostridium, Faecalibacterium,* and *Klebsiella* and lower levels of *Bilophila* (Matija si c *et al.*, 2014; Ruengsomwong *et al.*, 2016).

Dietary interventions have a significant impact on gut microbiota but short-term and long-term interventions need a detailed study to see if the short-term intake has a long-standing impact on gut microbiome diversity. Beta diversity can be dealt with short-term interventions but it is hard to deal with alpha diversity. Detailed studies are required to investigate the changes in microbial composition in response to habitual dietary strategies.

If the host acquires microbial resilience, the diversity returns to its original state after interventions otherwise it tends to establish a new microbiome profile (Smits *et al.*, 2017). Therefore, diet and gut microbiota act synergistically to provide resilience against diseases.

# Effects of fasting or starvation on human gut microbiota

Fasting in humans leads to an abundance of *Christensenellaceae* species. A decreased abundance of firmicutes such as *Lachnospiraceae* and *Ruminococcaceae* and an increase of *E. coli* and *Bilophila wadsworthia* with a calorie-restricted diet is reported (Mesnage *et al.*, 2019). While Ramadan fasting led to elevated levels of *Faecalibacterium prausnitzii*, *Roseburia, Eubacterium, Akkermansia, Butyricicoccus pullicaecorum* (Ozkul *et al.*, 2020) and *Bacteroides fragilis* (Ozkul *et al.*, 2019).

Within one week of calorie-restricted diet intake, an initial increase in abundance of *Lactobacilli* and *Enterobacteria* is seen with a subsequent decline by the end of the intervention. Genera *Clostridium-XIV*, *Coprococcus*, and *Lachnospiracea* decrease due to fasting while *Faecalibacterium* is known to increase (Ozkul *et al.*, 2020). The abundance of *Lachnospiraceae*, *Blautia* and *Faecalibacterium* is observed after intermittent fasting. These bacterial genera produce butyrate in the gut and counterbalance dysbiosis in the gut of patients suffering from multiple sclerosis (MS). Therefore, dietary programs and fasting regimens can serve as important non-pharmacological interventions for the treatment of various diseases.

## **Nutritional Supplements**

Protective effects of specific nutrients like zinc, iron, iodide, and omega-3-PUFA and adverse effects of food coloring agents, sugar, and preservatives are found to be associated with increased risk of ADHD and obesity (Pelsser et al., 2020; Bowling et al., 2017). A deficiency of vitamins such as A, C, B<sub>6</sub>, B<sub>12</sub>, D, and folate has been found to trigger an onset of ASD (Fraguas et al., 2019). Deficit of nervonic acid (NA), a monounsaturated omega-9-fatty acid is known to develop psychosis in patients with high clinical risk (Amminger et al., 2012), while omega-3-fatty acid supplementation tends to reduce psychotic conversion rates (Moon, 2010; Cadenhead et al., 2017). Elevated Hcy serum levels (homocysteine) are related to the development of cardiovascular diseases, SZ, and Alzheimer's disease (Tinelli et al., 2019). A low level of serum Vitamin D is observed in SZ patients while Vitamin D is found to have a negative correlation with psychosis severity (Gracious et al., 2012). Low dietary intake of vitamin C is associated with an increased risk of SZ. So, clinical management of SZ, in patients is diet dependent and also associated with celiac disease (Cha and Yang, 2020). Alterations in the diet by introducing protein supplementation can have asignificant imbalance in gut microbiota. In studies conducted, a diet complemented with protein intake was found to be correlated with an increase in the population of Bacteriodetes and a decrease in health-related taxa including *Bifidobacterium* longum, Blautia and Roseburia. Also, Lactobacilli and other butyrate-producing bacteria are reduced in abundance (Ma et al., 2017) along with a decrease in SCFAs-producing phylum such as Coprococcus. Protein supplements with probiotics could form a part of future strategies to mitigate the imbalance of microbiota in place of only protein supplementation and can result inrecovering the dysbiosis. However, further research is required to determine sources and doses of protein in support of health benefits for the sports community. Therefore, long-term protein supplementation with proteins hurts gut microbiota. Protein overfeeding also leads to lower levels of malondialdehyde which is a marker for oxidative stress and it can lead to changes in populations of microbiota and their metabolites (Moraes et al., 2017). Vitamin A supplementation increases Bacteroidetes and decreases in Actinobacteria, Bifidobacterium, Clostridium, Enterobacter, Escherichia, and *Proteobacteria* populations.  $\beta$  -carotene intake results in decreased Bacteroides, and an increased

Firmicutes and Clostridium (Liu et al., 2017; Li et al., 2017). Increased Proteobacteria and Actinobacteria result due to retinol intake (Mandal et al., 2016). The duration of E.coli infections after supplementation with vitamin A is due to a decrease in IL-8 and monocyte chemoattractant protein-1 concentrations (Long et al., 2011).

Vitamin B<sub>2</sub> provides an anti-oxidative environment in the gut which favors the growth of Faecalibacterium prausnitzii and reduces E. coli colonization, thereby decreasing pro-inflammatory processes and hence applications **IBD** finding in treatment. supplementation is associated with reduced inflammatory effects due to low oxidative stress, C-reactive proteins, low IL-2, and decreased erythrocyte sedimentation rate (ESR) (von Martels et al., 2019). An increased intake of Vitamin B<sub>2</sub>, B<sub>5</sub>, B<sub>6</sub>, and B<sub>12</sub> leads to a corresponding increase in the abundance of Prevotella and a decrease in Bacteroides populations (Carrothers et al., 2015). Also, deficiency of cobalamine (Vit B<sub>12</sub>), vitamin B<sub>9</sub> (folic acid), and piroxidine (Vit B<sub>6</sub>) is found to be associated with schizophrenia (SZ) development. In other words, vitamin B supplementation results in increased microbial interactions, metabolism, and signaling besides ensuring enhanced microbial diversity, specifically beneficial microbes that are enriched by Vitamin A, D, and E.

Vitamin C, E, and B<sub>2</sub> reduce redox potential. Vitamin C ensures increased SCFA production and exhibits antibacterial, antiviral, and antimicrobial properties (Mousavi et al., 2019). Vitamins A and D lead to increased immune function and also enhance barrier functions. Diseases of the gastrointestinal tract such as inflammatory bowel disease are correlated with a deficiency of vitamin D (Cross et al., 2005) which alters microbiome diversity with an increased Coriobacteriaceae, Streptococcus, Bifidobacterium, Dorea and Coprococcus while decreased Odoribacter and Desulfovibrionaceae (Pham et al., 2021; Chatterjee et al., 2020).

Vitamin D supplementation also reduces the abundance of gamma proteobacteria like *Pseudomonas, E.coli,* and *Shigella. Haemophilus, Blautia,* and *Veillonella* while increased *Prevotella* and *Lachnospira.* Higher intake of vitamin E results in decreased *Proteobacteria.* Vitamin E and iron supplementation lead to the enrichment of butyrate-producing bacteria (Tang *et al.,* 2016) and decreased vitamin E can enhance pathogenic *Citrobacter* in mice models (Smith *et al.,* 2011). Vitamin K supports

bacterial diversity in the gut microbiome (Fenn *et al.*, 2017).

#### **Antibiotics**

Antibiotics play a significant role in eradicating some diseases. Abundance of overgrown *colitis* bacteria in patients can be reduced with certain antibiotics. Similarly, antibiotics such as Minocycline and Sulfazalazine reduce symptoms of rheumatoid arthritis patients which target the causative microbes that are bacteria (Stone *et al.*, 2003). Administration of antibiotics during pregnancy, infancy, childhood, or even during adolescence influences colonization and diversity of gut microbiome to determine various neurocognitive disorders later in life (Younge *et al.*, 2019; Burger *et al.*, 2020)

At the same time, the use of broad-spectrum antibiotics results in short-term and long-term perturbation of diversity in the resident microbiome community of the host that has detrimental impacts leading to the killing and reduction of beneficial microbes such as *Faecalibacterium prausnitzii*to cause deleterious effects on host health too. For instance, low levels of *F. prausnitzii* are found to be prevalent in IBD patients (Sokol *et al.*, 2009; Dubourg *et al.*, 2014). Antibiotic-associated diarrhea (AAD) can be another outcome of the elimination of beneficial gut microbiota, in the absence of which *Clostridium difficile* gets a chance to thrive well and can further lead to diarrhea and colitis (Buffie *et al.*, 2012; Mc Donald 2017).

Decreases in accompanied serotonin levels, secondary bile acids, and tryptophan hydrolase influence gut motility (Ge et al., 2017). Also, there is a decrease in SCFA production with corresponding low levels of butyrate and propionate. (Mu et al., 2017) The absence of useful microbes under antibiotic influence results in immunological and physiological changes to the gut environment. Mucus thickness and gastric motility are reduced with improper functioning of intestinal cells and immune cells (Cehenzli et al., 2013) and chances of invasion by pathogens and subsequent inflammations become high (Wlodarska et al., 2011). Barrier functions of intestinal cells are also impaired which can lead to the development of ulcerative colitis, Salmonella, and Helicobacter infections (Machills et al., 2013; Gillis et al., 2018).

Altered microbial composition due to antibiotics results

in deficiencies of certain vitamins and metabolites that are produced by beneficial bacteria. Antibiotic intervention alters the gut microbiota and is known to reduce the severity of multiple sclerosis (MS) in human murine models. This is attributed to reduced secretion of pro-inflammatory cytokines such as IL-17.

Antibiotics therefore create persistent off-target microbiome disturbance called dysbiosis that can further lead to disorders and diseases related to host immunity and result in increased abundance of antibiotic-specific resistance genes. Antibiotics result in reduced colonization resistance with a concomitant increase in susceptibility to specific opportunistic pathogens such as Azithromycin-resistant *Clostridium difficile, Salmonella,* and *Vancomycin-resistant Enterococci* (Brandl *et al.,* 2008; Sequeira *et al.,* 2020; Wu *et al.,* 2020).

More research is needed to understand how different antibiotics alter gut microbiomes to better regulate the indiscriminate usage of antibiotics and avoid risks of antimicrobial-resistant infections. Consistent use of antibiotics during childhood results in the development and abundance of bacterial antimicrobial resistant (AMR) genes called resistome which posesa risk of obesity, diabetes, asthma, allergy, and later adiposity in life (Block et al., 2018; Marra et al., 2009; Chen et al., 2021; Stokholm et al., 2014). Class of antibiotics, duration of administration, usage, and follow-up time impact the diversity and composition of the microbiota. The use of other therapies such as probiotics, synbiotics, and FMT (Fecal Microbiota Transplantation) must be emphasized post-antibiotic treatment to restore the gut microbiota community.

Several therapies such as bacteriophage therapy, bacteriocins use, FMT, probiotics, synbiotics, and use of monoclonal antibodies are important alternatives to the use of antibiotics that pose target-specific results with minimal damage to the microbiota thus emphasizing the need to develop therapies in addition to methods of conserving and restoring the perturbed microbial-communities post-antibiotic treatments.

#### **Probiotics**

The live microorganisms which when consumed in adequate amounts bring about health benefits to the host or active bodies with essential functions for promoting health aspects (Gasbarrini *et al.*, 2016).

Lower levels of Bifidobacteria and Blautia gut bacteria

in mice are linked with the production of less tryptophan which acts as the serotonin precursor (Golubeva et al., 2017). Bacteroides fragilis or Lactobacillus reuteri, when administered, bring about gastrointestinal and behavioral changes in ASD (Pama, 2019). ASD-suffering children report GI pain and increased levels of Clostridiales, decreased levels of Veillonellaceae, Coprococcus, and Prevotella (Luna et al., 2017), butyrate-producing taxa Ruminococcaceae, Eubacterium, Lachnospiraceae, and Erysipelotrichaceae together with low production of SCFAs in adults (Liu et al., 2019). Selected strains of Bifidobacterium longum infantis and B. bifidum can abolish food-derived opioid peptides to contribute to host health (Olivares et al., 2018). Different probioticstrains such as Lactobacillus rhamnosus. L. reuteri, and Bifidobacterium infantis, when administered in combination, reduce incidences of seizures and epileptic activity due to increased GABA levels in the brain and accompanied by reduced oxidative stress in the brain (Bagheri et al., 2019). A reduction in Bacteroides, Prevotella, Bacteroides fragilis, Bifidobacterium and Eubacterium, Clostridium, and Coccoides groups is seen in rheumatoid arthritis patients (Scher et al., 2013). When inoculated in a probiotic combination, this provides chances for rheumatoid arthritis management. Also, members of families Lactobacillaceae of Firmicutes, Rikenelleceae, and Porphyromonadaceae suggest an immune-regulative role in diabetes.

Probiotics strains including *Bacillus spp* (*Bacillus breve*, *B. bifidum*, *B. subtilis*, *B. longum*, *B infantis*), *Lactobacillus casei*, *L. acidophilus*, *L. delbrueckii*, *L. helveticus*, *L. plantarum*, *L. salivarius*, *L. rhamnosus*, *Lactococcus lactis lactis*, and *Streptococcus thermophiles* when administered in MS patients, reduce IL-6 and BDNF levels (Rahimlou *et al.*, 2020) with a decreased release of IFN-γ, IL-1β and increased expression of anti-inflammatory IL-4, IL-5, and IL-8 (Dargahi *et al.*, 2020). Gastrointestinal bacteria activate stress circuits via vagus pathways (Lyte *et al.*, 2006).

A probiotic mixture of *Bacillus mesentericus*, *Clostridium butyricum*, *and Streptococcus faecalis* is known to reduce the symptoms of Schizophrenia (SZ) (Nagamine *et al.*, 2012). While a mixture of ten strains of *Lactobacillus* and four strains of *Saccharomyces* upregulates G-protein-coupled receptor 43/41 (GPR 43/41) and triggers GLP-1 secretion with subsequent release of insulin (Wei *et al.*, 2015) and holding the potential for management of diabetes. Certain probiotics show beneficial effects in relieving stress-related behaviors and

anxiety thus modulating depressive states in rats. Parallel studies are required in humans too (Dinan *et al.*, 2013). Administration of *Lactobacillus plantarum* is correlated with a reduction in infarct size and improved functioning of the left ventricle after myocardial infarction. *L rhamnosus* is known to reduce left ventricle hypertrophy. A high abundance of *Odoribacter* is associated with low blood pressure in obese people due to more production of butyrate and other SCFAs (Gomez-Arango *et al.*, 2016). Therefore probiotics used in adjunctive medications give more reliable and effective outcomes in various cardiovascular disorders (Lam *et al.*, 2012; Gan *et al.*, 2014).

The use of probiotics and synbiotics can prove as a good option to restore the microbiota community post-antibiotic treatment. Probiotics produce antimicrobial peptides and bacteriocins and together these suppress the growth of non-commensals, enhance the barrier function of the gut, and modulate immunity (Cazorla *et al.*, 2018). FMT supplementation followed with three strains of *Lactobacillus* can restore IL-22 production and reduce inflammations which opens avenues of probiotics as therapeutic agents. However, FMT is considered more beneficial as probiotics use cannot regain the entire microbial balance in the gut (Suez *et al.*, 2018).

#### **Prebiotics**

Resistant to gastric acidity and digestive enzymes of the host, non-absorbable, fermentable by gut bacteria, and having the potential to activate gut microbiota selectively, these probiotics are exploited towards the human interests of maintaining good health. Galactooligosaccharides (GOS) and resistant starches (RS) can stimulate *Bifidobacteria*, *Lactobacilli* largely and *Bacteroidetes*, *Enterobacteria*, and *Fermicutes* to some extent (Louis et. al. 2016, Ze, *et al.*, 2012).

These bacteria degrade prebiotics to produce SCFA which are small enough to diffuse from enterocytes and impact extra-intestinal organs such as the central nervous system (CNS), cardiovascular system, immune system, etc. Prebiotic supplementation resultsin an elevated *Bifidobacteria* population to improve Crohn's disease, IBS, IBD, and even UC (Lindsay *et al.*, 2006). Dysbiosis of microbiota in Crohn's disease is well elucidated with low levels of immunoregulatory *Faecalibacterium prausnitzii* (Sokol *et al.*, 2009). Prebiotics show antiadherence properties and prevent the binding of bacterial pathogens to attachment sites by disrupting adhesion-

oligosaccharides, thus reducing the chances of gastrointestinal infections (Sharon, 2006). Similarly, pectic oligosaccharides, chitin oligosaccharides, and mannose oligosaccharides also act as adherence agents.

Synbiotic therapy comprising of *Bifidobacterium lactis*, Lactobacillus, Rhamnosus, and inulin reduce the chances and rate of colorectal cancer by improving the intestinal barrier strength (Candela et al., 2011; Pool-Zobel, 2005). The population of harmful pathogens can be kept under check by prebiotics through direct binding or cytokine production. Prebiotic metabolites tend to improve the fetal immune system by crossing through the placenta. Prebiotic intake also reduces the use of antibiotics and reduces the duration of the disease. B( $2\rightarrow 1$ ) fructans lead to an abundance of interleukin-4 (IL-4) in serum, toll-like receptor-2 (TLR-2) mediated immune response, CD<sub>282+</sub>/ TLR<sub>2+</sub> myeloid dendritic cells (Clarke et al., 2016). Blood levels of IL-8, IL-1β, IL-10, and C-reactive protein (CRP) become high due (galactooligosaccharides) intake and also better working of NK cells consuming GOS (Vulevic et al., 2008; Vulevic et al., 2015). This leads to reduced expression of IL-6 and phagocytosis in monocytes and granulocytes (Guigoz et al., 2002). FOS (Fructooligosaccharides) and GOS show regulatory effects on neurotransmitters Nmethyl-D-aspartate (NMDA) and synaptic disorders by altered gut microbiota diversity (Smith et al., 2015; Waworuntu et al., 2014). Higher gastrointestinal disorders are correlated with autism (Adams et al., 2011). known FOS supplementation is to restore Bifidobacterium, Bacteroides, Clostridium, Roseburia, and Phascolartobacterium faecium with a positive impact on anorexia nervosa (Liu et al., 2021).

Lactulose has preventive potential for hepatic encephalopathy (Mudd et al., 2016; Müller et al., 1966) by lowering the pH of the lumen and reducing the production of ammonia due to inhibited glutaminase activity. The side effects of lactulose like nausea and flatulence are overcome by another prebiotic lactitol which is better than lactulose and equally effective for curing hepatic encephalopathy (Blanc et al., 1992). Prebiotics play a significant role in improving allergic skin problems like atopic dermatitis (Grüber et al., 2010), lowering the risk of CVD, reducing the level of total cholesterol, low-density lipoprotein (LDL) (Tiwari et al., 2011) and inflammatory elements. Non-alcoholic fatty liver disease (NAFLD) can be managed with prebiotics more efficiently along with medication (Tarantino and Finelli, 2015) and also enhances calcium absorption and

acts as a promising agent to improve the overall health of humans. A link has been found to exist between gut microbiota and NAFLD (Abu-Shanab and Quigley, 2010), non-alcoholic steatohepatitis (NASH), and obesity (Chierico et al., 2017). Risk of sepsis, bacteremia, and malnutrition endocarditis in compromised patients are attributed to prebiotics due to an incompetent intestinal epithelial barrier (Tsai et al., 2019). Intake of low dietary prebiotics reduces saccharolytic bacteria specifically Bifidobacteria. SCFAs produced from the action of colonic bacteria on prebiotic fibers modulate cytokine production and expansion of Tcells to favor the homeostasis of colonocytes and decrease the possibilities of inflammations (Lee and Hase, 2014).

In diabetes T2DM, butyrate-producing bacteria are scarce while *Clostridiales sp.*, *Eubacterium rectale*, *Faecalibacterium prausnitzii*, *Roseburia inulinivorans* and *Roseburia intestinalis* are enriched. Pathogens like *Bacteroides caecae*, *Akkermansia muciniphila*, *Desulfovibrio*, *E.coli*, and *Clostridium* are also enriched (Qin *et al.*, 2012).

For the reduction of resident gut microbiota-based disorders and keeping in mind the needs of each society and specific population, designing specific prebiotics is a challenging task and a topic of future research.

