

Review Article

<https://doi.org/10.20546/ijcmas.2022.1109.020>

Exomicrobiology: The Effects of Outer Space and its Potential Scope for Mankind

S. Rohith^{ID} and Radhika Oke-Velankar^{ID*}

Modern College of Arts, Science and Commerce (Autonomous), Shivajinagar, Pune, Maharashtra, India

*Corresponding author

ABSTRACT

Ever since the early days of space exploration and research, studies related to the survival of life in the outer space has gained much of momentum. As time passed, the advancements in the technology have given us the ability to develop advanced facilities to study and dive deeper into the responses of microorganisms to the stress conditions existing in the space. After the Theory of Panspermia was proposed by Swedish scientist Svante Arrhenius and later by Sir Fred Hoyle, it's been almost half a century that the research related to microbial behavior in space has begun and is still continuing. The International Space station has been the regular site which functions as the observatory for understanding the effects of space related factors such as Microgravity, Radiation, Vacuum etc. on microorganisms. Their adaptability and survival have always been the keen area of interest along with to identify the mechanisms carried out by predominant species in these extreme conditions. These adaptations can be either useful or be dangerous for both the spacecrafts as well as the crew members onboard. Therefore, it is necessary to monitor and study how the conditions like radiation and microgravity could influence the microorganisms. Advanced techniques in -omics studies have helped in genomic level research of microorganisms exposed to space conditions. Addressing these microbial alterations will help in designing the counter measures to their damaging effects and allow us to utilize their properties that are potentially useful. Various processes are hence being studied in both *in situ* and *ex situ* environments. In this review we will also discuss how these researches would be stepping stones for sustenance in the future longer manned space missions to Mars and beyond.

Keywords

Space exploration,
Panspermia,
Radiation,
Microgravity

Article Info

Received:

02 August 2022

Accepted:

30 August 2022

Available Online:

10 September 2022

Introduction

The outer space is an extreme environment which poses a great challenge for the survival of life of any sort. The outer space is cold, immense in size and is continuously under radiation. Ever since the earth has evolved for more than 4 billion years, it has protected us from these hostile conditions. It's been more than half a century of extensive research and development in the field of space technology, which

has enabled us to study the on-site effects of these hostile conditions on life beyond the safe zones of earth (Horneck *et al.*, 2010).

Every year, the earth receives more than 10^{18} live cells from the outer space in the form of deposition on various comet particles that lands up in the earth's atmosphere. This theory of Panspermia has paved the way for probing how microorganism could survive in the extreme environments of the

outer space (Hoyle *et al.*, 1999). Early biological experiments with microorganisms involving viruses, bacteriophage, unicellular organisms like bacteria were conducted in balloon flights and various unmanned and manned spacecrafts like STS and Soyuz missions. Now the international space station hosts various microorganisms that are contaminants, which are used in experiments and which are present in the natural microbiota of the crew onboard (Dickson, 1991; Venkateswaran *et al.*, 2014).

The first experiment to study about the extent of survival of the microbial life in the *in-situ* space condition was done during the Gemini 9 and Gemini 12 mission of NASA in 1966. This experiment directly exposed T1 bacteriophage and spores of *Penicillium roqueforti* to the outer space conditions like UV rays and X rays for 16 hr and 6 hr and they could successfully survive (Hotchin *et al.*, 1968).

Investigation and analysis of the economically important microbial life to the space conditions will give us more knowledge about how to efficiently engineer and utilize their production of important compounds (Blachowicz *et al.*, 2019).

Some microorganisms which are otherwise commensal in nature can turn into pathogens when exposed to the space conditions (Klaus and Howard, 2006).

It is a known fact that the microorganisms change their physiological and genetic composition to adapt to the stressful environment, so does in the environment of the International Space Station (Horneck *et al.*, 2010).

Several microorganisms which were isolated from the Space Station consisted of both commercially important microbes as well as pathogens (ChecinskaSielaff *et al.*, 2019).

Analyzing these adaptations will help us to design various strategies to reduce the risk factor posed by pathogenic microbes so that future spaceflights and stay at the ISS can be made safe (Taylor *et al.*,

1997). This could help us in implementing more safer measures for long-term space flights and manned space exploration missions.

Effect of Outer Space Environment on Microbial Life

Microgravity

The international space station (ISS) revolves around the earth at an altitude of 400km in the thermosphere layer of the Earth's atmosphere and has a stable orbit known as the Low Earth Orbit (LEO) (Castro *et al.*, 2013).

Hence, in the LEO, the microorganism will undergo tremendous stress conditions. They will experience low gravity (10^{-3} to $10^{-6}g$), low pressure (10^{-7} to 10^{-4} Pa) and a long range of variable temperatures (153 to 393 K). The LEO also consists of ionizing radiations such as galactic cosmic radiation (GCR), solar cosmic radiation (SCR), and also a radiation belt trapped by the magnetosphere of the earth (Horneck *et al.*, 2010).

The majority of the study and experiments on the Space Station are carried out in pressurized space capsules equipped with life supporting systems. Therefore the microorganisms are protected from the extreme most conditions with exception of micro gravity and cosmic radiations. Hence, we can conclude that most differences observed will be on the basis of these two factors (Senatore *et al.*, 2018).

The other external factors would be considered if the study was particularly done outside the environment of the ISS. Herethe term microgravity refers to the phenomena of weightlessness caused because of the less force exerted by gravity, but it is never zero (Horneck and Zell, 2012; Nickerson *et al.*, 2004).

From the experiments it was concluded that one of factor that affected the microbial life because of microgravity was their motility (Benoit and Klaus, 2007). Regarding the non-motile cells, the affected factors because of microgravity were observed as

reduction in lag phase and increase in cell density as compared to that of earth conditions (Zea *et al.*, 2017).

This was found out with a fact that the fluidity around the non-motile cells remains inactive which results in less nutrient uptake and reduced metabolism. This could have caused because of change in the chemical composition of the cell. On the other hand motile cells exhibited better mass transfer due to flagellar movements (Thévenet *et al.*, 1996).

Some of the microorganism like *Escherichia coli* exhibited increased resistance to drug in microgravity because of increased membrane fluidity but *Pseudomonas aeruginosa* did not show any change in membrane fluidity at all (Baker *et al.*, 2004; England *et al.*, 2003).

From this it is evident that effect of microgravity shows a variety of effects on microorganisms. A study showed increased rate of plasmid exchange in gram positive bacteria under these space condition, which might help them to adapt to these stress conditions of space (De Boever *et al.*, 2007).

Another study on *Salmonella* under microgravity showed that expression of about 100 genes were altered which included transcription factors, virulence factors, enzymes for synthesis of various products etc. (Wilson *et al.*, 2002).

Cosmic Radiation

The second factor that affects the microorganism along with microgravity are the Cosmic Radiations. These radiations can induce mutations and influence the microbial metabolism either through indirect interaction of radiation particles with the biomolecules or through the direct energy absorption by the biomolecules (Huang *et al.*, 2018).

In addition to these, experiments with *E. coli*, *B. subtilis* and *D. radiodurans* showed that DNA is damaged because of the radiation, causing breakage

in the double strand which leads to mutations (Micke *et al.*, 1994; Schafer *et al.*, 1994; Zimmermann *et al.*, 1994).

The DNA repair mechanism of microorganisms plays a crucial role in how efficiently they can tolerate the radiation. They carry out this phenomenon by homologous recombination or by joining non-homologous ends. Microorganism *D. radiodurans* was found 5 times more resistant than the spores of *B. subtilis* on its exposure to the space radiation. Hence, it was clear that the ability to repair DNA and the resulting mutations were responsible and the key for the survivability (Senatore *et al.*, 2018).

Apart from the DNA, other biomolecules like proteins and lipids also get altered when they are exposed to cosmic radiations and other radiations like UV and Solar radiations, as a result of which reactive oxygen is produced, causing the damage (Senatore *et al.*, 2018).

Some fungal species and spores were also found to be radiation resistant along with bacteria.

When exposed to UV- C radiation, some fungi could show enhanced UV resistance than the *Bacillus* endospores (Blachowicz *et al.*, 2019).

These increased mutations played a crucial role in the survivability of the microorganism as they altered various pathways of metabolism in them.

Adaptations of Microorganisms in Space Conditions

It is now evident that Microgravity and Cosmic radiation could seriously affect various microbial processes such as gene expression, growth, virulence, drug resistance etc. (Baker *et al.*, 2004; Demain and Fang, 2001; Kacena *et al.*, 1999; Kim *et al.*, 2013).

This is due to the reduction of uptake of nutrients (mass transfer) causing the change in the

metabolism due to low gravity (Zea *et al.*, 2016). Thus, the study of the phenomena like drug resistance, increased virulence activity and alterations in the production of secondary metabolites, resulting in giving out greater quantities of industrially important compounds or even newer compounds is of a huge scope (Wilson *et al.*, 2007; Benoit *et al.*, 2006; Lam *et al.*, 2002).