## **Synbiotics**

Probiotics and prebiotics when consumed as a mixture constitute synbiotics. This is based upon formulating probiotics such as Bifidobacterium and Lactobacillus clubbed with prebiotics such as inulin, cellobiose, psyllium, GOS, lactitol, FOS, and β-glucan for different time durations. Synbiotics have the potential to modulate microbiota and benefit diseases such as IBS, obesity, diabetes, infections, and chronic kidney disease with significant enrichment of Bifidobacterium (McFarlane et al., 2019). A combination of B. longum and inulin and another combination of Lactobacillus and GOS enhances the Bifidobacterium to different extents. Synbiotics comprising Lactobacillus plantarum, lactulose, and arabinose effectively adjust the blood glucose, lipid profile, and body weight of T2DM patients to ideal levels. Lactulose produces low molecular weight organic acids to reduce the pH of the intestine (Aït-Aissa and Aïder, 2014), and L-arabinose checks sucrose breakdown to reduce weight and manage diabetes (Panesar and Kumari, 2011). The presence of L. plantarum is

favorable for the thriving of Bifidobacterium which in turn reduces plasma endotoxins and ammonia in the blood. Therefore, synbiotics on the whole serve to play a role in detoxification, hypolipidemia, and hypoglycemia in T2DM patients as compared to a single probiotic or prebiotic. Synbiotics, therefore maintain homeostasis of intestinal flora and provide a new solution for managing T2DM. Lactobacillus rhamnosus, in a combined interventional approach for immunotherapy, provides promising results for the prevention of food allergies as compared to probiotics alone, thereby corroborating synbiotics as a more integrated, refined, and successful approach for the management of diseases through gutmicrobiome modulation. The release of bacterial toxins and production of pro-inflammatory cytokines also contribute to dysbiosis that needs to be addressed to with synbiotics intervention. Total Bifidobacterium spp., Actinobacteria. Actinobacteriota. Lactobacillus. Parabacteroides. Ruminococcaceae. Firmicutes. Methanobrevibacter, Prevotella enrich upon synbiotics supplementation Enterobacteriaceae, whereas Bacteriodetes, Proteobacteria. Desulfovibrio, Oscillospira, Verrucomicrobiota, Bacteroidota, Akkermansia muciniphila, Roseburia and Zonulindecline in population (Ouwehand et al., 2009; Costabile et al., 2017; Kanazawa, 2021; Krumbeck et al., 2018).

The effect of synbiotics on gut microbiome results in enhanced SCFA-producing microbes and corresponding improvised gut health. Increased production of butyrate and propionate with pronounced anti-inflammatory effects and a more preserved intestinal barrier with more expression of inflammatory cytokines have a direct impact on immune and neuronal modulation (Gill et al., Alloprevotella, 2018). Microbes such as Ruminococcaceae, Prevotellaceae, and Catenibacterium are more abundant with the intake of a synbiotic fiberrich diet (Neyrinck et al., 2021) and mediate their action by improving the intestinal barrier (Chen et al., 2019), reducing inflammations, increasing more SCFAs producing bacteria such as Eubacterium ruminantrim, modulating action of enzymes to manage obesity and showing the synergistic effect on T2DM. On the contrary, the intake of synbiotics results in decreased abundance of pathogenic Erysipelato, and Clostridium and with corresponding low chances of diet-induced obesity (Jo et al., 2021; Mukherjee et al., 2024; Oh et al., 2021). Synbiotics lead to an abundance of Alistipes and Parasutterella to maintain bile acid levels and regulate the metabolism of cholesterol (Mukherjee et al., 2020) with the later while Alistipes offers protection against

colitis, fibrosis, cancer, and CVD (Parker et al., 2020). Decreased Ruminococcus gnavus is detected due to synbiotics intake with the corresponding effect of the same against Crohn's disease. Dialister and Prevotella increase upon intake of Bifidobacterium bifidum, B. longum, Lactobacillus acidophilus, and L. rhamnosus as synbiotics. Prevotellaceae UCG-003 acts as a modulator of inflammations in the intestine. Streptococcus salivarius increases due to synbiotic intake with corresponding reduced inflammations (Kaci et al., 2014) while Colidextribacter levels are lowered with corresponding reduced inflammatory metabolites (Gu et al., 2022).

The potential of *Ruminococcus albus* and *R. flavefacines* lies in producing butyrate to provide energy to the intestinal cells while acting as anti-cancerous entities (Sergeev *et al.*, 2020).

Further research is required to work out the best possible combinations of probiotics and prebiotics, their proportions, dosage, duration, and mode of delivery for the most effective regimens to meet the need for microbiota-modulating therapies.

## **Postbiotics**

inanimate microorganisms bioactive Various or compounds including inactivated cell preparations or microbial cells metabolites, non-toxic and non-viable compounds, or food constituents derived from microorganisms that offer health benefits come under the category of postbiotics. These include SCFA, bacteriocins. **EPS** (exopolysaccharides), vitamins. enzymes, and peptides in inactivated cell preparations and mediate their health effects by strengthening the gut barrier, reducing gut inflammations, and promoting antimicrobial actions against gut pathogens. SCFAs activate their receptors on intestinal epithelial cells to mediate signals for the maintenance of the epithelial barrier and also for the regulation of the immune system (Sun et al., 2016). All the five SCFAs viz. acetic acid, propionic acid, butyric acid, pentanoic acid, and hexanoic acid increase after intake of postbiotics along with the increase of other metabolites such as p-methoxy cinnamic acid and α-linolenic acid while piperine, capsaicin, theophylline, phenylalanine, tryptophan, 5hydroxy tryptophan, aromatic amino acids, kynurenine and other related metabolites decrease. A primary bile acid Chenodeoxycholic acid also decreases which is linked with diarrhea (Yu et al., 2018; Panpetch et al.,

2021; Hill *et al.*, 1991; Chaiyasit and Wiwanitkit, 2016; Zheng *et al.*, 2020).

Intestinal motility is regulated by propionic acid, valeric acid, citric acid, malic acid, lactic acid, and butyric acid through different mechanisms such as affecting colonic smooth muscles functioning, vagus nerve and stimulating mucosal receptors (Rondeau *et al.*, 2003) by altering intestinal pH and eliminating harmful bacterial metabolites (Sun *et al.*, 2017; Bosi *et al.*, 2007). Production of anti-inflammatory interleukin IL-10 is enhanced by 3-indole acrylic acid to suppress cytokine IL-6 and IL-1β, posing a good anti-inflammatory effect. 5 HT (5 hydroxy tryptamine) mediates its effect by enhancing the gut mucosal barrier to elicit anti-diarrheal properties (Iancu *et al.*, 2023). Heat-treated *Lactobacillus* (LB) has the potential to mitigate diarrhea which is accompanied by post-antibiotic treatments.

Anxiety and sleep disturbance associated with diarrhea are taken care of by postbiotic interventions effectively (Nishida *et al.*, 2019). Quality of life in IBS patients is improved by *Lactobacillus* (LB) by relieving bloating, and abdominal pain, reducing weekly stools and frequency of diarrhea (Nocerino *et al.*, 2017).

Butyrate is anti-inflammatory and anti-carcinogenic (Barcenilla et al., 2000). UC, Crohn's disease, and colorectal cancer patients become devoid of butyrateproducing bacteria. Propionate also has anti-carcinogenic effects and has the potential to check hypertensive cardiovascular damage. While acetate shows antiinflammatory effects. Therefore, butvrate propionate-rich postbiotic preparations produced from bacterialcultures are administered to handle diseased conditions. Butyrate-producing bacteria being foodunsafe are replaced by food-safe lactate-producing bacteria which produce lactate and subsequently later can be converted into butvrate or propionate via different pathways (Flint et al., 2014; Gänzle, 2015; Louis et al., 2022) to rule the option of foodborne infections from Salmonella typhimurium (Barbara, 2006).

Similarly, EPS from *Lactiplantibacillus plantarum* improves the strength of the intestinal barrier by enhancing the tight junction proteins expression and preventing pro-inflammatory cytokine expression (Zhou *et al.*, 2018; Neurath, 2014; Chen and Sundrud, 2016; Luettig *et al.*, 2015).

EPS from other bacteria such as lactic acid bacteria has anti-proliferative and anti-oxidative potential and can be

delivered to the gut from postbiotic preparations (Kodali and Sen, 2008; Sharma *et al.*, 2014). Antimicrobial peptides called bacteriocins inhibit specific microbes without affecting the beneficial ones dwelling in the gut (Gálvez *et al.*, 2007).

Postbiotics facilitate the flourishing of celluloseproducing bacteria and reduce the population of methanogens. The proliferation of beneficial genera such as Bifidobacterium, and Lactobacillus is promoted while that of pathogenic ones like E.coli and Enterococci is inhibited with postbiotics intake (Liu et al., 2021). Increased beneficial gut bacteria include Faecalibacterium, Prausnitzii, Microviridae, Fournierella. Lawsonibacter. Ruminococcu. Bifidobacterium aerophilum, Dialister hominis, Angelakisella, Dysosmobacter, Pseudoruminococcus while decreased population include Amedibacterium intestinale. Succinivibrio. Fusobacterium, Duodenibacillus, Alistipes, Megamonas funiformis, Bifidobacterium adolescentis, Blautia, Holdemania filiformis, Metalysinibacillus, Ruminococcus.and pathogens like Megamonas after postbiotic intervention (Scott et al., 2022; Thu et al., 2011; Izuddin et al., 2019). Cell wall components and cytoplasmic extracts obtained from these beneficial bacteria act as the most common probiotic producers for effective postbiotics which in turn can manage several health conditions by acting as antiproliferative, anti-inflammatory, anti-oxidative, immunomodulatory, anti-obesogenic and antibacterial therapeutic agents with plasma sugar reducing, cholesterol reducing potential (Sokol et al., 2008; Jensen et al., 2010; Jensen et al., 2007).

Lactococcus stimulates immune cells to produce cytokines for better overall. Pediococcus acidilactici and produce bacteriocins P. pentosaceus and other antimicrobial metabolites showing anti-adipogenic effects). Similarly, β-glucan obtained from Saccharomyces cerevisiae cell wall has the effective role of scavenging hydroxyl radicals and serves as an antioxidant (Pourahmad et al., 2011), and acts as postbiotic to regulate the immune system via TLR-2-MyD88-nuclearfactor (NF)-κB signaling pathway (Diaz et al., 2018; Jin et al., 2019). Lipoteichoic acid (LTA), exopolysaccharides, and cell surface proteins from Lactic acid bacteria also act as antioxidants. Mannan from yeast walls stimulates immunocytes to produce immunoglobulins and cytokines. Postbiotics have been explored for human health effects beyond the gut including skin, oral cavity, and vagina besides being implicated in commercial products for human health.

Being non-proliferative, these have to be supplied regularly and in appropriate dosage to induce positive effects on the host for successful therapeutics. Postbiotics thus serve as promising tools for the management of metabolic disorders, GI tract diseases, mental disorders, respiratory problems, cancer, etc. These tend to safeguard against bone loss with reduced insulinemia, total plasma cholesterol, reduced hyperuricemia, reduced obesity, and T2DM (Tang and Li, 2021). Also, postbiotic intake as *L. acidophilus* and *L. paracasei* result in lower chances of childhood diarrhea, gastroenteritis, otitis media, and pharyngitis in children (Humphrey and Williamson, 2001; Haukioja *et al.*, 2006; Corsello *et al.*, 2017).

## **Psychobiotics**

Psychobiotics refer to microbiota-targeted interventions which have the potential to treat psychiatric disorders. Probiotics and prebiotics together support optimal mental health by influencing microbial profiles to bring about overall homeostasis and improve cognitive abilities and are considered psychobiotics. A high abundance of beneficial bacteria like Bifidobacterium bifidum is associated with the intake of vegetables, dietary fibers, and milk products with concomitant decreased depression scores (Uemura et al., 2019). Dietary fibers and fermented foods show anti-inflammatory effects (Swann et al., 2020; Dürholz et al., 2020; Wouw et al., 2020; Marco et al., 2017). An altered gut microbiome is considered for the pathogenesis of neuropsychiatric diseases. Psychobiotics include probiotics that mediate their effect on the human brain through the gut-brain axis. Therapeutic potential of psychobiotics in various mental health outcomes such as stress, anxiety, autism spectrum disorder (ASD), depression, insomnia, anorexia nervosa, Parkinson's disease, diabetic neuropathy, and multiple sclerosis (MS),ADHD. SZ neurodegenerative disorders like dementia must be promoted and explored further to alleviate the burden of mental ailments (Sudo, 2019).

Psychobiotics mediate their action by regulating neurotransmitters such as gamma amino butyric acid (GABA), serotonin, brain-derived neurotrophic factor (BDNF), SCFAs, and enteroendocrine hormones by lowering pro-inflammatory cytokines and elevating the number of anti-inflammatory cytokines such as IL-10 (O'Mahony et al., 2015; Cheng et al., 2019; O'Riordan et al., 2022; Dinan et al., 2013). The possible underlying mechanism includes the controlled release of corticosterone and adrenocorticotrophic hormone after the recolonization of bacteria with synbiotics.

Table.1 Effects of antibiotics on colonization of human gut microbiota.

Antibiotic Class	Examples	Increasing colonization rates	Decreasing colonization rates	Reference
Penicillin	Pen V Amoxicillin Ampicillin Oxacillin Amox + clavulanate	Enterobacteria Bacteroidaceae	Bifidobacteria Lactobacilli Eubacteria Lachnospiraceae	Les Dethlefsen et al., 2008
Cephalosporins	Cefalor Cefotaxime Ceftizidine Cefuroxime Cefepime	Clostridia Bacteroides sp.	E. coli Bifidobacteria Enterobacteriaceae	Les Dethlefsen et al., 2008
Macrolides	Azithromycin Clarithromycin Erythromycin Spiramycin	Bacteroidetes Proteobacteria Resistant Enterobacteria Streptococci Enterococci	Actinobacteria Lachnospiraceae Veillonella Clostriales	Les Dethlefsen et al., 2008
Quinolone Fluoroquinolone	Ciprofloxacin Norfloxacin	Resistant <i>E.coli</i>	Lachnospiraceae Coprococcus Enterobacteriaceae	Les Dethlefsen et al., 2008
Carbapenems	Carbapenems Meropenems Ertapenem	Enterococci	Eubacteria Lactobacillus Bacteroides Bifidobacteria Streptococci Clostridia Enterobacteria	Jernberg et al., 2007
Lincomycin	Clindamycin	Enterobacteriaceae	Blautia, Bacteroides	Jernberg <i>et al.</i> , 2007

Table.2 Bacteriocins source and target bacteria.

Name of the Bacteriocin	Source	Target	References (from Emma Scott)
Curvacin A and Sakacin 1	Latilactobacillus sakei subsp. sakei	Listeria monocytogenes	Camargo <i>et al.</i> , 2018
Plantaricin L-1	Lactiplantibacillus plantarum subsp. plantarum	Listeria monocytogenes	Zhou, 2007
Plantaricin MG	Lactiplantibacillus plantarum subsp. plantarum	Listeria monocytogenes,Salmonella typhimurium	Gong, 2010
BM1157	Companilactobacillus crustorum	Listeria monocytogenes	Camargo <i>et al.</i> , 2018
Gassericin A Gassericin T	Lactobacillus gasseri	Listeria monocytogenes, Bacillus cereus,Staphylococcus aureus	Pandey, 2013
Antilisterial ABP-118	Ligilactobacillus salivarius	Listeria monocytogenes, Enterococcus,Bacillus, Listeria, Staphylococcus,Salmonella species	Gálvez, 2007; Patel, 2015

## Int.J.Curr.Microbiol.App.Sci (2025) 14(01): 72-107

Table.3 Correlation between common metabolic diseases (CMDs) and human phagosomes.

Diseases (CMDs)	Effect of phages	Reference
Metabolic Syndrome	<ul> <li>↓ Phagosome diversity &amp; richness</li> <li>↓ Clostridiaceae phages</li> <li>↓ Bifidobacteriaceae phages</li> <li>↓ Ruminococcaceae phages</li> <li>↑ CrAssphages</li> <li>↑ Bacteriophage phages</li> </ul>	Ma <i>et al.</i> , 2018; Borin <i>et al.</i> , 2023; Rasmussen <i>et al.</i> , 2020
Type 2 Diabetes	↑ Gram-negative phages ↑ Enterobacteriaceae phages ↑ Klebsiella phages ↑ Shigella phages	Han et al., 2018; Sandoval-Vargas et al., 2021; Chen et al., 2020; Ma et al., 2018; Han et al., 2018;
Atherosclerotic Cardiovascular Disease	↑ Enterobacteriaceae phages ↑ Streptococcus phages	Valles-Colomer et al., 2023; Jie et al., 2017
Pre-Hypertension	No change in diversity  ↑ Enterobacterial phage (mEp390)  ↑ Pseudomonas phage (phi2)  ↑ Cronobacter phages  ↑ Salmonella phages  ↑ Serratia phage (phiMAM1)	Yan <i>et al.</i> , 2017
Hypertension	No change in diversity  ↑ phage 86  ↑ Cyanophage (S-T1M5)  ↑ Klebsiella phage (KP32)  ↑ Salmonella phage (FSL-SP-004)	Han et al., 2018
Non-alcoholic fatty liver disease	↑ Streptococcus phages ↑ Leuconostoc phages ↑Escherichia and Enterobacteria phages ↑ Blood glucose levels ↓ Lactococcus phages ↓ BMI, HbA1C levels	Lang <i>et al.</i> , 2020; Mao <i>et al.</i> , 2023; Caussy <i>et al.</i> , 2019

Table.4 Gut dysbiosis and altered gut metabolite levels in GI Symptoms (Krishnamurthy et al., 2023).

GI Symptom	Gut Microbiota	Gut Metabolite	Altered levels
Flatulence Bacteria	Klebsiella, Pneumonia, Proteus, E.coli, Clostridium, Actinobacteria, Phascolactobacterium, Bacteroides, Coprococcus, Blautia, Bifidobacteriales, Oscillospira, Ruminococcaceae, bacteriodales, Clostridales	↑H <sub>2</sub> , CO <sub>2</sub> , H <sub>2</sub> S, CH <sub>4</sub>	<b>↑</b>
Hydrogen & Carbon Dioxide-Producing Bacteria	Bacteroidetes, Firmicutes	↑H <sub>2</sub> , CO <sub>2</sub> , H <sub>2</sub> S, CH <sub>4</sub>	1
Sulfate-Reducing Bacteria	Desulfovibrio Sp	↑H <sub>2</sub> , CO <sub>2</sub> , H <sub>2</sub> S, CH <sub>4</sub>	<b>↑</b>
Methane –Producing Archaea	M. Smithii, M stadtmanae	$\uparrow$ H <sub>2</sub> , CO <sub>2</sub> , H <sub>2</sub> S, CH <sub>4</sub>	<b>↑</b>
Constipation Bacteria	Coprococcus, Ruminococcus, Blautia, Anaerotruncus, Bifidobacterium, Lactobacillus Bacteroides, Prevotella, Roseburia	Butyrate, Acetate, Propionate, Methane ↑	<b>↑</b>
Diarrhea Bacteria	Streptococcus spp, Blautia, Faecalibacterium Lachnospiraceae, Ruminococcaceae Bacteroides, Lactobacillus, Bifidobacteriaceae	Butyrate Acetate Propionate ↓	<b>↑</b>
Diarrhea Fungi	C. albicans, C. tropicalis C. Krusei, Torulopsis glabrata, Trichosporon spp, Geotrichum spp	Butyrate Acetate Propionate↓	<b>†</b>
Diarrhea Virus	Rotavirus, Adenovirus, Norovirus, Anellovirus Calcivirus, Astrovirus, Picobirnavirus, Enterovirus, Dependovirus, Sapovirus, Bufavirus, Bocavirus	Butyrate Acetate Propionate ↓	<b>↑</b>
Abdominal Pain	Methane–Producing Archaea: <i>M. smithii</i> Fungi, Aspergillus spp.	Methane ↑	<b>↑</b>
Bloating Bacteria	Proteobacteria, Faecalibacterium Actinobacteria, Bacteroides Uniformis Bifidobacterium adolescentis Methane Producing Archaea: M. smithii	Methane ↑	↑ ↓ ↑

Figure.1 Chronic diseases implicated from gut microbiome.

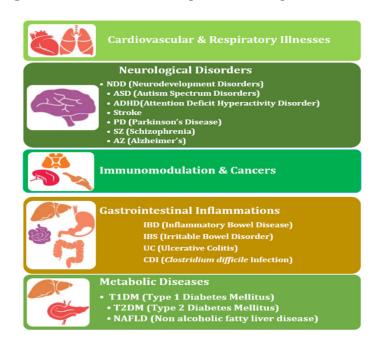


Figure.2 Challenges of Microbiome modulation based therapies.



Figure.3 Mechanism of action of gut microbiome.

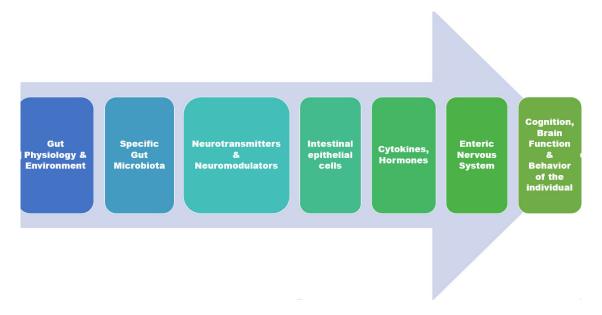


Figure.4 Microbiome modulation-based therapeutic interventions.

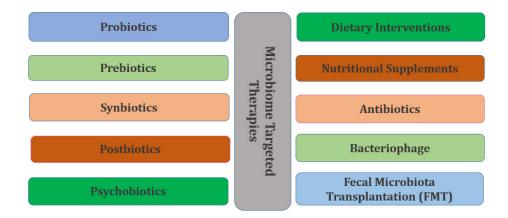


Figure.5 Mechanism of action of gut microbiota associated with CVDs.

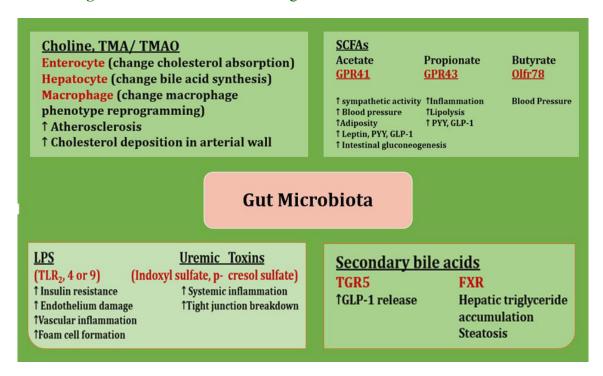
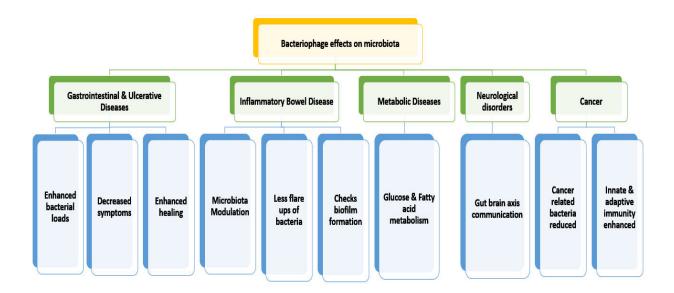


Figure.6 Role of bacteriophage in specific gut microbiota-related diseases.



Increased pro-inflammatory cytokines increase bloodbrain barrier permeability by activating the hypothalamus-pituitary axis thereby leading to decreased serotonin levels and causing depression which can be managed by affecting the hypothalamus pituitary adrenal axis and hence reducing the production of inflammatory

cytokines and enhancing the production of SCFAs, proteins, and neurotransmitters (Sudo *et al.*, 2004; Dowlati *et al.*, 2010; Barbosa *et al.*, 2020). SCFAs help proliferate regulatory T-cells and production of cytokines, maturation, and functioning of microglia (Cowan and Petri, 2018). GABA and glutamate-

producing gut bacteria result in neuronal excitability and enhance synaptic plasticity for improvising cognition. GABA also plays a significant role in preventing neurological diseases, type-1 diabetes, cancer, and immune disorders (Diez-Gutierrez et al., 2020). SCFAs lead to the synthesis of serotonin and the expression of tryptophan hydroxylase-1, a precursor for the synthesis of serotonin. Gut microbiota also influences the expression of BDNF, the lower levels of which are correlated with anxiety and depression (Bistoletti et al., 2019). Psychobiotics also delay the reuptake of neurotransmitters from the synaptic cleft, thereby increasing their duration and release in the synaptic cleft. These also reduce hs-CRP, increase GSH, reduce triglycerides, and insulin levels, and increase antioxidant capacity (Tamtaji et al., 2019). Increased levels of peripheral tumor necrosis factor  $\alpha$  (TNF- $\alpha$ ) have been correlated with enhanced decline in cognition and infections in Alzheimer's disease. Neuro-inflammations are also linked with high levels of TNF-α, transforming growth factor beta (TGF-β) in cerebrospinal fluid (CSF), and increased activation of microglia in dementia (Heneka and Kummer, 2014; Marogianni et al., 2020).

Administration of *Lactobacillus plantarum* has been found to prevent gliosis and subsequently improve cognitive behavior in animal models of AD with reduced neuro-inflammations (Huang *et al.*, 2021). Probiotics containing *Bifidobacterium bifidum*, *Lactobacillus acidophilus*, *L. fermentum*, *and L. reuteri* are found to be associated with decreased TNF-α, IL-1, IL-8, and increased levels of TGF-β, PPAR-γ gene expression which shows anti-inflammatory properties (Borzabadi *et al.*, 2018).

Psychobiotics can be obtained from functional foods containing those probiotic species that can produce GABA and induce the production of neuro-hormones and neurotransmitters. *Bacillus spp.* and *Enterobacter xiangfangensis* are known as GABA-producing bacteria (Luang *et al.*, 2020). *Acetobacter aceti, Acetobacter sp, L. fructivorans, L. fermentum, Leuconostoc spp., Enterococcus faecium, L. kefiranofaciens, Candida krusei,* and *Candida format* found in Kefir grains have potential to improve memory and language functions in AZ patients (Ton *et al.*, 2020). Similarly, fermented milk contains *L. helveticus* to improve cognition in aged adults (Chung *et al.*, 2014).

GABA-producing *L. brevis* is known to have antidepressant action without side effects (Ko *et al.*,

2013). Foods prepared after the fermentation of Laminiaria japonica and adding L. brevis provide protection against dementia (Reid et al., 2018). Similarly, anti-stress and anti-fatigue effects of rice bran fermented with Saccharomyces cerevisiae are seen (Kim et al., 2002). Probiotic yogurt containing B. lactis and L. acidophilus helps to reduce depression, anxiety, and stress (Mohammadi et al., 2016). Bacillus subtilis, Lactobacillus spp., L. rhamnosus can produce nitric oxide (NO), a neurotransmitter involved in regulating gut functions (Gusarov et al., 2013). The peptide hormone GLP-1 produced by gut microbiota acts neuroprotection causing proliferation and apoptosis ofneural cells, lowering Aß plaques, improving memory, stimulating nerve regeneration. and reserving dopaminergic neurons (Kim et al., 2017). Probiotics also improve the metabolism of glucose, reduce neuroinflammation, and check the progression of AD.

Psychobiotics are therefore safe with a low risk of causing an illness. Individuals may show mild symptoms abdominal psychobiotic ofdiscomfort with supplementation till the prevailing microbiota adjusts. Sometimes rashes, itching, and other allergies such as endocarditis, bacteremia, and fungenia may occur as a response to a few strains of bacteria especially in immunocompromised persons but these allergies will go away once psychobiotic intake is discontinued (Sotoudegan et al., 2019). Psychobiotics have been used in dairy, fermented products, and soybean products to be included in the diet for significant mental health (Barbosa, 2020).

Therefore, psychobiotics show their probiotic potential by reducing levels of pro-inflammatory and enhancing anti-inflammatory effects shifting the gut microbiome composition towards more regulated inflammatory pathways involving modulation of the immune system through the production of neuroactive compounds and influencing gut barrier functions (Mohammadi *et al.*, 2019). Research on psychobiotics must be carried out further to see their effectiveness, dosage, safety, long-term effects, and possible side effects while ensuring high standards for the quality and purity of their products so that these form a valuable tool for the treatment of neurological problems.

## **Bacteriophage**

These are the viruses that infect bacteria and serve as key drivers of the composition and functioning of the bacterial community thus playing a vital role in gastrointestinal and cardio-metabolic diseases (CMDs) such as IBD, IBS, diabetes type 1 (T1DM), diabetes type 2 (T2DM) and Clostridium difficile infection (CDI) by preventing the formation of biofilms (Liu et al., 2022; Norman et al., 2015; de Jonge et al., 2022; Yang et al., 2021; Zhao et al., 2017). (Fig.6.) Phages are therefore considered the most potent therapeutic intervention, these biological agents being active for a longer duration than antibiotics after administration and having a narrow target host range with minimum side effects on humans as they replicate within the target bacterium without mutilating mammalian cells. These possess a higher degree of specificity as selectively target the pathogens making their action more effective by producing better. specific, and safe bacteriolytic agents as compared to antibiotics (Kim et al., 2019).

The role of phages is mediated through phage therapy and fecal virome transplantation (FVT). Bacteriophage attaches to mucin, checks the attachment of bacteria and its colonization to the mucin layer, and prevents the death of epithelial cells. The E. coli bacteriophage is known to contribute to amyloid secretion which further induces islet amyloid polypeptide (IAPP) to cause breakdown of beta cells and production of beta antigens that play a significant role in type 1 diabetes (T1DM). IAPP also acts as an autoantigen to develop either autoimmunity or T1DM (Tetz et al., 2019). Besides this, prophages produce various neurotoxins and endotoxins for causing botulism, cholera, diphtheria, shigellosis, and scarlet fever (Waldor and Mekalanos, 1996, Jun et al., 2013; Parajuli et al., 2017; Muthuirulandi et al., 2019; Sakaguchi, 2005). Phage administration leads to an increased abundance of Eubacterium spp and a decrease in Clostridium perfringens (Shkoporov and Hill, 2019). Phage administration leads to reduced levels of Ruminococcus gnavus and C. sporogenes with decreased synthesis of tryptamine. Likewise, phage VD13 results in a decreased number of E. faecalis and hence decreased tyramine and cytolysin production (Dorrestein et al., 2014; Wahida et al., 2021). Phages alter the bacterial function of metabolism and absorption of bile salts from the human gut (Hsu et al., 2019). Phages activate B and T cells to produce antibodies and cytokines by the gut bacteria (Gogokhia et al., 2019).

Phage therapy is successful in urinary tract infection (UTI), gastrointestinal diseases and antibiotic-resistant infections (Duan *et al.*, 2021). While FVT emerges as a therapy for managing obesity and T2DM (Rasmussen *et* 

al., 2020; Manrique et al., 2021; Borin et al., 2023). FVT involves the transfer of viral components from the stools of a healthy person to the recipient who has dysbiosis to restore its microbiome (Biazzo et al., 2022).

Making the transfer free from unwanted viruses and bacteria poses the biggest challenge in phage therapy. The potential role of the bacterium Faecalibacterium prausnitzii has been indicated in the pathophysiology of IBD (Ma et al., 2018). Schizophrenia (SZ) patients include Lactobacillus phage phi-adh more predominantly (Yolken et al., 2015). A high abundance of Acanthocystis turfaea chlorella virus-1 (ATCV-1) is correlated with decreased cognitive functioning due to altered gene expression (Yolken et al., 2014). Phage therapy is in use against E. coli. Streptococcus, and Klebsiella pneumoniae infections, and combinations of phages are implicated in targeting E. coli for gastrointestinal infections (Febvre et al., 2019). Engineered phages are delivered to the gut to modify the composition of disease-associated bacteria by promoting the growth of SCFAs producing bacteria to restore and establish the lost dysbiosis. Therefore, phages can be exploited for diagnosis and prognosis of CMDs while leaving the beneficial bacteria unaltered and also are capable of acting synergistically with antibiotics making them more effective. Possibilities of FVT-based therapeutic studies find extended and promising use in endometriosis (Kitaya and Yasuo, 2023), rheumatoid arthritis (Koziel and Potempa, 2022), and periodontitis where the vagina and oral microbiome are also involved. Therefore, the concept of manipulation of the gut microbiome holds the potential for management of diseases.

## **Fecal Microbiota Transplantation (FMT)**

Among various interventions to restore the diversity and composition of the microbiota, fecal microbiota transplantation (FMT) aims at the transfer of fecal microbial content from a healthy individual to the intestine of a diseased person. FMT has been successfully effective in recurrent *Clostridium difficile* infections (CDI). Treatment success of CDI is found to rely upon bacteriophage transfer during FMT (Zuo *et al.*, 2018). This technique is suggested to treat diarrhea in HIV patients due to *Clostridium difficile* infection. (Elopre and Rodriguez, 2013). FMT in humans forms the basis of treatment of several diseased conditions elucidating that gut microbiota could be the cause of disease rather than a consequence of it (Smits *et al.*, 2013; Aroniadis and Brandt, 2013). Safety concerns are

important, especially in advanced gastrointestinal tract complications that may occur post-FMT (Wortelboer *et al.*, 2019; Ooijevaar *et al.*, 2019; DeFilipp and Bloom, 2019). FMT can be exploited for the management of IBD, IBS, and hepatic encephalopathy.

In IBS patients, the SCFA-producing species such as *Bifidobacterium* and *Ruminococcus sp* are enriched after FMT while *Akkermansia muciniphila* reduces at 12 months period after FMT (Parada *et al.*, 2019; Cruz *et al.*, 2019). FMT infusions from lean donors result in an increased population of *Eubacterium hallii* and *Roseburia intestinalis* with enhanced insulin sensitivity (Duncan *et al.*, 2007) as an association is observed between *Roseburia* and glucose homeostasis in addition to the cardiometabolic role of FMT.

Antimicrobial drugs fail to provide a long-term solution for the management of diseases and chronic conditions. The treatment efficiency is increased only through the engraftment of beneficial species through FMT (Keshteli *et al.*, 2017). It plays a role as a new treatment strategy for malignant tumors, metabolic syndromes, nervous system diseases, and autoimmune disorders.

A non-invasive method of implementing FMT includes the use of FMT-freeze-drying capsules or frozen capsules which are replacing the invasive methods where fresh fecal matter is administered through the distal part of the gut under strictly anaerobic conditions to acquire eubiosis of the gut (Nimgampalle and Kuna, 2017; Tamtaji *et al.*, 2019). Overall success and therapeutic effect of FMT rely upon the match between the donor and the recipient. A large number of small molecules produced by microorganisms, significant SCFAs such as butyrate, production of anti-inflammatory substances lead to strengthening the gut barrier functions leading to the effective success of FMT microbiota inoculated (Zheng *et al.*, 2022).

Therefore gut microbiota is resilient and can be reshaped and reconstructed. Microbiome-targeted therapies aim to rehabilitate disturbed microbiomes to ensure restoring dysbiosis, preventing and curing diseases, and attempting to develop precise therapeutics and treatments. Under normal physiological conditions, gut microbiota helps in the digestion of food, checks the entry of invading pathogens, produces a variety of metabolites good for human health, and strengthens the immune system. Gut microbiota can serve as a key component, the composition of which is influenced by dietary

interventions, starvation or fasting, nutritional supplementation, antibiotics, probiotics, prebiotics, postbiotics, synbiotics, psychobiotics, bacteriophages, and FMT. The introduction of beneficial microbiota forms the platform to develop alternate strategies to preview and cure several chronic ailments, especially autoimmune disorders.

Diseased conditions have been successfully managed and cured in different animal models by manipulating gut microbiota, still, data from such human interventions is less conclusive as the ability of introduced microbes for successful colonization in the gut depends upon the already prevailing microbiota. So, animal model-based clinical trials need to be clinically brought into practice to explain the discrepancies between success and failure in human trials. Thus, a comprehensive analysis of human gut microbiomesbased on their genetics and metabolic predispositions needs further exploration of how various microbiome modulation-based interventions can result in effective prognosis, prevention, and treatment to monitor their progress in combating the comorbidities and to anticipate future complications.

## **Future Prospectives**

The mechanism of the factors that lead to neurodegenerative diseases, immunomodulation and certain types of cancers, gastrointestinal inflammations, metabolic disorders, cardiovascular, and respiratory diseases, and how these are associated with gut microbiome profiles needs to be unveiled. Exhaustive research is required to create individual microbiota profiles and bring them into practice to know the status of gut health, and their correlation with diseases and to decipher the possibilities of the development of diseases and attempting to treat these diseases by manipulating gut microbiota towards the homeostasis. Eventually, this knowledge must form the platform for improving human health by bringing it into clinical practice. Important therapeutic strategies for gut modulation included in this review are dietary interventions, FMT, phage therapy, use of prebiotics, probiotics, postbiotics, synbiotics, and psychobiotics. FMT needs to be made more effective by working out the effective dosage, its transplantation method, and adopting different pre-treatments of frozen preparations. Phage therapy though has elucidated work on metabolic disorders, further research is required to standardize the technique to bring it to medical practice. The potential of psychobiotics to interact with gut microbiota modulation towards a healthy one needs to be

accurately and more precisely uncovered opening the avenues of psychobiotics in adjunct therapies for improving the quality of life for the entire mankind.

Funding: None

#### **Author Contributions**

Neena K. Dhiman: Investigation, formal analysis, writing—original draft.

## **Data Availability**

The datasets generated during and/or analyzed during the current study are available from the corresponding author on reasonable request.

#### **Declarations**

**Ethical Approval** Not applicable.

**Consent to Participate** Not applicable.

Consent to Publish Not applicable.

Conflict of Interest The authors declare no competing interests.

#### References

- Abu-Shanab and E.M.M. Quigley, The role of the gut microbiota in nonalcoholic fatty liver disease, *Nat Rev Gastroenterol Hepatol.*7, 2010, 691–701.
- Adak, A., Khan, M.R. (2019). An insight into gut microbiota and its functionalities. *Cell Mol Life Sci.* 76:473–93.
- Adams, J.B., Johansen, L.J., Powell, L.D., Quig, D., Rubin, R.A. (2011).Gastrointestinal flora and gastrointestinal status in children with autism–comparisons to typical children and correlation with autism severity. *BMC Gastroenterol*. 11: 22.
- Aït-Aissa, A., Aïder, M. (2014). Lactulose: Production and use in functional food, medical and pharmaceutical applications. Practical and critical review. *Int. J. Food Sci. Technol.* 49: 1245–1253.
- Amminger, G.P., Schäfer, M.R., Klier, C.M., Slavikm J.M., Holzer, I., Holub, M. *et al.*, (2012).Decreased nervonic acid levels in erythrocyte membranes predict psychosis in help-seeking ultra-high-risk individuals. *Mol Psychiatry*. 17:1150–2. https://doi.org/10.1038/mp.2011.167.
- Angoorani, P., Ejtahed, H.S., Hasani-Ranjbar, S., Siadat, S.D., Soroush, A.R., Larijani, B. (2021). Gut microbiota modulation as a possible mediating mechanism for

- fasting-induced alleviation of metabolic complications: A systematic review, *Nutr. Metab.* 18: 105.
- Aoyama, M., Kotani, J., Usami, M. (2010). Butyrate and propionate induced activated or non-activated neutrophil apoptosis via HDAC inhibitor activity but without activating GPR-41/GPR-43 pathways. *Nutrition*. 26:653-661.
- Aroniadis, O.C., Brandt, L.J. (2013). Fecal microbiota transplantation: past, present and future. *Curr. Opin. Gastroenterol.* 29: 79–84, http://dx.doi.org/10.1097/MOG.0b013e32835a4b3e.
- Atarashi, K., Tanoue, T., Oshima, K., Suda, W., Nagano, Y., Nishikawa, H., Fukuda, S., Saito, T., Narushima, S., Hase, K. *et al.*, (2013). Treg induction by a rationally selected mixture of *Clostridia* strains from the human microbiota. *Nature*. 500:232–236. https://doi.org/10.1038/nature12331.
- Atarashi, K., Tanoue, T., Shima, T., Imaoka, A., Kuwahara, T., Momose, Y. *et al.*, (2011). Induction of colonic regulatory T cells by indigenous *Clostridium* species. *Science*. 331 (6015):337–41.
- Baek, D., Park, Y. (2013). Association between erythrocyte n-3 polyunsaturated fatty acids and biomarkers of inflammation and oxidative stress in patients with and without depression. *Prostaglandins Leukot Essent Fat Acids*. 89: 8. https://doi.org/10.1016/j.plefa.2013.09.008
- Bagheri, S., Heydari, A., Alinaghipour, A., Salami, M. (2019). Effect of probiotic supplementation on seizure activity and cognitive performance in pTZ-induced chemical kindling. *Epilepsy Behav*. 95:38. <a href="https://doi.org/10.1016/j.yebeh.2019.03.038">https://doi.org/10.1016/j.yebeh.2019.03.038</a>.
- Bailón, E., Cueto-Sola, M., Utrilla, P. *et al.*, (2010). Butyrate in vitro immune-modulatory effects might be mediated through a proliferation-related induction of apoptosis. *Immunobiology*. 215: 863-873.
- Barbara, G. (2006). Mucosal Barrier Defects in Irritable Bowel Syndrome. Who Left the Door Open? *Am. J. Gastroenterol.* 101:1295–1298.
- Barcenilla, A., Pryde, S.E., Martin, J.C., Duncan, S.H., Stewart, C.S., Henderson, C., Flint, H.J. (2000). Phylogenetic Relationships of Butyrate-Producing Bacteria from the Human Gut. *Appl. Environ. Microbiol.* 66: 1654–1661.
- Barrea Webster, A., Staley, C., Hamilton, M.J., Huang, M., Fryxell, K., Erickson, R. *et al.*, (2019). Influence of short-term changes in dietary sulfur on the relative abundances of intestinal sulfate-reducing bacteria. *Gut Microb*.10 (4):447–57.
- Barrea, L., Annunziata, G., Muscogiuri, G., Laudisio, D., Di Somma, C., Maisto, M. *et al.*, (2019). Trimethylamine Noxide, Mediterranean diet, and nutrition in healthy, normal-weight adults: also a matter of sex? *Nutr Burbank Los Angel Cty Calif.* 62:7–17.
- Biazzo, M., and Deidda, G. (2022). Fecal Microbiota Transplantation as New Therapeutic Avenue for Human Diseases. *J. Clin. Med.* 11, 4119.