Drug Resistance

Experimental studies have shown different expression of responses to stress conditions in microorganisms in the outer space conditions, the drug resistance phenomenon being among the prominent ones. Altered drug resistance is a matter of great concern as it affects the health and well-being of the crew members on board a spacecraft.

Studies on bacteria like *E. coli* and *S. aureus* showed that they were more resistant to all antibiotics tested in space conditions as compared to the ground tests (Lapchine *et al.*, 1986).

Apart from the drug resistance, *E. coli* and *S. typhimurium* which were grown in microgravity showed better resistance to thermal stress, osmotic stress and greater ability to survive in macrophages (Lynch *et al.*, 2004).

In *E. coli*, analysis showed that 50 stress response genes were upregulated in the microgravity conditions (Aunins *et al.*, 2018).

Studies also showed the greater probability of horizontal gene transfer in mostly gram-positive bacteria, giving them the drug resistance capacity. It was found that plasmid borne erythromycin and tetracycline genes were horizontally transferred from *Staphylococci* (Vaishampayan and Grohmann, 2019; Sobisch *et al.*, 2019).

B. thuringiensis also showed greater antibiotic resistance and virulence because of the higher chances of the horizontal gene transfer (Beuls *et al.*, 2009).

Virulence Factor

Pathogenic organisms like *P. aeruginosa*, *A. fumigatus* and *Fusarium oxysporum* showed alteration in regulatory signals in the space conditions, which lead to changes in expression of genes and enhancing their pathogenicity. This increased levels of virulence and pathogenicity could be a threat to the crew members on-board long-distance spaceflights with compromised immunity and hence it is crucial to identify their mechanisms and virulent strains (Crabbé *et al.*, 2010; Knox *et al.*, 2016; Urbaniak *et al.*, 2019).

In the studies and isolations conducted at the ISS, the isolated *A. fumigatus* were found more deadlier than the ones in clinical trials in ground conditions (Knox *et al.*, 2016).

Drosophila infected with pathogen *Serratia marcescens* at the ISS were found to have significant shorter life span as compared to the control. In other microorganisms like

C. albicans, the transition into its hyphal form in microgravity showed virulence (Gilbert *et al.*, 2020; Altenburg *et al.*, 2008).

Therefore, above it's clear that how important it is to address the pathogenicity and virulence of microorganisms in space for having safer long- term space missions.

Secondary Metabolites Production

It is evident from the above facts that there is significant effect of microgravity on the growth and metabolism of the microorganisms (Taylor, 1974).

The importance of the secondary metabolite production is of greater interest because of their biotechnological evidences. Many experiments that were carried out in the space and in the simulated environment on ground, showed excess production of secondary metabolites in the microbes that were pharmaceutically important than the normal earth

conditions (Blachowicz *et al.*, 2019; Knox *et al.*, 2016; Romsdahl *et al.*, 2019).

The secondary metabolite production generally increases in the space conditions. Studies on *Humicola fuscoatra* samples on the STS- 77 mission showed higher production of Monorden antibiotic when compared to the ground conditions (Lam *et al.*, 1998).

Similarly, *Streptomyces plicatus* showed increased production of Actinomycin D antibiotic in spaceflight conditions (Lam *et al.*, 2002).

Microorganism *Aspergillus fumigatus* isolated from ISS showed production of novel metabolites of industrial importance, with increased production of Fumigaclavine A which is an antibacterial compound. It was later found out that Frame shift mutation caused these increased Fumigaclavine A production (Knox *et al.*, 2016).

Other *Aspergillus* species like *A. nidulans* and *A.niger* isolated from ISS produced significantly more amount of Asperthicin, an anthraquinone pigment and antioxidant Pyranonigrin (Romsdahl *et al.*, 2020).

These studies indicate that the increased amount of production of these metabolites may act as a protection for these fungi from the space radiations and also how space conditions would be helpful in recovering these commercially important metabolites.

Even after many positive results, many other studies have shown that the microgravity can affect inversely in the secondary metabolite production. In *Streptomyces hygrosopicus*, Rapamycin, which is an immunosuppressive drug, was reduced by almost 90% as compared with the normal conditions (Fang *et al.*, 2000).

Benoit *et al.*, (2006) interestingly pointed out that the enhanced Actinomycin D production in *S. plicatus* was seen between 8-12 days in

microgravity, but the ground control production surpassed it for the rest of the mission.

These prolonged experiments indicated that the activation of specific pathways and production of secondary metabolites by the microorganism could be a time bound activity.