## https://doi.org/10.3390/JCM11144119.

- Bikel, S., Lo' pez-Leal, G., Cornejo-Granados, F., Gallardo-Becerra, L., Garci'a-Lo' pez, R., Sa' nchez, F., Equihua-Medina, E., Ochoa-Romo, J.P., Lo' pez-Contreras, B.E., Canizales-Quinteros, S., *et al.*, (2021). Gut dsDNA virome shows diversity and richness alterations associated with childhood obesity and metabolic syndrome. *iScience*. 24:102900. https://doi.org/10.1016/J.ISCI.2021.102900.
- Bistoletti, M., Caputi, V., Baranzini, N., Marchesi, N., Filpa, V., Marsilio, I., Cerantola, S., Terova, G., Baj, A., Grimaldi, A., *et al.*, (2019). Antibiotic treatment-induced dysbiosis differently affects BDNF and TrkB expression in the brain and in the gut of juvenile mice. PLoS ONE. 14: e0212856.
- Blanc, P., Daures, J.P., Rouillon, J.M., Peray, P., Pierrugues, R., Larrey, D., Gremy, F., Michel, H. (1992). Lactitol or lactulose in the treatment of chronic hepatic encephalopathy: Results of a meta-analysis. *Hepatology*.15: 222–228.
- Block, J.P., Bailey, L.C., Gillman, M.W., Lunsford, D., Daley, M.F., Eneli, I., *et al.*, (2018). Early Antibiotic Exposure and Weight Outcomes in Young Children. *Pediatrics*. 142:e20180290. <a href="https://doi.org/10.1542/peds.2018-0290">https://doi.org/10.1542/peds.2018-0290</a> PMID: 30381474
- Bonder, M.J., Tigchelaar, E.F., Cai, X., Trynka, G., Cenit, M.C., Hrdlickova, B. *et al.*, (2016). The influence of a short-term gluten-free diet on the human gut microbiome. *GenomeMed*. 8(1):45.
- Borin, J.M., Liu, R., Wang, Y., Wu, T. C., Chopyk, J., Huang, L., Kuo, P.,Ghose, C., Meyer, J.R., Tu, X.M. *et al.*, (2023). Fecal virome transplantation is sufficient to alter fecal microbiota and drive lean and obese body phenotypes in mice. Preprint at bioRxiv. https://doi.org/10.1101/2023.02.03.527064.
- Borzabadi, S., Oryan, S., Eidi, A., Aghadavod, E., Daneshvar Kakhaki, R., Tamtaji, O.R., Taghizadeh, M., Asemi, Z. (2018). The Effects of Probiotic Supplementation on Gene Expression Related to Inflammation, Insulin and Lipid in Patients with Parkinson's Disease: A Randomized, Double-blind, Placebo Controlled Trial. *Arch. Iran. Med.* 21: 289–295.
- Bosi, P., Sarli, G., Casini, L., De Filippi, S., Trevisi, P., Mazzoni, M., Merialdi, G. (2007). The influence of fat protection of calcium formate on growth and intestinal defence in *Escherichia coli* K88-challenged weanling pigs. *Anim Feed Sci Technol*. 139 (3–4):170–185. <a href="https://doi.org/10.1016/j.anifeedsci.2006.12.006">https://doi.org/10.1016/j.anifeedsci.2006.12.006</a>.
- Bough, K.J., Rho, J.M.(2007). Anticonvulsant Mechanisms of the Ketogenic Diet. *Epilepsia*, 48: 43–58.
- Boursier, J., Mueller, O., Barret, M., Machado, M., Fizanne, L., Araujo-Perez, F. *et al.*, (2016). The severity of nonalcoholic fatty liver disease is associated with gut dysbiosis and shift in the metabolic function of the gut microbiota. *Hepatology*. 63 (3):764–75.
- Bowling, A., Davison, K., Haneuse, S., Beardslee, W., Miller,

- D.P. (2017). ADHD medication, dietary patterns, physical activity, and bmi in children: a longitudinal analysis of the ECLS-K Study. *Obesity*. 25:21949. https://doi.org/10.1002/oby.21949.
- Brandl, K., Plitas, G., Mihu, C.N., Ubeda, C., Jia, T., Fleisher, M. *et al.*, (2008).Vancomycin-resistant *enterococci* exploit antibiotic-induced innate immune deficits. *Nature*. 455:804–807. <a href="https://doi.org/10.1038/nature07250">https://doi.org/10.1038/nature07250</a> PMID: 18724361
- Buffie, C. G., Jarchum, I., Equinda, M., Lipuma, L., Gobourne, A., Viale, A., Ubeda, C., Xavier, J., & Pamer, E. G. (2012). Profound alterations of intestinal microbiota following a single dose of clindamycin results in sustained susceptibility to Clostridium difficile-induced colitis. *Infection and Immunity*. 80 (1), 62–73. https://doi.org/10.1128/IAI.05496-11.
- Buffie, C.G., Pamer, E.G. (2013). Microbiota-mediated colonization resistance against intestinal pathogens. *Nat Rev Immunol*. 13:790–801. https://doi.org/10.1038/nri3535.
- Burger, M., Hoosain, M., Einspieler, C., Unger, M., Niehaus *et al.*, (2020).Maternal perinatal mental health and infant and toddler neurodevelopment evidence from low and middle-income countries. A systematic review. *J Affect Disord*. 268:23. https://doi.org/10.1016/j.jad.2020.03.023.
- Cadenhead, K., Addington, J., Bearden, C., Cannon, T., Cornblatt, B., Mathalon, D. *et al.*, (2017). Dietary omega 3 and erythrocyte omega 3 are associated with symptoms, functioning and psychotic conversion in a clinical high risk population. *Neuropsychopharmacology*.
- Cahenzli, J., Köller, Y., Wyss, M., Geuking, M. B., & McCoy, K. D. (2013). Intestinal microbial diversity during early-life colonization shapes long-term IgE levels. *Cell Host and Microbe*. 14(5): 559–570. https://doi.org/10.1016/j.chom.2013.10.004.
- Camargo, A.C., Todorov, S.D., Chihib, N.E., Drider, D., Nero, L.A. (2018). Lactic Acid Bacteria (LAB) and Their Bacteriocins as Alternative Biotechnological Tools to Control *Listeria monocytogenes* Biofilms in Food Processing Facilities. *Mol. Biotechnol.*, 60: 712–726.
- Candela, M.; Guidotti, M.; Fabbri, A.; Brigidi, P.; Franceschi, C.; Fiorentini, C. Human intestinal microbiota:Cross-talk with the host and its potential role in colorectal cancer. Crit. Rev. Microbiol. 2011, 37, 1–14.
- Cani, P.D., Jordan, B.F. (2018). Gut microbiota-mediated inflammation in obesity: A link with gastrointestinal cancer. *Nat. Rev. Gastroenterol. Hepatol.* 15: 671–682.
- Carrothers, J.M., York, M.A, Brooker, S.L., Lackey, K.A., Williams, J.E., Shafii, B. *et al.*, (2015). Fecal microbial community structure is stable over time and related to variation in macronutrient and micronutrient intakes in lactating women. *J Nutr*.145:2379–88. <a href="https://doi.org/10.3945/jn.115.211110">https://doi.org/10.3945/jn.115.211110</a>.
- Castro, K., Faccioli, L.S., Baronio, D., Gottfried, C., Perry, I.S., dos Santos Riesgo, R. (2015). Effect of a ketogenic

- diet on autism spectrum disorder: a systematic review. *Res Autism Spectr Disord*. 20: 31–8. https://doi.org/10.1016/j.rasd.2015.0 8.005.
- Caussy, C., Tripathi, A., Humphrey, G., Bassirian, S., Singh, S., Faulkner, C., Bettencourt, R., Rizo, E., Richards, L., Xu, Z.Z., *et al.*, (2019). A gut microbiome signature for cirrhosis due to nonalcoholic fatty liver disease. *Nat. Commun.* 10:1406. <a href="https://doi.org/10.1038/s41467-019-09455-9">https://doi.org/10.1038/s41467-019-09455-9</a>.
- Cazorla, S. I., Maldonado-Galdeano, C., Weill, R., De Paula, J., & Perdigón, G. (2018). 'Oral administration of probiotics increases Paneth cells and intestinal antimicrobial activity'. *Frontiers in Microbiology*. 9: 1–14. https://doi.org/10.3389/fmicb.2018.00736.
- Cha, H.Y., Yang, S.J. (2020).Anti-inflammatory diets and schizophrenia. *Clin Nutr Res.* 9:241. https://doi.org/10.7762/cnr.2020.9.4.241.
- Chaiyasit, K., Wiwanitkit, V. (2016). Black pepper: stimulation of diarrhea in patient with underlying short bowel syndrome. *Ancient Sci Life*. 35 (3):185. https://doi.org/10.4103/0257-7941.179872.
- Chambers, E.S., Preston, T., Frost, G., Morrison, D.J. (2018) Role of Gut Microbiota-Generated Short-Chain Fatty Acids in Metabolic and Cardiovascular Health. *Curr. Nutr. Rep.* 7:198–206.
- Chassaing, B., Koren, O., Goodrich, J.K., Poole, A.C., Srinivasan, S., Ley, R.E. *et al.*, (2015). Dietary emulsifiers impact the mouse gut microbiota promoting colitis and metabolic syndrome. *Nature*. 519 (7541):92–6.
- Chassaing, B., Van de Wiele, T., De Bodt, J., Marzorati, M., Gewirtz, A.T. (2017). Dietary emulsifiers directly alter human microbiota composition and gene expression ex vivo potentiating intestinal inflammation. *Gut.* 66 (8):1414–27.
- Chatterjee, I., Lu, R., Zhang, Y., Zhang, J., Dai, Y., Xia, Y. *et al.*, (2020). Vitamin D receptor promotes healthy microbial metabolites and microbiome. Sci Rep.10:7340. https://doi.org/10.1038/s41598-020-64226-7.
- Chen, J., Yu, B., Chen, D., Zheng, P., Luo, Y., Huang, Z., Luo, J., Mao, X., Yu, J., He, J. (2019). Changes of porcine gut microbiota in response to dietary chlorogenic acid supplementation. *Appl. Microbiol. Biotechnol.* 103: 8157–8168.
- Chen, M.L., Sundrud, M.S. (2016). Cytokine Networks and T-Cell Subsets in Inflammatory Bowel Diseases. *Inflamm. Bowel Dis.* 22: 1157–1167.
- Chen, Q., Ma, X., Li, C., Shen, Y., Zhu, W., Zhang, Y., Guo, X., Zhou, J., and Liu, C. (2020). Enteric Phageome Alterations in Patients With Type 2 Diabetes. *Front. Cell. Infect. Microbiol.* 10:575084. <a href="https://doi.org/10.3389/FCIMB.2020.575084">https://doi.org/10.3389/FCIMB.2020.575084</a>.
- Chen, R.Y., Mostafa, I., Hibberd, M.C., Das, S., Mahfuz, M., Naila, N.N, et.al.(2021). Microbiota-Directed Food Intervention for Undernourished Children. *N Engl J Med*. 384:1517–1528.

- https://doi.org/10.1056/NEJMoa2023294 PMID: 33826814
- Cheng, L.H., Liu, Y.W., Wu, C.C., Wang, S., Tsai, Y.C. (2019). Psychobiotics in mental health, neurodegenerative and neurodevelopmental disorders. *J. Food Drug Anal.* 27: 632–648.
- Chung, Y-C., Jin, H-M., Cui, Y. *et al.*, (2014). Fermented milk of *Lactobacillus helveticus* IDCC3801 improves cognitive functioning during cognitive fatigue tests in healthy older adults. *J. Funct. Foods.* 10:465–474.
- Clarke, S., Green-Johnson, J., Brooks, S., Ramdath, D., Bercik, P., Avila, C., Inglis, G., Green, J., Yanke, L., Selinger, L. (2016). B2-1 fructan supplementation alters host immune responses in a manner consistent with increased exposure to microbial components: Results from a double-blinded, randomised, cross-over study in healthy adults. *Br. J. Nutr.* 115: 1748–1759.
- Cohen, J (2016). Vaginal microbiome affects HIV risk. *Science*. 353(6297):331.
- Coker, M.O., Laue, H.E., Hoen, A.G., Hilliard, M., Dade, E., Li, Z. *et al.*, (2021).Infant feeding alters the longitudinal impact of birth mode on the development of the gut microbiota in the first year of life. *Front Microbiol*. 12197. https://doi.org/10.3389/fmicb.2021.642197
- Collado, M.C., Salminen, S., Vinderola, G. (2021). Chapter 11-Postbiotics: Defining the impact of inactivated microbes and their metabolites on promotion of health. In: The Human Microbiome in Early Life. (Koren, O., Rautava, S., Eds.), Academic Press: Cambridge, MA, USA. pp. 257–268.
- Corsello, G., Carta, M., Marinello, R., Picca, M., De Marco, G., Micillo, M., Ferrara, D., Vigneri, P., Cecere, G., Ferri, P., *et al.*, (2017). Preventive Effect of Cow's Milk Fermented with *Lactobacillus paracasei* CBA L74 on Common Infectious Diseases in Children: A Multicenter Randomized Controlled Trial. *Nutrients* 9:669. <a href="https://doi.org/10.3390/nu9070669">https://doi.org/10.3390/nu9070669</a>.
- Costabile, A., Bergillos-Meca, T., Rasinkangas, P. *et al.*, (2017). Effects of soluble corn fiber alone or in synbiotic combination with *Lactobacillus rhamnosus* GG and the pilus-deficient derivative GG-PB12 on fecal microbiota, metabolism, and markers of immune function: a randomized, double-blind, placebo-controlled, crossover study in healthy elderly (saimes study). *Front Immunol*. 8:1443. https://doi.org/10.3389/fimmu.2017.01443
- Cowan, C.S.M, Stylianakis, A.A., Richardson, R. (2019).Early-life stress. microbiota, brain development: probiotics reverse the effects of maternal separation on neural circuits underpinning fear expression and extinction in infant rats. Dev Cogn Neurosci. 37:627. https://doi.org/10.1016/j.dcn.2019.100627.
- Cowan, M., Petri, W.A. Jr. (2018). Microglia: Immune Regulators of Neurodevelopment. *Front. Immunol.* 9: 2576.
- Cox, M.A., Jackson, J., Stanton, M., et al., (2009). Short-chain

- fatty acids act as anti-inflammatory mediators by regulating prostaglandin E and cytokines. *World J.Gastroentero* 15: 5549-5557.
- Cross, H.S., Bises, G., Lechner, D., Manhardt, T., Kállay, E. (2005). The Vitamin D endocrine system of the gut—Its possible role in colorectal cancer prevention. *J SteroidBiochem Mole Biol.* 97:121–8. https://doi.org/10.1016/j.jsbmb.2005.06.005.
- Cruz-Aguliar, R.M., Wantia, N., Clavel, T., Vehreschild, M.J.G.T., Buch, T., Bajbouj, M., Haller, D., Busch, D., Schmid, R., Stein-Thoeringer, C. *et al.*, (2019). An openlabeled study on fecal microbiota transfer in irritable bowel syndrome patients reveals improvement in abdominal pain associated with the relative abundance of *akkermansia muciniphila*. *Digestion*. 100:127–138. https://doi.org/10.1159/000494252.
- Cryan, J.F., Dinan, T.G. (2012).Mind-altering microorganisms: the impact of the gut microbiota on brain and behavior. *Nat Rev Neurosci*. 13:701–12. https://doi.org/10.1038/nrn3346.
- Dapito, D.H., Mencin, A., Gwak, G.Y., Pradere, J.P., Jang, M.K., Mederacke, I. *et al.*, (2012). Promotion of hepatocellular carcinoma by the intestinal microbiota and TLR4. *Cancer Cell*. 21(4):504–16.
- Dargahi, N., Matsoukas, J., Apostolopoulos, V. (2020). Streptococcus thermophiles ST285 alters proinflammatory to anti-inflammatory cytokine secretion against multiple sclerosis peptide in mice. Brain Sci. 10:126.
- Dasgupta, S., Erturk-Hasdemir, D., Ochoa-Reparaz, J., Reinecker, H.C. and Kasper, D.L. (2014). Plasmacytoid dendritic cells mediate anti-inflammatory responses to a gut commensal molecule via both innate and adaptive mechanisms. *Cell Host Microbe*. 15:413–423. https://doi.org/10.1016/j.chom.2014.03.006.
- David, L.A., Maurice, C.F., Carmody, R.N., Gootenberg, D.B., Button, J.E., Wolfe, B.E. *et al.*, (2014).Diet rapidly and reproducibly alters the human gut microbiome. *Nature*. 505 (7484):559–63.
- de Jonge, P.A., Wortelboer, K., Scheithauer, T.P.M., van den Born, B.J.H., Zwinderman, A.H., Nobrega, F.L., Dutilh, B.E., Nieuwdorp, M., and Herrema, H. (2022). Gut virome profiling identifies a widespread bacteriophage family associated with metabolic syndrome. *Nat. Commun.* 13:3594. <a href="https://doi.org/10.1038/S41467-022-31390-5">https://doi.org/10.1038/S41467-022-31390-5</a>.
- De Moraes, W.M.A.M., Mendes, A.E.P., Lopes, M.M.M., Maia, F.M.M.(2017). Protein overfeeding is associated with improved lipid and anthropometric profile thus lower malondialdehyde levels in resistance-trained athletes. *Int. J. Sports Sci.* 7: 87–93.
- de Moura e Dias, M., Pais Siqueira, N., Lopes da Conceiç~ao, L., Aparecida dos Reis, S., Xavier Valente, F., Maciel dos Santos Dias, M. *et al.*, (2018). Consumption of virgin coconut oil in Wistar rats increases saturated fatty acids in the liver and adipose tissue, as well as adipose tissue

- inflammation. J Funct Foods. 48: 472–80.
- DeFilipp, Z., Bloom, P.P., Torres Soto. M. *et al.*, (2019). Drugresistant *E. coli* bacteremia transmitted by fecal microbiota transplant. *N Engl J Med.* 381(21):2043-2050.
- Del Toro-Barbosa, M., Hurtado-Romero, A.; Garcia-Amezquita, L.E.; García-Cayuela, T. Psychobiotics: Mechanisms of Action, Evaluation Methods and Effectiveness in Applications with Food Products. Nutrients 2020, 12, 3896.
- Desbonnet, L., Clarke, G., Shanahan, F., Dinan, T.G., Cryan, J.F. (2014). Microbiota is essential for social development in the mouse. *Mol Psychiatry*. 19:146–8. https://doi.org/10.1038/mp.2013.65
- Devkota, S., Wang, Y., Musch, M.W., Leone, V., Fehlner-Peach, H., Nadimpalli, A. *et al.*, (2012). Dietary-fat-induced taurocholic acid promotes pathobiont expansion and colitis in II10-/- mice. *Nature*. 487(7405):104–8.
- Diez-Gutierrez, L., San Vicente, L., Barron, L.J.R. et. al. (2020). Gammaaminobutyric acid and probiotics: multiple health benefits and their future in the global functional food and nutraceuticals market. *J. Funct. Foods* 64:103669
- Dinan, T.G., Stanton, C., Cryan, J.F. (2013). Psychobiotics: A novel class of psychotropic. *Biol. Psychiatry*. 74: 720– 726
- Dorrestein, P., Mazmanian, S., Knight, R. (2014). Finding the missing links among metabolites, microbes, and the Host. *Immunity*. 40 (6):824–832. https://doi.org/10.1016/j.immuni.2014.05.015.
- Dowlati, Y., Herrmann, N., Swardfager, W., Liu, H., Sham, L., Reim, E.K., Lanctôt, K.L. (2010). A Meta-Analysis of Cytokines in Major Depression. *Biol. Psychiatry*. 67: 446–457.
- Duan, Y., Young, R., and Schnabl, B. (2021). Bacteriophages and their potential for treatment of gastrointestinal diseases. *Nat. Rev. Gastroenterol. Hepatol.* 19: 135–144. https://doi.org/10.1038/s41575-021-00536-z.
- Dubourg, G., Lagier, J. C., Robert, C., Armougom, F., Hugon, P., Metidji, S., Dione, N., Dangui, N. P. M., Pfleiderer, A., Abrahao, J., Musso, D., Papazian, L., Brouqui, P., Bibi, F., Yasir, M., Vialettes, B., & Raoult, D. (2014). Culturomics and pyrosequencing evidence of the reduction in gut microbiota diversity in patients with broad-spectrum antibiotics. *International Journal of Antimicrobial Agents*. 44(2): 117–124. <a href="http://doi.org/10.1016/j.ijantimicag.2014.04.020">http://doi.org/10.1016/j.ijantimicag.2014.04.020</a>.
- Duncan, S., Belenguer, A., Holtrop, G. et al., (2007). Reduced dietary intake of carbohydrates by obese subjects results in decreased concentrations of butyrate and butyrate producing bacteria in feces. *Appl Environ Microb*.73: 1073-1078.
- Dupuis, N., Curatolo, N., Benoist, J.F., Auvin, S. (2015). Ketogenic diet exhibits anti-inflammatory properties. *Epilepsia*. 56:38. <a href="https://doi.org/10.1111/epi.13038">https://doi.org/10.1111/epi.13038</a>.
- Dürholz, K., Hofmann, J., Iljazovic, A., Häger, J., Lucas, S., Sarter, K. (2020). Dietary short term fiber interventions

- in arthritis patients increase systemic SCFA levels and regulate inflammation. *Nutrients*.12:3207.
- Elopre, L., Rodriguez, M. (2013).Fecal microbiota therapy for recurrent *Clostridiumdifficile* infection in HIV-infected persons, *Ann. Intern. Med.* 158:779–780, <a href="http://dx.doi.org/10.7326/0003-4819-158-10-201305210-00021">http://dx.doi.org/10.7326/0003-4819-158-10-201305210-00021</a>.
- Del Chierico et. al. (2017). Gut microbiota profiling of pediatric NAFLD and obese patients unveiled by an integrated meta-omics based approach, *Hepatology*. 65(2):451-464. https://doi.org/ 10.1002/hep.28572 https://doi.org/10.1002/hep.28572.
- Fan, G., Cao, F., Kuang, T., Yi, H., Zhao, C., Wang, L., Peng, J., Zhuang, Z., Xu, T., Luo, Y., *et al.*, (2023). Alterations in the gut virome are associated with type 2 diabetes and diabetic nephropathy. *Gut Microb*. 15:2226925.
- Febvre, H.P., Rao, S., Gindin, M., Goodwin, N.D.M., Finer, E., Vivanco, J.S., Lu, S., Manter, D.K., Wallace, T.C., and Weir, T.L. (2019). PHAGE Study: Effects of Supplemental Bacteriophage Intake on Inflammation and Gut Microbiota in Healthy Adults. *Nutrients*. 11: 666. https://doi.org/10.3390/NU11030666.
- Fei, N., Zhao, L. (2013). An opportunistic pathogen isolated from the gut of an obese human causes obesity in germfree mice. *IsmeJ.* 7: 880-884.
- Fenn, K., Strandwitz, P., Stewart, E.J., Dimise, E., Rubin, S., Gurubacharya, S. *et al.*, (2017). Quinones are growth factors for the human gut microbiota. *Microbiome*. 5:161. https://doi.org/10.1186/s40168-017-0380-5.
- Fernandez-Real, J.M., Serino, M., Blasco, G., Puig, J., Daunisi-Estadella, J., Ricart, W. *et al.*, (2015).Gut microbiota interacts with brain microstructure and function. *J Clin EndocrinolMetab*. 100: 4505–13. https://doi.org/10.1210/jc.2015-3076.
- Flint, H.J., Duncan, S.H., Scott, K.P., Louis, P. (2014). Links between diet, gut microbiota composition and gut metabolism. *Proc. Nutr. Soc.* 74: 13–22.
- Foster, J. A., McVey Neufeld, K.A. (2013).Gut-brain axis: how the microbiome influences anxiety and depression. *Trends Neurosci.* 36:305–12. https://doi.org/10.1016/j.tins.2013.01.005
- Fraguas, D., Díaz-Caneja, C.M., Pina-Camacho, L., Moreno, C., Durán-Cutilla, M., Ayora, M., *et al.*, (2019).Dietary interventions for autism spectrum disorder: a Meta-analysis. *Pediatrics*. 144:3218. https://doi.org/10.1542/peds.2018-3218.
- Fransen, F., A.A. van Beek, T. Borghuis, B. Meijer, F. Hugenholtz, C. van der Gaast-de Jongh, H.F. Savelkoul, M.I. de Jonge, M.M. Faas, M.V. Boekschoten *et al.*, (2017). The impact of gut microbiota on gender-specific differences in immunity. *Front. Immunol.* 8:754. <a href="https://doi.org/10.3389/fimmu.2017.00754">https://doi.org/10.3389/fimmu.2017.00754</a>
- Gálvez, A., Abriouel, H., López, R.L., Ben Omar, N. (2007). Bacteriocin-based strategies for food biopreservation. *Int. J. Food Microbiol.* 120: 51–70.
- Gaman, A.; Kuo, B. Neuromodulatory processes of the brain-

- gut axis. Neuromodulation 2008, 11, 249-259.
- Gan, X.T., Ettinger, G., Huang, C.X., Burton, J.P., Haist, J.V., Rajapurohitam, V., Sidaway, J.E., Martin, G., Gloor, G.B., Swann, J.R., Reid, G., Karmazyn, M.(2014). Probiotic administration attenuates myocardial hypertrophy and heart failure after myocardial infarction in the rat. *Circ Heart Fail*. 7:491– 499. <a href="https://doi.org/0.1161/CIRCHEARTFAILURE.113.0009">https://doi.org/0.1161/CIRCHEARTFAILURE.113.0009</a> 78.
- Gänzle, M.G. (2015).Lactic metabolism revisited: Metabolism of lactic acid bacteria in food fermentations and food spoilage. *Curr. Opin. Food Sci.* 2:106–117.
- Garcia Diaz, T., Ferriani Branco, A., Jacovaci, F.A., Cabreira Jobim, C., Bolson, D.C., and Pratti Daniel, J.L. (2018). Inclusion of live yeast and mannan-oligosaccharides in high grain-based diets for sheep: Ruminal parameters, inflammatory response and rumen morphology. PLoS One.13: e0193313. https://doi.org/10.1371/journal.pone.0193313.
- Gasbarrini, G., Bonvicini, F. and Gramenzi, A. (2016).
- Probiotics history. J. Clin. Gastroenterol. 50: S116–S119. https://doi.org/10.1097/MCG.000000000000000697.
- Ge, X., Ding, C., Zhao, W., Xu, L., Tian, H., Gong, J., Zhu, M., Li, J., & Li, N. (2017). Antibiotics-induced depletion of mice microbiota induces changes in host serotonin biosynthesis and intestinal motility. *Journal of Translational Medicine*. 15(1): 1–9. <a href="https://doi.org/10.1186/s12967-016-1105-4">https://doi.org/10.1186/s12967-016-1105-4</a>.
- Gevers, D., Kugathasan, S., Denson, L.A., Vázquez-Baeza, Y., Van Treuren, W., Ren, B. *et al.*, (2014). The treatment-naive microbiome in new-onset Crohn's disease, *Cell Host Microbe*. 15:382–392, <a href="http://dx.doi.org/10.1016/j.chom.2014.02.005">http://dx.doi.org/10.1016/j.chom.2014.02.005</a>.
- Gill, P.A., van Zelm, M.C., Muir, J.G. *et al.*, (2018). Review article: short chain fatty acids as potential therapeutic agents in human gastrointestinal and inflammatory disorders. *Aliment Pharmacol Ther*. 48:15–34. https://doi.org/10.1111/apt.14689
- Gillis, C. C., Hughes, E. R., Spiga, L., Winter, M. G., Zhu, W., Furtado de Carvalho, T., Chanin, R. B., Behrendt, C. L., Hooper, L. V., Santos, R. L., & Winter, S. E. (2018). Dysbiosis- associated change in host metabolism generates lactate to support Salmonella growth. *Cell Host and Microbe*. 23(1):54–64. https://doi.org/10.1016/j.chom.2017.11.006.
- Gogokhia, L., Buhrke, K., Bell, R., Hoffman, B., Brown, D.G., Hanke-Gogokhia, C., Ajami, N.J., Wong, M.C., Ghazaryan, A., Valentine, J.F. *et al.*,(2019). Expansion of bacteriophages is linked to aggravated intestinal inflammation and colitis. *Cell Host Microbe*.25(2):285–299.e8. https://doi.org/10.1016/j.chom.2019.01.008.
- Golubeva, A.V., Joyce, S.A., Moloney, G., Burokas, A., Sherwin, E., Arboleya, S. *et al.*, (2017).Microbiotarelated changes in bile acid, tryptophan metabolism are associated with gastrointestinal dysfunction in a mouse model of autism. *EBioMedicine*. 24:20.

- https://doi.org/10.1016/j.ebiom.2017.09.020.
- Gomez-Arango, L.F., Barrett, H.L., McIntyre, H.D., Callaway, L.K., Morrison, M., Dekker Nitert, M. (2016). SPRING Trial Group. Increased systolic and diastolic blood pressure is associated with altered gut microbiota composition and butyrate production in early pregnancy. *Hypertension*. 68:974–981.
  - https://doi.org/10.1161/HYPERTENSIONAHA.116.07910.
- Gong, H.S., Meng, X. C., Wang, H. (2010). Mode of action of plantaricin MG, a bacteriocin active against *Salmonella typhimurium*. 50, S37–S45.
- Gordillo Altamirano, F.L., and Barr, J.J. (2019). Phage Therapy in the Postantibiotic. *Era. Clin. Microbiol. Rev.* 32: e00066-18. https://doi.org/10.1128/CMR.00066-18.
- Gow, R.V. (2013). The Omega-3 Fatty Acid Deficiency Syndrome: Opportunities for Disease Prevention. McNamara RK, editor. Nova Science Publishers, Inc.
- Gracious, B.L., Finucane, L., Friedman-Campbell, M., Messing, S., Parkhurst, M.N. (2012). Vitamin D deficiency and psychotic features in mentally ill adolescents: a cross-sectional study. *BMC Psychiatry*. 12:38. https://doi.org/10.1186/1471-244X-12-38.
- Graf, D., Monk, J.M., Lepp, D., Wu, W., McGillis, L., Roberton, K. *et al.*, (2019).Cooked red lentils dose-dependently modulate the colonic microenvironment in healthy C57Bl/6 male mice. *Nutrients*. 11 (8):1853.
- Grüber, C., van Stuijvenberg, M., Mosca, F., Moro, G., Chirico, G., Braegger, C.P., Riedler, J., Boehm, G.,Wahn, U. (2010). MIPS 1 Working Group. Reduced occurrence of early atopic dermatitis because of immunoactive prebiotics among low-atopy-risk infants. J. Allergy Clin. Immunol. 126:791–797.
- Gu, W., Zhang, L., Han, T., Huang, H., Chen, J. (2022). Dynamic Changes in Gut Microbiome of Ulcerative Colitis: Initial Study from Animal Model. *J. Inflamm. Res.* 15: 2631–2647.
- Guigoz, Y., Rochat, F., Perruisseau-Carrier, G., Rochat, I., Schiffrin, E. (2002). Effects of oligosaccharide on the faecal flora and non-specific immune system in elderly people. *Nutr.Res.* 22: 13–25.
- Gusarov, I., Gautier, L., Smolentseva, O. *et al.*, (2013). Bacterial nitric oxide extends the lifespan of *C. elegans*. *Cell* 152:818–830.
- Sokol, B. Pigneur, L. Watterlot, O. Lakhdari, L.G. Bermudez-Humaran, J.- J. Gratadoux, S. Blugeon, C. Bridonneau, J.-P. Furet, G. Corthier, C. Grangette, N. Vasquez, P. Pochart, G. Trugnan, G. Thomas, H.M. Blottiere, J. Dore, P. Marteau, P. Seksik, P. Langella. (2008). Faecalibacterium prausnitzii is an anti-inflammatory commensal bacterium identified by gut microbiota analysis of Crohn disease patients, PNAS. 105 16731–16736, https://doi.org/10.1073/pnas.0804812105.
- Hamada, H., Haruma, K., Mihara, M., Kamada, T., Yoshihara, M., Sumii, K. *et al.*, (2000). High incidence of reflux oesophagitis after eradication therapy for *Helicobacter*

- *pylori*: impacts of hiatal hernia and corpus gastritis. *Aliment Pharmacol Ther*. 14 (6):729–35.
- Han, M., Yang, P., Zhong, C. and Ning, K. (2018). The Human Gut Virome in Hypertension. *Front. Microbiol.* 9: 3150. https://doi.org/10.3389/FMICB.2018.03150/FULL.
- Haukioja, A., Yli-Knuuttila, H., Loimaranta, V., Kari, K., Ouwehand, A.C., Meurman, J.H., and Tenovuo, J. (2006). Oral adhesion and survival of probiotic and other *lactobacilli* and *bifidobacteria* in vitro. Oral Microbiol. Immunol. 21, 326–332. <a href="https://doi.org/10.1111/j.1399-302X.2006.00299.x">https://doi.org/10.1111/j.1399-302X.2006.00299.x</a>.
- Heiman, M.L., Greenway, F.L. (2016). A healthy gastrointestinal microbiome is dependent on dietary diversity. *Mol. Metab.* 5: 317–320.
- Henao-Mejia, J., Elinav, E., Jin, C., Hao, L., Mehal, W.Z., Strowig, T. *et al.*, (2012). Inflammasome-mediated dysbiosis regulates progression of NAFLD and obesity, *Nature*. 482 :179–185, http://dx.doi.org/10.1038/nature10809.
- Heneka, M.T., Kummer, M.P., Latz, E. (2014). Innate immune activation in neuro-degenerative disease. *Nat. Rev. Immunol.* 14: 463–477.
- Hill, D.B., Henderson, L.M., Mcclain, C.J. (1991). Osmotic diarrhea induced by sugar-free theophylline solution in critically III patients. *J Parenteral Enter Nutr*. 15(3):332–336. https://doi.org/10.1177/0148607191015003332.
- Hsu, B.B., Gibson, T.E., Yeliseyev, V., Liu, Q., Lyon, L., Bry, L., Silver, P.A., Gerber, G.K. (2019). Dynamic modulation of the gut microbiota and metabolome by bacteriophages in a mouse Model. *Cell Host Microbe*. 25 (6):803–814. e5. <a href="https://doi.org/10.1016/j.chom.2019.05.001">https://doi.org/10.1016/j.chom.2019.05.001</a>.
- Huang, H.J., Chen, J.L., Liao, J.F., Chen, Y.H., Chieu, M.W., Ke, Y.Y., Hsu, C.C., Tsai, Y.C., Hsieh-Li, H.M. (2021).
   Lactobacillus plantarum PS128 prevents cognitive dysfunction in Alzheimer's disease mice by modulating propionic acid levels, glycogen synthase kinase 3 beta activity, and gliosis. BMC Complement Med. 21: 259.
- Humphrey, S.P., and Williamson, R.T. (2001). A review of saliva: normal composition, flow, and function. J. Prosthet. Dent 85, 162–169. https://doi.org/10.1067/mpr.2001.113778.
- Huycke, M.M., Abrams, V., Moore, D.R.(2002). *Enterococcus faecalis* produces extracellular superoxide and hydrogen peroxide that damages colonic epithelial cell DNA. Carcinogenesis. 23(3):529–36.
- Iancu, M.A., Profir, M., Roşu, O.A., Ionescu, R.F., Cretoiu, S.M., Gaspar, B.S. (2023). Revisiting the intestinal microbiome and its role in diarrhea and constipation. *Microorganisms*. 11(9):11. https://doi.org/10.3390/microorganisms11092177.
- Imhann, F., M.J. Bonder, A. Vich Vila, J. Fu, Z. Mujagic, L.
  Vork, E.F. Tigchelaar, S.A. Jankipersadsing, M.C. Cenit,
  H.J. Harmsen, *et al.*, (2016). Proton pump inhibitors affect the gut microbiome. *Gut*. 65:740–748.