Formation of Biofilm

Under the extreme space conditions involving microgravity and cosmic radiation, the formation of biofilm was first seen in microorganism *P. aeruginosa* (McLean *et al.*, 2001).

Also, other studies have shown that microorganisms forming biofilm on surfaces of spacecrafts, on its water systems could be dangerous to the health of the crew on board or it may cause corrosion of the surfaces making it vulnerable to disasters (Novikova, 2004; Song and Leff, 2005; Gu *et al.*, 1998).

The water systems including pipes, air conditioners, recyclers, electric connectors were the prime spots where these microbial growths were observed (Novikova *et al.*, 2006).

Out of all the microorganisms forming the biofilms, more of the polymer biodegradation were shown by the fungal species (Novikova, 2004).

Bacterial species like *Sphingomonas*, *Methylobacterium* and other coliforms were also found in the drinking water and wastewaters of the space station (Novikova *et al.*, 2006; Koenig and Pierson, 1997).

Studies have indicated that the biofilm formation in microorganisms could be due to increasing cell clumping, enhanced production of extracellular matrix as a result of the extreme space conditions. Hence, the focus of construction of future spaceflights should be given in making it less microbial biodegrading and biofilm resistant (Wilson *et al.*, 2007; Zea *et al.*, 2018).

Changes in Natural Microbiota of the Crew on Board

We are aware that the microbial life is also an integral composition of the human body and it could also get affected due to the exposure to space conditions. Apart from the adverse effects on human health, microbes are also helpful for us in many ways. Studies related have shown that space conditions have enhanced the growth of microorganisms in the Gastrointestinal (GI) sample taken from the crew members, but their diversity was considerably decreased (Taylor *et al.*, 1971).

Samples from the nasal, oral and gastrointestinal areas showed significant difference in the microbial count in the outer space environment. (Brown *et al.*, 1976; Decelle and Taylor, 1976)

The increased amount of pathogenic bacterial species was also seen to show more rate of transfer amongst the crew members. Some of the crew also reported allergic tendencies in the environment of the ISS which might have caused due to prolonged exposure to some fungal pathogens (Venkateswaran *et al.*, 2014; Lencner *et al.*, 1984).

Studies have also showed that the reduction of the natural microbiota of the human body might be due to the limited and confined diet and lifestyle of the crew members (Hao *et al.*, 2018).

The natural microbial diversity of the human body can have very considerable effect on the immunity, immunity responses and healthy conditions of the body (Heyde and Ruder, 2015; Siddiqui *et al.*, 2021). So, it is important to maintain a healthy microbial composition in the crew members to avoid diseases if their immunity is compromised.

Future Prospects of Study and Applications of Microorganism in Missions to Mars and Beyond

With the advanced study of omics, it has become possible of for us to isolate and identify the novel species of microorganisms in the space conditions

(Bijlani *et al.*, 2021; Venkateswaran *et al.*, 2017). The use of Genomics, proteomics, transcriptomics and metabolomics has helped us in understanding the variety of microbial life in the spaceflight conditions. Using the shot gun sequencing technique, we could study the diversified composition of microorganisms at different time in different areas in the space conditions (Checinska *et al.*, 2015).

Studies like metagenomics have helped us in concluding the presence of *Klebsiella pneumoniae* in the ISS at different point of time. It also has aided in understanding their virulence factors and genes of antimicrobial resistance which could be controlled with the ground conditions (Singh *et al.*, 2018).

Analysis of transcriptome, proteome and genome have paved the ways for understanding which proteins were responsible for the adaptations of microorganisms like *A. fumigatus* to space conditions, its stress responses, secondary metabolism etc. (Blachowicz *et al.*, 2019).

All in all, the advancements in the omics have led us to establish a relation between the phenotypical and genotypical responses in the microbial life in outer space environment. This will help us to develop precautionary remedies to pathogenic microorganisms and provide safer manned space flights to mars and beyond in future (Schmidt and Goodwin, 2013).

The curiosity regarding the changes in the interaction between the microorganisms, their pathogenicity, adaptations with other species have revealed us the secrets of microbial life in space conditions (NASA, 2016).

Despite the pathogenicity of microorganisms are a matter of concern the future studies should be focused more towards how beneficial they can be in life supporting processes, how they can be utilized in the waste degradation, waste recovery, oxygen production etc. (Acevedo-Rocha *et al.*, 2019; Carillo *et al.*, 2020).

Studies carried out using certain fungi, bacteria and cyanobacteria were found to produce enhanced vitamins, recycled water, managed wastes, degraded plastics and decontaminated air in the space environment (Roberts *et al.*, 2004; Cortesão *et al.*).