- https://doi.org/10.1136/gutjnl-2015-310376.
- Izuddin, W.I., Loh, T.C., Samsudin, A.A., Foo, H.L., Humam, A.M., and Shazali, N. (2019). Effects of postbiotic supplementation on growth performance, ruminal fermentation and microbial profile, blood metabolite and GHR, IGF-1 and MCT-1 gene expression in postweaning lambs. BMC Vet. Res. 15, 315. <a href="https://doi.org/10.1186/s12917-019-2064-9">https://doi.org/10.1186/s12917-019-2064-9</a>.
- Jarmołowska, B., Bukało, M., Fiedorowicz, E., Cie'sli'nska, A., Kordulewska, N.K., Moszy'nska, M. et al., (2019).Role of milk-Derived opioid peptides and proline dipeptidyl peptidase-4 in autism spectrum disorders. Nutrients. 11:87. https://doi.org/10.3390/nu11010087.
- Jensen, G.S., Benson, K.F., Carter, S.G., Endres, J.R.(2010). GanedenBC30TM cell wall and metabolites: antiinflammatory and immune modulating effects in vitro, BMC Immunol. 11-15, <a href="https://doi.org/10.1186/1471-2172-11-15">https://doi.org/10.1186/1471-2172-11-15</a>.
- Jensen, G.S., Hart, A.N. and A.G. (2007). An antiinflammatory immunogen from yeast culture induces activation and alters chemokine receptor expression on human natural killer cells and B lymphocytes in vitro. *Nutr. Res.* 27:327–335. https://doi.org/10.1016/j.nutres.2007.04.008.
- Jernberg, C., Löfmark, S., Edlund, C., & Jansson, J. K. (2007). Long-term ecological impacts of antibiotic administration on the human intestinal microbiota. *ISME Journal*. 1(1): 56–66. https://doi.org/10.1038/ismej.2007.3.
- Jernberg, C., Löfmark, S., Edlund, C., & Jansson, J. K. (2010). Long-term impacts of antibiotic exposure on the human intestinal microbiota. *Microbiology*. 156(11): 3216–3223. https://doi.org/10.1099/mic.0.040618-0.
- Jie, Z., Xia, H., Zhong, S.L., Feng, Q., Li, S., Liang, S., Zhong, H., Liu, Z., Gao, Y., Zhao, H., et al., (2017). The gut microbiome in atherosclerotic cardiovascular disease. Nat. Commun. 8: 845. <a href="https://doi.org/10.1038/s41467-017-00900-1">https://doi.org/10.1038/s41467-017-00900-1</a>.
- Jin, X., Zhang, M., and Yang, Y.F. (2019). Saccharomyces cerevisiae b-glucan-induced SBD-1 expression in ovine ruminal epithelial cells is mediated through the TLR-2-MyD88-NF-kB/MAPK pathway. Vet. Res. Commun. 43: 77–89. https://doi.org/10.1007/s11259-019-09747-x.
- Jo, J. K., Seo, S. H., Park, S. E., Kim, H. W., Kim, E. J., Kim, J. S., Pyo, J. Y., Cho, K. M., Kwon, S. J., Park, D. H. et al., (2021). Gut Microbiome and Metabolome Profiles Associated with High-Fat Diet in Mice. Metabolites. 11: 482.
- Jun, J.W., Kim, J.H., Shin, S.P., Han, J.E., Chai, J.Y. and Park, S.C. (2013). Characterization and complete genome sequence of the *shigella* bacteriophage pSf-1. *ResMicrobiol*.164 (10):979–986. https://doi.org/10.1016/j.resmic.2013.08.007.
- Kaci, G., Goudercourt, D., Dennin, V., Pot, B., Doré, J., Ehrlich, S.D., Renault, P., Blottière, H.M., Daniel, C., Delorme, C. (2014) Anti-inflammatory properties of Streptococcussalivarius, a commensal bacterium of the

- oral cavity and digestive tract. *Appl. Environ. Microbiol.* 80: 928–934.
- Kanazawa, A., Aida, M., Yoshida, Y. et al., (2021). Effects of synbiotic supplementation on chronic inflammation and the gut microbiota in obese patients with type 2 diabetes mellitus: a randomized controlled study. Nutrients.13:1– 19. https://doi.org/10.3390/nu13020558.
- Karbach, S.H., Schonfelder, T., Brandao, I., Wilms, E.,
  Hormann, N., Jackel, S., Schuler, R., Finger, S., Knorr,
  M., Lagrange, J., Brandt, M., Waisman, A., Kossmann,
  S., Schafer, K., Munzel, T., Reinhardt, C., Wenzel, P.
  (2016). Gut microbiota promote angiotensin ii-induced arterial hypertension and vascular dysfunction. J Am Heart Assoc.5.
- Kedia, S., Virmani, S., Vuyyuru, K.S., Kumar, P., Kante, B., Sahu, P. *et al.*, (2022). Faecal microbiota transplantation with anti-inflammatory diet (FMT-AID) followed by anti-inflammatory diet *al.*, one is effective in inducing and maintaining remission over 1 year in mild to moderate ulcerative colitis: a randomised controlled trial. *Gut.* 71 (12):2401–13.
- Keshteli, A.H., Millan, B. and Madsen, K.L.(2017). Pretreatment with antibiotics may enhance the efficacy of fecal microbiota transplantation in ulcerative colitis: a meta-analysis. *Mucosal Immunol*. 10:565–566. https://doi.org/10.1038/mi.2016.123.
- Khan, S., Waliullah, S., Godfrey, V., Khan, M.A.W., Ramachandran, R.A., Cantarel, B.L. *et al.*, (2020). Dietary simple sugars alter microbial ecology in the gut and promote colitis in mice. *Sci Transl Med*.12 (567): eaay6218.
- Kim, B., Kim, E.S., Yoo, Y., Bae, H., Chung, I.Y., Cho, Y. (2019). Phage-derived antibacterials: harnessing the simplicity, plasticity, and diversity of phages. *Viruses*.11 (3):268. https://doi.org/10.3390/v11030268.
- Kim, D.S., Choi, H-I., Wang, Y. et. al. (2017). A new treatment strategy for Parkinson's disease through the gut-brain axis: the glucagonlike peptide-1 receptor pathway. *Cell Transplant*. 26:1560–1571.
- Kim, K.M., Yu, K.W., Kang, D.H., Suh, H.J. (2002). Antistress and anti-fatigue effect of fermented rice bran. *Phyther. Res. an Int J.* Devoted to *Pharmacol. Toxicol. Eval. Nat. Prod.Deriv.* 16:700–702
- Kim, S. *et al.*,(2021). Gram-negative bacteria and their lipopolysaccharides in Alzheimer's disease: pathologic roles and therapeutic implications. *Transl Neurodegener*. 10(1):1–23.
- Kitaya, K., and Yasuo, T. (2023). Commonalities and Disparities between Endometriosis and Chronic Endometritis: Therapeutic Potential of Novel Antibiotic Treatment Strategy against Ectopic Endometrium. *Int. J. Mol. Sci.* 24: 2059. https://doi.org/10.3390/IJMS24032059.
- Ko, C.Y., Lin, H-TV., Tsai, G.J. (2013). Gamma-aminobutyric acid production in black soybean milk by *Lactobacillus* brevis FPA 3709 and the antidepressant effect of the

- fermented product on a forced swimming rat model. *Process Biochem.* 48:559–568.
- Kodali, V.P., Sen, R. (2008). Antioxidant and free radical scavenging activities of an exopolysaccharide from a probiotic bacterium. *Biotechnol. J.* 3: 245–251.
- Koeth, R.A., Wang, Z., Levison, B.S., Buffa, J.A, Org, E., Sheehy, B.T. *et al.*, (2013). Intestinal microbiota metabolism of l-carnitine, a nutrient in red meat, promotes atherosclerosis. *Nat Med.* 19: 576–85. <a href="https://doi.org/10.1038/n.m.3145">https://doi.org/10.1038/n.m.3145</a>.
- Koziel, J., and Potempa, J. (2022). Pros and cons of causative association between periodontitis and rheumatoid arthritis. *Periodontol*. 89: 83–98. https://doi.org/10.1111/PRD.12432.
- Krishnamurthy, H.K., Pereira, M., Bosco, J., George, J., Jayaraman, V., Krishna, K., Wang, T., Bei, K. and Rajasekaran, J.J. (2023) Gut commensals and their metabolites in health and disease. *Front. Microbiol.* 14:1244293.
  - https://doi.org/10.3389/fmicb.2023.1244293.
- Krumbeck, J.A., Rasmussen, H.E., Hutkins, R.W. et. al.(2018). Probiotic *Bifidobacterium* strains and galactooligosaccharides improve intestinal barrier function in obese adults but show no synergism when used together as synbiotics. *Microbiome*. 6:121. <a href="https://doi.org/10.1186/s40168-018-0494-4">https://doi.org/10.1186/s40168-018-0494-4</a>.
- Laffin, M., Fedorak, R., Zalasky, A., Park, H., Gill, A., Agrawal, A. *et al.*, (2019). A high-sugar diet rapidly enhances susceptibility to colitis via depletion of luminal short-chain fatty acids in mice. *Sci Rep.* 9:12294.
- Lakshmi, C.P., Ghoshal, U.C., Kumar, S., Goel, A., Misra, A., Mohindra, S. *et al.*, (2010). Frequency and factors associated with small intestinal bacterial overgrowth in patients with cirrhosis of the liver and extra hepatic portal venous obstruction. *DigDis Sci.* 55(4):1142–8.
- Lam, V., Su, J., Koprowski, S., Hsu, A., Tweddell, J.S., Rafiee, P., Gross, G.J., Salzman, N.H., Baker, J.E. (2012). Intestinal microbiota determine severity of myocardial infarction in rats. *FASEB J.* 26 :1727–1735. https://doi.org/10.1096/fj.11-197921.
- Lang, S., Demir, M., Martin, A., Jiang, L., Zhang, X., Duan, Y., Gao, B., Wisplinghoff, H., Kasper, P., Roderburg, C. et al., (2020). Intestinal Virome Signature Associated With Severity of Nonalcoholic Fatty Liver Disease. Gastroenterology 159: 1839–1852. https://doi.org/10.1053/J.GASTRO.2020.07.005.
- Lee, W.J. and Hase, K. (2014).Gut microbiota—generated metabolites in animal health and disease, *Nat Chem Biol*10: 416–424.
- Lei, X., Zhang, W., Liu, T., Xiao, H., Liang, W., Xia, W. et al., (2013).Perinatal supplementation with omega-3 polyunsaturated fatty acids improves sevoflurane-Induced neurodegeneration and memory impairment in neonatal rats. PLoS One. 8:645. <a href="https://doi.org/10.1371/journal.pone.0070645">https://doi.org/10.1371/journal.pone.0070645</a>.
- Leone, V., Gibbons, S.M., Martinez, K., Hutchison, A.L.,

- Huang, E.Y., Cham, C.M *et al.*, (2015). Effects of diurnal variation of gut microbes and high-fat feeding on host circadian clock function and metabolism. *Cell Host Microbe*. 17 (5):681–9.
- Les Dethlefsen, S. H., Sogin, M. L., & Relman, D. A. (2008). The pervasive effects of an antibiotic on the human gut microbiota, as revealed by deep 16S rRNA sequencing. *PLoS Biology*. 6(11): e280. https://doi.org/10.1371/journal.pbio.0060280.
- Li, J., Zhao, F., Wang, Y., Chen, J., Tao, J., Tian, G., Wu, S., Liu, W., Cui, Q., Geng, B., *et al.*, (2017). Gut microbiota dysbiosis contributes to the development of hypertension. *Microbiome*. 5: 14.
- Li, L., Krause, L., Somerset, S. (2017). Associations between micronutrient intakes and gut microbiota in a group of adults with cystic fibrosis. *Clinical Nutrition*. 36:1097–104. https://doi.org/10.1016/j.clnu.2016.06.029.
- Li, P., Zheng, J., Bai, Y., Wang, D., Cui, Z., Li, Y., Zhang, J., Wang, Y. (2020). Characterization of kynurenine pathway in GUT MICROBES 23patients with diarrheapredominant irritable bowel syndrome. *Eur J Histochem*. 64. https://doi.org/10.4081/ejh.2020.3132.
- Lindefeldt, M., Eng, A., Darban, H., Bjerkner, A., Zetterström, C.K, Allander, T. *et al.*, (2019). The ketogenic diet influences taxonomic and functional composition of the gut microbiota in children with severe epilepsy. *NPJ Biofilms Microbiomes*. 5 (1): 5.
- Lindsay, J.O., Whelan, K., Stagg, A.J., Gobin, P., Al-Hassi, H.O.,Rayment, N., Kamm, M., Knight, S.C.,Forbes, A. (2006). Clinical, microbiological, and immunological effects of fructo-oligosaccharide in patients with crohn's disease. *Gut.* 55: 348–355.
- Lindsay, K.L., Buss, C., Wadhwa, P.D., Entringer, S. (2019).The interplay between nutrition and stress in pregnancy: implications for fetal programming of brain development. *BiolPsychiatry*. 85:135–49. <a href="https://doi.org/10.1016/j.biopsych.2018.06.021">https://doi.org/10.1016/j.biopsych.2018.06.021</a>
- Ling, Z., Jin, C., Xie, T., Cheng, Y., Li, L., Wu, N. (2016). Alterations in the fecal microbiota of patients with HIV-1 infection: an observational study in a Chinese population. *Sci Rep.*6:30673.
- Ling, Z., Li, Z., Liu, X., Cheng, Y., Luo, Y., Tong, X. *et al.*,(2014). Altered fecal microbiota composition associated with food allergy in infants. *Appl Environ Microbiol*. 80(8):2546–54.
- Liu, A., Gao, W., Zhu, Y., Hou, X., and Chu, H. (2022). Gut Non-Bacterial Microbiota: Emerging Link to Irritable Bowel Syndrome. Toxins 14, 596. https://doi.org/10.3390/TOXINS14090596.
- Liu, J., Liu, X., Xiong, X.Q., Yang, T., Cui, T., Hou, N.L. *et al.*, (2017). Effect of vitamin A supplementation on gut microbiota in children with autism spectrum disorders a pilot study. *BMC Microbiol*. 17:204. <a href="https://doi.org/10.1186/s12866-017-1096-1">https://doi.org/10.1186/s12866-017-1096-1</a>.
- Liu, L., Poveda, C., Jenkins, P.E., Walton, G.E. (2021). An in vitro approach to studying the microbial community and

- impact of pre and probiotics under anorexia nervosa related dietary restrictions. *Nutrients*.13:4447
- Liu, P., Liu, M., Liu, X., Xue, M., Jiang, Q., Lei, H. (2021).
   Effect of α- linolenic acid (ALA) on proliferation of probiotics and its adhesion to colonic epithelial cells.
   Food Sci Technol. 42:e71921.
   https://doi.org/10.1590/fst.71921.
- Liu, S., Li, E., Sun, Z., Fu, D., Duan, G., Jiang, M. *et al.*, (2019). Altered gut microbiota and short chain fatty acids in chinese children with autism spectrum disorder. *Sci Rep.* 9:30. <a href="https://doi.org/10.1038/s41598-018-36430-z">https://doi.org/10.1038/s41598-018-36430-z</a>
- Long, K.Z., Santos, J.I., Rosado, J.L., Estrada-Garcia, T., Haas, M., Al Mamun, A. *et al.*, (2011). Vitamin A supplementation modifies the association between mucosal innate and adaptive immune responses and resolution of enteric pathogen infections. *Am J ClinNutr*. 93:578–85. https://doi.org/10.3945/ajcn.110.003913.
- Louis, P., Duncan, S.H., Sheridan, P.O., Walker, A.W., Flint, H.J. (2022). Microbial lactate utilisation and the stability of the gut microbiome. *Gut Microbiome*. 3:e3.
- Louis, P., Flint, H.J., Michel, C. (2016). How to manipulate the microbiota: Prebiotics. In Microbiota of the Human Body; Springer: Basel, Switzerland, pp. 119–142.
- Luang-In, V., Katisart, T., Konsue, A. *et al.*, (2020). Psychobiotic effects of multi-strain probiotics originated from thai fermented foods in a rat model. *Food Sci. Anim. Resour.* 40:1014.
- Luettig, J., Rosenthal, R., Barmeyer, C., Schulzke, J. (2015). Claudin-2 as a mediator of leaky gut barrier during intestinal inflammation. *Tissue Barriers*. 3:e977176.
- Luna, R.A., Oezguen, N., Balderas, M., Venkatachalam, A., Runge, J.K, Versalovic, J. *et al.*, (2017). Distinct microbiome-Neuroimmune signatures correlate with functional abdominal pain in children with autism spectrum disorder. *CMGH*. 3: 8. https://doi.org/10.1016/j.jcmgh.2016.11.008.
- Luster, A.D., Alon, R., von Andrian, U.H. (2005). Immune cell migration in inflammation: present and future therapeutic targets. *Nat Immunol*. 6: 1182-1190.
- Lyte, M., Li, W., Opitz, N. et. al. (2006). Induction of anxiety-like behavior in mice during the initial stages of infection with the agent of murine colonic hyperplasia *Citrobacterrodentium*. *Physiol Behav* 89:350–357.
- Ma, N., Tian, Y., Wu, Y., Ma, X. (2017). Contributions of the interaction between dietary protein and gut microbiota to intestinal health. *Curr. Protein Pept. Sci.* 18: 795–808.
- Ma, Y., You, X., Mai, G., Tokuyasu, T., and Liu, C. (2018). A human gut phage catalog correlates the gut phageome with type 2 diabetes. *Microbiome*. 6: 24. <a href="https://doi.org/10.1186/S40168-018-0410-y/FIGURES/6">https://doi.org/10.1186/S40168-018-0410-y/FIGURES/6</a>.
- MacFabe, D.F., Cain, n.E., Boon, F., Ossenkopp, K.P., Cain et. al. Effects of the enteric bacterial metabolic product propionic acid on object-directed behavior, social behavior, cognition, and neuroinflammation in adolescent rats: Relevance to autism spectrum disorder. *Behav Brain*

- *Res.* 217:47–54. https://doi.org/10.1016/j.bbr.2010.10.005.
- Machiels, K., Joossens, M., Sabino, J., De Preter, V., Arijs, I., Eeckhaut, V., Ballet, V., Claes, K., Van Immerseel, F., Verbeke, K., Ferrante, M., Verhaegen, J., Rutgeerts, P., & Vermeire, S. (2013). A decrease of the butyrate-producing species Roseburia hominis and Faecalibacterium prausnitzii defines dysbiosis in patients with ulcerative colitis. *Gut*. 63:1–9. https://doi.org/10.1136/gutjnl-2013-304833.
- Malesza, I.J., Malesza, M., Walkowiak, J., Mussin, N., Walkowiak, D., Aringazina, R. *et al.*, (2021). High-fat, western-style diet, systemic inflammation, and gut microbiota: a narrative review. *Cells*. 10(11): 3164.
- Mandal, S., Godfrey, K.M., McDonald, D., Treuren, W.V., Bjornholt, J.V., Midtvedt, T. *et al.*, (2016). Fat and vitamin intakes during pregnancy have stronger relations with a pro-inflammatory maternal microbiota than does carbohydrate intake. *Microbiome*. 4: 55. https://doi.org/10.1186/s40168-016-0200-3.
- Manrique, P., Zhu, Y., van der Oost, J., Herrema, H., Nieuwdorp, M., de Vos, W.M., and Young, M. (2021). Gut bacteriophage dynamics during fecal microbial transplantation in subjects with metabolic syndrome. *Gut Microb*. 13: 1–15.
- Mao, X., Larsen, S.B., Zachariassen, L.S.F., Brunse, A., Adamberg, S., Castro Mejia, J.L., Adamberg, K., Nielsen, D.S., Hansen, A.K., Friis Hansen, C.H., and Rasmussen, T.S. (2023). Transfer of modified fecal viromes alleviates symptoms of non-alcoholic liver disease and improve blood glucose regulation in an obesity mouse model. *Preprint at bioRxiv*. https://doi.org/10.1101/2023.03.20.532903.
- Marco, M.L., Heeney, D., Binda, S., Cifelli, C.J., Cotter, P.D., Foligne, B. *et al.*, (2017). Health benefits of fermented foods: microbiota and beyond. *Curr OpinBiotechnol*. 44: 94–102.
- Mariano Del Toro-Barbosa, Alejandra Hurtado-Romero, Luis Eduardo Garcia-Amezquita and Tomás García-Cayuela. (2020). Psychobiotics: Mechanisms of Action, Evaluation Methods and Effectiveness in Applications with Food Products. *Nutrients*. 12: 3896. https://doi.org/10.3390/nu12123896.
- Marogianni, C., Sokratous, M., Dardiotis, E., Hadjigeorgiou, G.M., Bogdanos, D., Xiromerisiou, G. (2020). Neurodegeneration and Inflammation-An Interesting Interplay in Parkinson's Disease. *Int. J. Mol. Sci.* 21: 8421.
- Marra, F., Marra, C.A., Richardson, K., Lynd, L.D., Kozyrskyj, A., Patrick, D.M. *et al.*, (2009). Antibiotic use in children is associated with increased risk of asthma. *Pediatrics*. 123:1003–1010. https://doi.org/10.1542/peds.2008-1146
- Matija si'c, B.B., Obermajer, T., Lipoglav sek, L., Grabnar, I., Avgu stin, G., Rogelj, I. (2014). Association of dietary type with fecal microbiota in vegetarians and omnivores

- in Slovenia. Eur J Nutr. 53(4):1051-64.
- McDonald, L. C. (2017). Effects of short- and long-course antibiotics on the lower intestinal microbiome as they relate to traveller's diarrhea. *Journal of Travel Medicine*. 24(1): S35–S38. https://doi.org/10.1093/jtm/taw084.
- McFarlane, C. Ramos, C.I., Johnson, D.W. *et al.*, (2019). Prebiotic, probiotic, and synbiotic supplementation in chronic kidney disease: a systematic review and meta-analysis. *J Ren Nutr.* 29: 209–220. https://doi.org/10.1053/j.jrn.2018.08.008.
- Mesnage, R., Grundler, F., Schwiertz, A., Le Maho, Y., Wilhelmi de Toledo, F. (2019). Changes in human gut microbiota composition are linked to the energy metabolic switch during 10 d of Buchinger fasting. *J. Nutr. Sci.* 8: e36.
- Milani, C., Duranti, S., Bottacini, F., Casey, E., Turroni, F., Mahony, J., Belzer, C., Delgado Palacio, S., Arboleya Montes, S., Mancabelli, L., et al., (2017). The First Microbial Colonizers of the Human Gut: Composition, Activities, and Health Implications of the Infant Gut Microbiota. Microbiol. Mol. Biol. Rev. 81; 10–1128.
- Mohammadi, A.A., Jazayeri, S., Khosravi-Darani, K. *et al.*, (2016). The effects of probiotics on mental health and hypothalamic–pituitary–adrenal axis: a randomized, double-blind, placebo-controlled trial in petrochemical workers. *Nutr. Neurosci.* 19:387–395.
- Mohammadi, G., Dargahi, L., Peymani, A., Mirzanejad, Y., Alizadeh, S.A., Naserpour, T., Nassiri-Asl, M. (2019). The Effects of Probiotic Formulation Pretreatment (*Lactobacillus helveticus* R0052 and *Bifidobacterium longum* R0175) on a Lipopolysaccharide Rat Model. *J. Am. Coll. Nutr.* 38: 209–217.
- Moon, K.T. (2010). Preventing psychotic disorders in high-risk patients. *Am Fam Physician*. 82.
- Mousavi, S., Bereswill, S., Heimesaat, M.M. (2019). Immunomodulatory and Antimicrobial Effects of Vitamin C. *Eur J Microbiol Immunol* (Bp).9: 73–9. https://doi.org/10.1556/1886.2019.00016.
- Mu, C., Yang, Y., Yu, K., Yu, M., Zhang, C., Su, Y., & Zhu, W. (2017). Alteration of metabolomic markers of amino-acid metabolism in piglets with in-feed antibiotics. *AminoAcids*, 49(4): 771–781. https://doi.org/10.1007/s00726-017-2379-4.
- Mudd, A.T., Alexander, L.S., Berding, K., Waworuntu, R.V., Berg, B.M., Donovan, S.M., Dilger, R.N. (2016). Dietary prebiotics, milk fat globule membrane, and lactoferrin affects structural neurodevelopment in the young piglet. *Front. Pediatr.* 4: 4.
- Mukherjee, A. Lordan, C. Ross, R.P. Cotter, P.D. Cronin, P. Joyce, S.A. O'Toole, P.W. O'Connor, E.M. Chen, L. Liu, B. *et al.*, (2020). Increasing levels of *Parasutterella* in the gut microbiome correlate with improving low-density lipoprotein levels in healthy adults consuming resistant potato starch during a randomised trial. *Sci. Rep.* 13: 8096.
- Mukherjee, A., Lordan, C., Ross, R.P., Cotter, P.D. (2024).

- Gut microbes from the phylogenetically diverse genus *Eubacterium* and their various contributions to gut health. *Gut Microbes*. 12 (1): 1802866. https://doi.org/10.1080/19490976.2020.1802866.
- Müller, J.B., Guggenheim, P., Haemmerli, U. (1966). Treatment of chronic portal-systemic encephalopathy with lactulose. *Lancet*. 287: 890–893.
- Muthuirulandi Sethuvel, D.P., Subramanian, N., Pragasam, A.K., Inbanathan, F.Y., Gupta, P., Johnson, J., Sharma, N.C., Hemvani, N., Veeraraghavan, B., Anandan, S. et al., (2019). Insights to the diphtheria toxin encoding prophages amongst clinical isolates of *Corynebacterium diphtheriae* from India. *Indian J Med Microbiol*. 37(3):423–425.
  - https://doi.org/10.4103/ijmm.ijmm\_19\_469.
- Nagamine, T., Sato, N., Seo, G. (2012). Probiotics reduce negative symptoms of schizophrenia: a case report. *Int. Med. J.* 19:72–73.
- Neurath, M. (2014). Cytokines in inflammatory bowel disease. *Nat. Rev. Immunol.* 14: 329–342.
- Neyrinck, A.M., Rodriguez, J., Zhang, Z.,Seethaler, B., Sánchez, C.R., Roumain, M., Hiel, S., Bindels, L.B., Cani, P.D., Paquot, N. *et al.*,(2021). Prebiotic dietary fibre intervention improves fecal markers related to inflammation in obese patients: Results from the Food4Gut randomized placebo-controlled trial. *Eur. J. Nutr.* 60: 3159–3170.
- Nimgampalle, M., Kuna, Y. (2017). Anti-Alzheimer Properties of Probiotic, *Lactobacillusplantarum* MTCC 1325 in Alzheimer's Disease induced Albino Rats. *J. Clin. Diagn. Res.*, 11: KC01–KC05.
- Nishida, K., Sawada. D., Kuwano.Y., Tanaka. H., Rokutan. K. (2019). Health benefits of *Lactobacillus gasseri* CP2305 tablets in young adults exposed to chronic stress: a randomized, double-blind, placebo-controlled study. *Nutrients*. 11 (8):1859. https://doi.org/10.3390/nu11081859.
- Nocerino, R., Paparo, L., Terrin, G., Pezzella, V., Amoroso, A., Cosenza, L., Cecere, G., De Marco, G., Micillo, M., Albano, F. *et al.*, (2017). Cow's milk and rice fermented with *Lactobacillus paracasei* CBA L74 prevent infectious diseases in children: a randomized controlled trial. *Clin Nutr.* 36:118–125. https://doi.org/10.1016/j.clnu.2015. 12. 004.
- Noriega, B.S., Sanchez-Gonzalez, M.A., Salyakina, D., Coffman, J. (2016). Understanding the impact of omega-3 rich diet on the gut microbiota. *Case Rep Med*: 3089303.
- Norman, J.M., Handley, S.A., Baldridge, M.T., Droit, L., Liu, C.Y., Keller, B.C., Kambal, A., Monaco, C.L., Zhao, G., Fleshner, P., *et al.*, (2015). Disease-specific alterations in the enteric virome in inflammatory bowel disease. *Cell*. 160: 447–460.
  - https://doi.org/10.1016/J.CE.S.ALL.2015.01.002
- O'Mahony, S.M., Clarke, G., Borre, Y.E., Dinan, T.G., Cryan, J.F. (2015). Serotonin, tryptophan metabolism and the brain-gut-microbiome axis. Behav. Brain Res. 277: 32–

48.

- O'Riordan, K.J., Collins, M.K., Moloney, G.M., Knox, E.G., Aburto, M.R., Fulling, C., Morley, S.J., Clarke, G., Schellekens, H., Cryan, J.F.(2022). Short chain fatty acids: Microbial metabolites for gut-brain axis signalling. *Mol. Cell Endocrinol.* 546: 111572.
- Oh, J.K., Vasquez, R., Kim, S.H., Hwang, I.C., Song, J.H., Park, J.H., Kim, I.H., Kang, D.K. (2021). Multispecies probiotics alter fecal short-chain fatty acids and lactate levels in weaned pigs by modulating gut microbiota. *J. Anim. Sci. Technol.* 63: 1142–1158.
- Olivares, M., Schüppel, V., Hassan, A.M., Beaumont, M., Neyrinck, A.M., Bindels, L.B. *et al.*, (2018). The potential role of the dipeptidyl peptidase-4-Like activity from the gut microbiota on the host health. *Front Microbiol*. 9:1900. https://doi.org/10.3389/fmicb.2018.01900.
- Olson, C.A., Vuong, H.E., Yano, J.M., Liang, Q.Y., Nusbaum, D.J., Hsiao, E.Y. (2018). The gut microbiota mediates the anti-seizure effects of the ketogenic diet. *Cell.* 173:27. <a href="https://doi.org/10.1016/j.cell.2018.04.027">https://doi.org/10.1016/j.cell.2018.04.027</a>.
- Ooijevaar, R.E., Terveer, E.M., Verspaget, H.W/, Kuijper, E.J., Keller, J.J. (2019). Clinical application and potential of fecal microbiota transplantation. *Annu Rev Med.* 70:335-351.
- Ouwehand, A.C., Tiihonen, K., Saarinen, M. *et al.*, (2009).Influence of a combination of *Lactobacillus acidophilus* NCFM and lactitol on healthy elderly: intestinal and immune parameters. *Br J Nutr*.101:367–375. <a href="https://doi.org/10.1017/s0007114508003097">https://doi.org/10.1017/s0007114508003097</a>
- Ozkul, C., Yalinay, M., Karakan, T. (2019). Islamic fasting leads to an increased abundance of *Akkermansia muciniphila* and *Bacteroides fragilis* group: A preliminary study on intermittent fasting. *Turk. J. Gastroenterol.* 30: 1030–1035.
- Ozkul, C., Yalinay, M., Karakan, T. (2020). Structural changes in gut microbiome after Ramadan fasting: A pilot study. *Benef. Microbes.* 11: 227–233.
- Pama, C. (2019). Socialization by bacteria. Science (80-.). 364:39. https://doi.org/10.1126/science.364.6435.39-a.
- Pandey, N., Malik, R.K., Kaushik, J.K., Singroha, G. (2013). Gassericin A: A circular bacteriocin produced by Lactic acid bacteria *Lactobacillus gasseri*. *World J. Microbiol. Biotechnol.* 29:1977–1987.
- Panesar, P.S., Kumari, S. (2011). Lactulose: Production, purification and potential applications. *Biotechnol. Adv.* 29: 940–948.
- Panpetch, W., Visitchanakun, P., Saisorn, W., Sawatpanich, A., Chatthanathon, P., Somboonna, N., Tumwasorn, S., Leelahavanichkul, A. (2021). *Lactobacillus rhamnosus* attenuates Thai chili extracts induced gut inflammation and dysbiosis despite capsaicin bactericidal effect against the probiotics, a possible toxicity of high dose capsaicin. PLOS ONE. 16 (12):e0261189. <a href="https://doi.org/10.1371/journal.pone.0261189">https://doi.org/10.1371/journal.pone.0261189</a>.
- Parada Venegas, D., De la Fuente, M.K., Landskron, G., González, M.J., Quera, R., Dijkstra, G., Harmsen,

- H.J.M., Faber, K.N., Hermoso, M.A. (2019). Short chain fatty acids (SCFAs)-mediated gut epithelial and immune regulation and its relevance for inflammatory bowel diseases. *Front Immunol*. 10:277. https://doi.org/10.3389/fimmu.2019.00277.
- Parajuli, P., Adamski, M., Verma, N.K. (2017). Bacteriophages are the major drivers of *shigella flexneri* serotype 1c genome plasticity: a complete genome analysis. *BMC Genomics*. 18(1). https://doi.org/10.1186/s12864-017-4109-4.
- Parker, B.J., Wearsch, P.A., Veloo, A.C.M., Rodriguez-Palacios, A. (2020). The Genus *Alistipes*: Gut Bacteria with Emerging Implications to Inflammation, Cancer, and Mental Health. Front. *Immunol*. 11: 906.
- Patel, R., Dupont, H.L. (2015). New Approaches for Bacteriotherapy: Prebiotics, New-Generation Probiotics, and Synbiotics. *Clin. Infect. Dis.* 60: S108–S121.
- Pelsser, L., Frankena, K., Toorman, J., Rodrigues, Pereira R. (2020).Retrospective outcome monitoring of ADHD and nutrition (ROMAN): the effectiveness of the few-foods diet in general practice. Front Psychiatry. 11:96. https://doi.org/10.3389/fpsyt.2020.00096.
- Pham, V., Fehlbaum, S., Seifert, N., Richard, N., Bruins, M., Sybesma, W. *et al.*, (2021). Effects of colon-targeted vitamins on the composition and metabolic activity of the human gut microbiome—a pilot study. *Gut microbes.* 13. https://doi.org/10.1080/19490976.2021.1875774.
- Piwowarczyk, A., Horvath, A., Łukasik, J., Pisula, E., Szajewska, H. (2018). Gluten- and casein-free diet and autism spectrum disorders in children: a systematic review. *Eur J Nutr.* 57: 433–40. https://doi.org/10.1007/s00394-017-1483-2.
- Pool-Zobel, B.L. (2005).Inulin-type fructans and reduction in colon cancer risk: Review of experimental and human data. *Br. J. Nutr.* 93: S73–S90.
- Pourahmad, J., Shaki, F., Tanbakosazan, F., Ghalandari, R., Ettehadi, H.A., and Dahaghin, E. (2011). Protective effects of fungal b-(1/3)-D glucan against oxidative stress cytotoxicity induced by depleted uranium in isolated rat hepatocytes. *Hum. Exp. Toxicol.* 30: 173–181. https://doi.org/10.1177/0960327110372643.
- Qin, J. *et al.*, (2012). A metagenome-wide association study of gut microbiota in type 2 diabetes, *Nature*490: 55–60.
- Qin, L. *et al.*, (2007). Systemic LPS causes chronic neuroinflammation and progressive neurodegeneration. *Glia*. 55 (5):453–62.
- Rahimlou, M., Hosseini, S.A., Majdinasab, N. et. al. (2020). Effects of long term administration of Multi-Strain Probiotic on circulating levels of BDNF, NGF, IL-6 and mental health in patients with multiple sclerosis: a randomized, double-blind, placebo-controlled trial. *Nutr Neurosci*: 1–12.
- Rasmussen, T.S., Mentzel, C.M.J., Kot, W., Castro-Meji'a, J.L., Zuffa, S., Swann, J.R., Hansen, L.H., Vogensen, F.K., Hansen, A.K., and Nielsen, D.S. (2020). Faecal virome transplantation decreases symptoms of type 2

- diabetes and obesity in a murine model. *Gut* 69: 2122–2130. https://doi.org/10.1136/GUTJNL-2019-320005.
- Reid, S.N.S., Ryu, J., Kim, Y., Jeon, B.H. (2018). The effects of fermented *Laminariajaponica* on short-term working memory and physical fitness in the elderly. Evidence-Based Complement. Altern. Med. Evid Based Complement Alternat Med::8109621. https://doi.org/10.1155/2018/8109621.
- Rianda, D., Agustina, R., Setiawan, E.A., Manikam, N.R.M. (2019). Effect of probiotic supplementation on cognitive function in children and adolescents: A systematic review of randomised trials. *Beneficial Microbes*. 10: 68. https://doi.org/10.3920/BM2019.0068.
- Rivera, H.M., Christiansen, K.J., Sullivan, E.L. (2015). The role of maternal obesity in the risk of neuropsychiatric disorders. *Front Neurosci*. 9:194. https://doi.org/10.3389/fnins.2015.00194
- Rondeau, M.P., Meltzer, K., Michel, K.E., McManus, C.M., Washabau, R.J. (2003). Short chain fatty acids stimulate feline colonic smooth muscle contraction. *J Feline MedSurg*. 5 (3):167–173. <a href="https://doi.org/10.1016/s1098-612x(03)00002-0">https://doi.org/10.1016/s1098-612x(03)00002-0</a>.
- Ruengsomwong, S., La-Ongkham, O., Jiang, J., Wannissorn, B., Nakayama, J., Nitisinprasert, S. (2016). Microbial community of healthy Thai vegetarians and nonvegetarians, their core gut microbiota, and pathogen risk. *J.Microbiol. Biotechnol.* 26(10):1723–35.
- Ruiz-Ojeda, F.J., Plaza-Díaz, J., S'aez-Lara, M.J., Gil, A.(2019). Effects of sweeteners on the gut microbiota: a review of experimental studies and clinical trials. *Adv.Nutr.*(Suppl1): S31–48.
- Sakaguchi, Y., Hayashi, T., Kurokawa, K., Nakayama, K., Oshima, K., Fujinaga, Y., Ohnishi, M., Ohtsubo, E., Hattori, M., Oguma, K. (2005). The genome sequence of *Clostridium botulinum* type C neurotoxin-converting phage and the molecular mechanisms of unstable lysogeny. *Proc Natl Acad SciUSA*. 102 (48):17472–17477. https://doi.org/10.1073/pnas.0505503102.
- Sampson, T.R., Debelius, J.W., Thron, T., Janssen, S., Shastri, G.G., Ilhan, Z.E. *et al.*, (2016).Gut microbiota regulate motor deficits and neuro inflammation in a model of parkinson's disease. *Cell*. 167:1469–80.e12. <a href="https://doi.org/10.1016/j.cell.2016.11.018">https://doi.org/10.1016/j.cell.2016.11.018</a>.
- Sandoval-Vargas, D., Concha-Rubio, N.D., Navarrete, P., Castro, M., and Medina, D.A. (2021). Short communication: Obesity intervention resulting in significant changes in the human gut viral composition. *Appl. Sci.* 11:10039. https://doi.org/10.3390/APP112110039/S1.
- Sanguinetti, E. *et al.*, (2018). Microbiome-metabolome signatures in mice genetically prone to develop dementia, fed a normal or fatty diet. *Sci Rep.* 8(1):4907.
- Savignac, H.M., Corona, G., Mills, H., Chen, L., Spencer, J.P., Tzortzis, G., Burnet, P.W. (2013). Prebiotic feeding elevates central brain derived neurotrophic factor, nmethyl-d-aspartate receptor subunits and d-serine.

- Neurochem. Int. 63: 756-764.
- Scher, J.U., Sczesnak, A., Longman, R.S., Segata, N., Ubeda, C., Bielski, C. *et al.*, (2013). Expansion of intestinal *Prevotella copri* correlates with enhanced susceptibility to arthritis, *Elife*. 2: e01202, http://dx.doi.org/10.7554/eLife.01202.
- Scott, E., De Paepe, K., Van deWiele, T. (2022). Postbiotics and Their Health Modulatory Biomolecules. *Biomolecules*. 12: 1640. https://doi.org/10.3390/biom12111640.
- Sequeira, R.P., McDonald, J.A.K., Marchesi, J.R., Clarke, T.B. (2020).Commensal Bacteroidetes protect against *Klebsiella pneumoniae* colonization and transmission through IL-36 signalling. *Nat Microbiol.*; 5:304–313. https://doi.org/10.1038/s41564-019-0640-1
- Sergeev, I.N., Aljutaily, T., Walton, G., Huarte, E. (2020). Effects of Synbiotic supplement on human gut microbiota, body composition and weight loss in obesity. *Nutrients*. 12 (1):222. https://doi.org/10.3390/nu12010222.
- Sharma, R., Kapila, R., Kapasiya, M., Saliganti, V., Dass, G., Kapila, S. (2014). Dietary supplementation of milk fermented with probiotic *Lactobacillus fermentum* enhances systemic immune response and antioxidant capacity in aging mice. *Nutr. Res.*, 34: 968–981.
- Sharon, N. (2006). Carbohydrates as future anti-adhesion drugs for infectious diseases. *Biochim Biophys Acta* 1760: 527—537.
- Shiryaev, S.A., Remacle, A.G., Chernov, A.V., Golubkov, V.S., Motamedchaboki, K., Muranaka, N. *et al.*, (2013). Substrate cleavage profiling suggests a distinct function of *Bacteroides fragilis* metalloproteinases (fragilysin and metalloproteinase (II) at the microbiomeinflammation-cancer interface. *J Biol Chem.* 288 (48): 34956–67.
- Shkoporov, A.N., Hill, C. (2019). Bacteriophages of the human gut: the "known unknown" of the microbiome. *Cell Host Microbe*. 25(2):195–209. https://doi.org/10.1016/j.chom.2019.01.017.
- Shuai Guo, Teng Ma, Lai Yu Kwok, Keyu Quan, Bohai Li, Huan Wang, Heping Zhang, Bilige Menghe & Yongfu Chen.(2024). Effects of postbiotics on chronic diarrhea in young adults: a randomized, double-blind, placebo-controlled crossover trial assessing clinical symptoms, gut microbiota, and metabolite profiles. *Gut Microbes*. 16(1): 2395092,
  - https://doi.org/10.1080/19490976.2024.2395092.
- Simões, C.D., Maukonen, J., Scott, K.P., Virtanen, K.A., Pietiläinen, K.H., Saarela, M. (2014). Impact of a very low-energy diet on the fecal microbiota of obese individuals. *Eur. J. Nutr.* 53: 1421–1429.
- Slykerman, R.F., Coomarasamy, C., Wickens, K., Thompson, J.M.D., Stanley, T. V., Barthow, C. *et al.*, (2019).Exposure to antibiotics in the first 24 months of life and neurocognitive outcomes at 11 years of age. *Psychopharmacology* (Berl). 236:1573–82.

## https://doi.org/10.1007/s00213-019-05216-0.

- Smeekens, S.P., Huttenhower, C., Riza, A., van de Veerdonk, F.L., Zeeuwen, P.L.J.M., Schalkwijk, J. *et al.*, (2014).Skin microbiome imbalance in patients with STAT1/STAT3 defects impairs innate host defense responses, *J. Innate Immun.* 6 253–262, http://dx.doi.org/10.1159/000351912.
- Smith, A.D., Botero, S., Shea-Donohue, T., Urban, J.F. Jr. (2011). The pathogenicity of an enteric *Citrobacter rodentium* Infection is enhanced by deficiencies in the antioxidants selenium and vitamin E. *Infect Immun*. 79:1471–8. https://doi.org/10.1128/IAI.01017-10.
- Smith, A.P., Sutherland, D., Hewlett, P.(2015). An investigation of the acute effects of oligofructose-enriched inulin on subjective wellbeing, mood and cognitive performance. *Nutrients*. 7: 8887–8896.
- Smits, L.P., Bouter, K.E.C., de Vos, W.M., Borody, T.J., Nieuwdorp, M. (2013). Therapeutic potential of fecal microbiota transplantation. *Gastroenterology*.145: 946– 953, <a href="http://dx.doi.org/10.1053/j.gastro.2013.08.058">http://dx.doi.org/10.1053/j.gastro.2013.08.058</a>.
- Sokol, H., Seksik, P., Furet, J.P., Lakhdari, O., Bermúdez-Humatán, L.C, Cratadoux, J.J., Blugeon, S., Bridonneau, C, Futet, J.P., Cotthiet, C, Crangette, C, Vasquez, N., Pochatt, P., Ttugnan, C, Thomas, C, Blottière, H.M., Doté, J., Marteau, P., Seksik, P. and Langella, P. (2009). Low counts of Faecalibacterium prausnitzii in colitis microbiota. Inflammatory Bowel Disease 15: 1183-1189.
- Sokol, S.Y.(1999). Wnt signaling and dorsoventral axis specification in vertebrates. *Curr Opin Genet Dev.* 9(4):405–10.
- Sotoudegan, F., Daniali, M., Hassani, S., Nikfar, S., Abdollahi, M. (2019).Reappraisal of probiotics' safety in humans. *Food Chem. Toxicol.* 129: 22–29.
- Spor, A., Koren, O., Ley, R. (2011). Unraveling the effects of the environment and host genotype on the gut microbiome. *Nat. Rev. Microbiol.* 9: 279–290.
- Stokholm, J., Sevelsted, A., Bønnelykke, K., & Bisgaard, H. (2014). Maternal propensity for infections and risk of childhood asthma: A registry-based cohort study. *The Lancet Respiratory Medicine*. 2(8): 631–637. https://doi.org/10.1016/S2213-2600(14)70152-3.
- Stone, M., Fortin, P.R., Pacheco-Tena, C., Inman, R.D. (2003). Should tetracycline treatment be used more extensively for rheumatoid arthritis? Meta-analysis demonstrates clinical benefit with reduction in disease activity. *J. Rheumatol.* 30:2112–2122.
- Su, K.P., Lai, H.C., Yang, H.T., Su, W.P., Peng, C.Y., Chang, J.P.C. *et al.*, (2014). Omega-3 fatty acids in the prevention of interferon- alpha-induced depression: results from a randomized, controlled trial. *Biol Psychiatry*. 76:508–18. <a href="https://doi.org/10.1016/j.biopsych.2014.01.008">https://doi.org/10.1016/j.biopsych.2014.01.008</a>.
- Sudo, N. (2019). Biogenic Amines: Signals between Commensal Microbiota and Gut Physiology. *Front. Endocrinol*.10: 504.
- Sudo, N., Chida, Y., Aiba, Y., Sonoda, J., Oyama, N., Kubo,

- C., Koga, Y., Yu, X.N. (2004).Postnatal microbial colonization programs the hypothalamic–pituitary–adrenal system for stress response in mice. *J. Physiol.*, 558 Pt 1: 263–275.
- Suez, J., Korem, T., Zeevi, D., Zilberman-Schapira, G., Thaiss, C.A., Maza, O. *et al.*, (2014). Artificial sweeteners induce glucose intolerance by altering the gut microbiota. *Nature*. 514 (7521):181–6.
- Suez, J., Zmora, N., Zilberman-Schapira, G., Mor, U., Dori-Bachash, M., Bashiardes, S., Zur, M., Regev-Lehavi, D., Ben-Zeev Brik, R., Federici, S., Horn, M., Cohen, Y., Moor, A. E., Zeevi, D., Korem, T., Kotler, E., Harmelin, A., Itzkovitz, S., Maharshak, N., Elinav, E. (2018). Post-antibiotic gut mucosal microbiome reconstitution is impaired by probiotics and improved by autologous FMT. *Cell*. 174(6): 1406–1423. https://doi.org/10.1016/j.cell.2018.08.047
- Sun, M., Wu, W., Liu, Z., Cong, Y. (2016). Microbiota metabolite short-chain chain fatty acids, GPCR, and inflammatory bowel diseases. *J. Gastroenterol.* 52: 1–8.
- Sun, Y., Kim, S.W. (2017). Intestinal challenge with enterotoxigenic *Escherichia coli* in pigs, and nutritional intervention to prevent post-weaning diarrhea. *Anim Nutr.* 3 (4):322–330. https://doi.org/10.1016/j.aninu.2017.10.001.
- Swann, O.G., Kilpatrick, M., Breslin, M., Oddy, W.H. (2020). Dietary fiber and its associations with depression and inflammation. *Nutr Rev.* 78: 394–411.
- Swiątecka, D., Narbad, A., Ridgway, K.P., Kostyra, H. (2011). The study on the impact of glycated pea proteins on human intestinal bacteria. *Int J Food Microbiol*. 45(1):267–72.
- Tamtaji, O.R., Heidari-Soureshjani, R., Mirhosseini, N., Kouchaki, E., Bahmani, F., Aghadavod, E., Tajabadi-Ebrahimi, M., Asemi, Z.(2019). Probiotic and selenium co-supplementation, and the effects on clinical, metabolic and genetic status in Alzheimer's disease: A randomized, double-blind, controlled trial. Clin. Nutr. 38: 2569–2575.
- Tang, M., Frank, D.N., Sherlock, L., Ir, D., Robertson, C.E., Krebs, N.F. (2016). Effect of Vitamin E with therapeutic iron supplementation on iron repletion and gut microbiome in US iron deficient infants and toddlers. *J Pediatr Gastroenterol Nutr* 63:379. https://doi.org/10.1097/MPG.00000000000001154.
- Tang, R. and Li, L. (2021). Modulation of Short-Chain Fatty Acids as Potential Therapy Method for Type 2 Diabetes Mellitus. *Can. J. Infect Dis. Med. Microbiol.* 6632266. https://doi.org/10.1155/2021/6632266.
- Tang, W.H., Wang, Z., Kennedy, D.J., Wu, Y., Buffa, J.A., Agatisa-Boyle, B., Li, X.S., Levison, B.S., Hazen, S.L. (2015). Gut microbiota-dependent trimethylamine Noxide (TMAO) pathway contributes to both development of renal insufficiency and mortality risk in chronic kidney disease. *Circ Res.*116:448–455. https://doi.org/10.1161/CIRCRESAHA.116.305360.
- Tang, W.H., Wang, Z., Levison, B.S., Koeth, R.A., Britt, E.B.,

- Fu, X., Wu, Y., Hazen, S.L. (2013).Intestinal microbial metabolism of phosphatidylcholine and cardiovascular risk. *N Engl J Med.* 368:1575–1584.
- Tarantino, G., Finelli, C.(2015). Systematic review on intervention with prebiotics/probiotics in patients with obesity-related nonalcoholic fatty liver disease. *Future Microbiol.* 10: 889–902.
- Tetz, G., Brown, S.M., Hao, Y., Tetz, V. (2019).Type 1 diabetes: an association between autoimmunity, the dynamics of Gut amyloid-producing *E. coli* and their phages. *SciRep.* 9(1). <a href="https://doi.org/10.1038/s41598-019-46087-x">https://doi.org/10.1038/s41598-019-46087-x</a>.
- Thu, T.V., Loh, T.C., Foo, H.L., Yaakub, H., and Bejo, M.H. (2011). Effects of liquid metabolite combinations produced by *Lactobacillus plantarum* on growth performance, faeces characteristics, intestinal morphology and diarrhoea incidence in post weaning piglets. *Trop. Anim. Health Prod.* 43: 69–75. https://doi.org/10.1007/s11250-010-9655-6.
- Ticinesi, A., C. Milani, F. Lauretani, A. Nouvenne, L. Mancabelli, G.A. Lugli, F. Turroni, S. Duranti, M. Mangifesta, A. Viappiani, *et al.*, (2017). Gut microbiota composition is associated with polypharmacy in elderly hospitalized patients. *Sci. Rep.* 7:11102. https://doi.org/10.1038/s41598-017-10734-y.
- Tinelli, C., Di Pino, A., Ficulle, E., Marcelli, S., Feligioni, M. (2019). Hyperhomocysteinemia as a risk factor and potential nutraceutical target for certain pathologies. *Front Nutr.* 6: 49. https://doi.org/10.3389/fnut.2019.00049.
- Tiwari, U., Cummins, E.(2011). Meta-analysis of the effect of glucan intake on blood cholesterol and glucose levels. *Nutrition*, 27: 1008–1016.
- Ton, A.M.M., Campagnaro, B.P., Alves, G.A. et. al. (2020). Oxidative stress and dementia in Alzheimer's patients: effects of synbiotic supplementation. *Oxid Med Cell Longev*: 2638703. <a href="https://doi.org/10.1155/2020/2638703">https://doi.org/10.1155/2020/2638703</a>.
- Tsai, Y.-L., Lin, T.-L., Chang, C.-J., Wu, T.-R., Lai, W.-F., Lu, C.-C., Lai, H.-C. (2019). Probiotics, prebiotics and amelioration of diseases. *J. Biomed. Sci.* 26: 3.
- Uemura, M., Hayashi, F., Ishioka, K., Ihara, K., Yasuda, K., Okazaki, K. *et al.*,(2019). Obesity and mental health improvement following nutritional education focusing on gut microbiota composition in Japanese women: a randomised controlled trial. *Eur JNutr.* 58: 3291–302.
- Valles-Colomer, M., Menni, C., Berry, S.E., Valdes, A.M., Spector, T.D., and Segata, N. (2023). Cardiometabolic health, diet and the gut microbiome: a meta-omics perspective. *Nat. Med.* 29: 551–561. <a href="https://doi.org/10.1038/s41591-023-02260-4">https://doi.org/10.1038/s41591-023-02260-4</a>.
- van de Wouw, M., Walsh, A.M., Crispie, F., van Leuven, L., Lyte, J.M., Boehme. M. *et al.*, (2020). Distinct actions of the fermented beverage kefir on host behaviour, immunity and microbiome gut-brain modules in the mouse. *Microbiome*. 8:67.
- Vinolo, M.A.R., Rodrigues, H.G., Nachbar, R.T. et al., (2011).

- Regulation of Inflammation by Short Chain Fatty Acids. *Nutrients.* 3: 858-876.
- Visconti, C. I. Le Roy, F. Rosa *et al.*,(2019). "Interplay between the human gut microbiome and host metabolism." *Nature Communications*, 10 (1):4505.
- von Martels, J.Z.H., Bourgonje, A.R., Klaassen, M.A.Y., Alkhalifah, H.A.A., Sadaghian Sadabad, M., Vich Vila, A., *et al.*, (2019). Riboflavin supplementation in patients with Crohn's disease (RISE-UP study). *J Crohns Colitis*. https://doi.org/10.1093/ecco-jcc/jjz208.
- Vulevic, J., Drakoularakou, A., Yaqoob, P., Tzortzis, G., Gibson, G.R. (2008). Modulation of the fecal microflora profile and immune function by a novel trans-galactooligosaccharide mixture (b-gos) in healthy elderly volunteers. *Am. J. Clin. Nutr.* 88:1438–1446.
- Vulevic, J., Juric, A., Walton, G.E., Claus, S.P., Tzortzis, G., Toward, R.E., Gibson, G.R. (2015). Influence of galactooligosaccharide mixture (b-gos) on gut microbiota, immune parameters and metabonomics in elderly persons. *Br. J. Nutr.* 114: 586–595.
- Wahida, A., Tang, F., Barr, J.J. (2021).Rethinking phage-bacteria- eukaryotic relationships and their influence on human health. *Cell Host Microbe*. 29 (5):681–688. https://doi.org/10.1016/j.chom.2021.02.007.
- Waldor, M.K., Mekalanos, J.J. (1996). Lysogenic conversion by a filamentous phage encoding cholera toxin. *Science*. 272 (5270):1910–1914. https://doi.org/10.1126/science.272.270.1910.
- Waworuntu, R., Hain, H., Chang, Q., Thiede, L., Hanania, T., Berg, B. (2014). Dietary prebiotics improve memory and social interactions while reducing anxiety when provided early in life to normally developing rodents (637.5). *FASEB J.*, 28: 637.5.
- Wei, S-H., Chen, Y-P., Chen, M-J. (2015). Selecting probiotics with the abilities of enhancing GLP-1 to mitigate the progression of type 1 diabetes in vitro and in vivo. *J. Funct.Foods.* 18:473–486. https://doi.org/10.1016/j.jff.2015.08.016
- Wheeler, M.L., J.J. Limon, A.S. Bar, C.A. Leal, M. Gargus, J. Tang, J. Brown, V.A. Funari, H.L. Wang, T.R. Crother *et al.*, (2016). Immunological consequences of intestinal fungal dysbiosis. *Cell Host Microbe*.19:865–873. <a href="https://doi.org/10.1016/j.chom.2016.05.003">https://doi.org/10.1016/j.chom.2016.05.003</a>
- Williams, S., Chen, L., Savignac, H.M., Tzortzis, G., Anthony, D.C., Burnet, P.W. (2016). Neonatal prebiotic (bgos) supplementation increases the levels of synaptophysin, glun2a-subunits and bdnf proteins in the adult rat hippocampus. *Synapse*. 70: 121–124.
- Wlodarska, M., Willing, B., Keeney, K. M., Menendez, A., Bergstrom, K. S., Gill, N., Russell, S. L., Vallance, B. A., & Finlay, B. B. (2011). Antibiotic treatment alters the colonic mucus layer and predisposes the host to exacerbated Citrobacter rodentium-induced colitis. *Infection and Immunity*. 79(4): 1536–1545. https://doi.org/10.1128/IAI.01104-10.
- Wolters, M., Ahrens, J., Romaní-P'erez, M., Watkins, C.,

- Sanz, Y., Benítez-P'aez, A. *et al.*, (2019). Dietary fat, the gut microbiota, and metabolic health a systematic review conducted within the MyNewGut project. *Clin Nutr Edinb Scotl*. 38(6): 2504–20.
- Wortelboer, K., Nieuwdorp, M., Herrema, H. (2019). Fecal microbiota transplantation beyond *Clostridioides difficile* infections. *EBioMedicine*. 44:716-729.
- Wu, G.D., Chen, J., Hoffmann, C., Bittinger, K., Chen, Y.Y., Keilbaugh, S.A., Bewtra, M., Knights, D., Walters, W.A., Knight R. et al., (2011). Linking long-term dietary patterns with gut microbial enterotypes. Science. 334: 105–108.
- Wu, H., Ma, Y., Peng, X., Qiu, W., Kong, L., Ren, B., et al.,(2020). Antibiotic-induced dysbiosis of the rat oral and gut microbiota and resistance to Salmonella. Arch Oral Biol. 114:104730. https://doi.org/10.1016/j.archoralbio.2020.104730
- Xu, Y., Wang, N., Tan, H.Y., Li, S., Zhang, C., Feng, Y. (2022). Function of *Akkermansiamuciniphila* in obesity: interactions with lipid metabolism, immune response and gut systems. *Front microbiol [internet]*:11. <a href="https://www.frontiersin.org/articles/10.3389/fmicb.2020.">https://www.frontiersin.org/articles/10.3389/fmicb.2020.</a> 00219 2020.
- Yan, Q., Gu, Y., Li, X., Yang, W., Jia, L., Chen, C., Han, X., Huang, Y., Zhao, L., Li, P. et al., (2017). Alterations of the Gut Microbiome in Hypertension. Front. Cell. Infect. Microbiol.
  7: 381. https://doi.org/10.3389/FCIMB.2017.00381.
- Yang, K., Niu, J., Zuo, T., Sun, Y., Xu, Z., Tang, W., Liu, Q.,
  Zhang, J., Ng, E.K.W., Wong, S.K.H., et al., (2021).
  Alterations in the Gut Virome in Obesity and Type 2
  Diabetes Mellitus. Gastroenterology. 161: 1257–1269.e13.
  - https://doi.org/10.1053/J.GASTRO.2021.06.056.
- Yolken, R.H., Jones-Brando, L., Dunigan, D.D., Kannan, G., Dickerson, F., Severance, E. et al., (2014). Chlorovirus aTCV-1 is part of the human oropharyngeal virome and is associated with changes in cognitive functions in humans and mice. Proc Natl Acad Sci USA. 111:16106–11. https://doi.org/10.1073/pnas.1418895111
- Yolken, R.H., Severance, E.G., Sabunciyan, S., Gressitt, K.L., Chen, O., Stallings, C. *et al.*, (2015).Metagenomic sequencing indicates that the oropharyngeal phageome of individuals with schizophrenia differs from that of controls. *Schizophr Bull*. 41:197 https://doi.org/10.1093/schbul/sbu197.
- Younge, N.E., Newgard, C.B., Cotton, C.M., Goldberg, R.N, Muehlbauer, M.J., Bain, J.R. *et al.*, (2019). Disrupted maturation of the microbiota and metabolome among

- extremely preterm infants with postnatal growth failure. *Sci Rep.* 9:547. <a href="https://doi.org/10.1038/s41598-019-44547-v">https://doi.org/10.1038/s41598-019-44547-v</a>.
- Yu, L.M., Zhao, K.J., Wang, S.S., Wang, X., Lu, B. (2018). Gas chromatography/mass spectrometry based metabolomic study in a murine model of irritable bowel syndrome. *World J Gastroenterol*. 24(8):894–904. https://doi.org/10.3748/wjg.v24.i8.894
- Zaibi, M.S., Stocker, C.J., O'Dowd, J. et al., (2010). Roles of GPR41 and GPR43 in leptin secretory responses of murine adipocytes to short chain fatty acids. FEBS Lett. 584: 2381-2386.
- Ze, X., Duncan, S.H.; Louis, P.; Flint, H.J. Ruminococcus bromii is a keystone species for the degradation of resistant starch in the human colon. ISME J. 2012, 6, 1535–1543
- Zhang, X., G'erard, P. (2022). Diet-gut microbiota interactions on cardiovascular disease. *Comput Struct BiotechnolJ*. 20:1528–40.
- Zhao, G., Vatanen, T., Droit, L., Park, A., Kostic, A.D., Poon, T.W. *et al.*, (2017).Intestinal virome changes precede autoimmunity in type 1 diabetes susceptible children. *Proc Natl Acad Sci* USA. 114:114. https://doi.org/10.1073/pnas.1706359114.
- Zheng, Lie, Yong-Yi Ji, Xin, Li Wen, Sheng-Lei, Duan. (2022). Fecal microbiota transplantation in the metabolic diseases: Current status and perspectives *World J Gastroenterol*. 28(23): 2546-2560. <a href="https://doi.org/10.3748/wjg.v28.i23.2546">https://doi.org/10.3748/wjg.v28.i23.2546</a> ISSN 1007-9327 (print) ISSN 2219-2840 (online).
- Zhou, W., Liu, G.-R., Li, P.-L., Dai, Y.-Q., Zhou, K. (2007). Mode of action of plantaricin L-1, an antilisteria bacteriocin produced by *Lactobacillus plantarum*. *Acta Microbiol. Sin.*, 47, 260–264.
- Zhou, X., Qi, W., Hong, T., Xiong, T., Gong, D., Xie, M., Nie, S. P. (2018). Exopolysaccharides from *Lactobacillus plantarum* NCU116. Regulate Intestinal Barrier Function via STAT3 Signaling Pathway. *J. Agric. Food Chem.* 66: 9719–9727.
- Zhu, W., Gregory, J.C., Org, E. *et al.*, (2016).Gut microbial metabolite TMAO enhances platelet hyper-reactivity and thrombosis risk.*Cell*.165:111–124. https://doi.org/10.1016/j.cell.2016.02.011.
- Zuo, T., Wong, S.H., Lam, K., Lui, R., Cheung, K., Tang, W. et al., (2018). Bacteriophage transfer during faecal microbiota transplantation in clostridium difficile infection is associated with treatment outcome. Gut. 67:634–43. https://doi.org/10.1136/gutjnl-2017-313952.

#### How to cite this article:

Neena Kumar Dhiman. 2025. Microbiome Modulation-Based Therapeutic Interventions to Target Enhanced Human Health. *Int.J.Curr.Microbiol.App.Sci.* 14(01): 72-107. https://doi.org/https://doi.org/10.20546/ijcmas.2025.1401.007