Therefore, the best possible solution is to balance out the microbial flora to minimize the detrimental effects on spacecrafts and crew members and maximize the life supporting processes.

Development of vaccines and against pathogenic organisms like *Salmonella* which causes diarrhea and new therapeutic drugs for treatment in space environments will help us to carry out the same for the use at ground levels where organisms experience the same conditions. Viruses may be dangerous to the crew, but live attenuating these viruses and production of vaccine slow and cease the bacterial growth that are dangerous (Higginson *et al.*, 2016; Horneck *et al.*, 2010).

In general, we could see that how microbial life responds to the space environment, how they can be both detrimental in space conditions as well as how efficiently we can exploit their benefits. Numerous studies and tests were carried out and are being carried out on how the potential of microorganisms can be harnessed for the benefits of human for our long spaceflight missions. Advancement in technologies, advent of -omics have given us a better idea on various changes and have revealed how outer space stress conditions can affect the microbial life and how they respond to it. It is the left to us to develop newer techniques and strategies for sustaining the human life in a healthy manner in future manned long-term missions (Bijlani *et al.*, 2021).

References

Acevedo-Rocha CG, Gronenberg LS, Mack M, Commichau FM, Genee HJ. Microbial cell factories for the sustainable manufacturing of B vitamins. *Curr.Opin.Biotechnol.* 2019 56:18-29. doi: 10.1016/j.copbio.2018.07.006.

Altenburg SD, Nielsen-Preiss SM, Hyman LE.

Increased filamentous growth of *Candida albicans* in simulated microgravity. *Genomics Proteomics Bioinformatics.* 2008 Mar;6(1):42-50. doi: 10.1016/S1672-0229(08)60019-4.

Aunins TR, Erickson KE, Prasad N, Levy SE, Jones A, Shrestha S, Mastracchio R, Stodieck L, Klaus D, Zea L, Chatterjee A. Spaceflight Modifies *Escherichia coli* Gene Expression in Response to Antibiotic Exposure and Reveals Role of Oxidative Stress Response. *Front Microbiol.* 2018 Mar 16;9:310. doi: 10.3389/fmicb.2018.00310.

Baker PW, Meyer ML, Leff LG. *Escherichia coli* growth under modeled reduced gravity. *Microgravity Sci Technol.* 2004;15(4):39-44. doi: 10.1007/BF02870967.

Benoit MR, Klaus DM. Microgravity, bacteria, and the influence of motility. *Advances in Space Research.* 2007 Jan 1;39(7):1225-32.

Benoit MR, Li W, Stodieck LS, Lam KS, Winther CL, Roane TM, Klaus DM. Microbial antibiotic production aboard the International Space Station. *ApplMicrobiolBiotechnol.* 2006 Apr;70(4):403-11. doi: 10.1007/s00253-005-0098-3.

Beuls E, Van Houdt R, Leys N, Dijkstra C, Larkin O, Mahillon J. *Bacillus thuringiensis* conjugation in simulated microgravity. *Astrobiology.* 2009 Oct;9(8):797-805. doi: 10.1089/ast.2009.0383.

Bijlani S, Singh NK, Eedara VVR, Podile AR, Mason CE, Wang CCC, Venkateswaran K. *Methylobacteriumajmalii* sp. nov., Isolated From the International Space Station. *Front Microbiol.* 2021 Mar 15;12:639396. doi: 10.3389/fmicb.2021.639396.

Bijlani S, Stephens E, Singh NK, Venkateswaran K, Wang CCC. Advances in space microbiology. *iScience.* 2021 Apr 3;24(5):102395. doi: 10.1016/j.isci.2021.102395.

Blachowicz A, Chiang AJ, Romsdahl J, Kalkum M, Wang CCC, Venkateswaran K. Proteomic characterization of *Aspergillus fumigatus* isolated from air and surfaces of the International Space Station. *Fungal Genet Biol.* 2019 124:39-46. doi: 10.1016/j.fgb.2019.01.001.

Brown LR, Fromme WJ, Handler SF, Wheatcroft MG, Johnston DA. Effect of Skylab missions on clinical and microbiologic aspects of oral

- health. *J Am Dent Assoc.* 1976 Aug;93(2):357-63. doi: 10.14219/jada.archive.1976.0502.
- Carillo P, Morrone B, Fusco GM, De Pascale S, Roupael Y. Challenges for a sustainable food production system on board of the international space station: A technical review. *Agronomy.* 2020 May;10(5):687.
- Castro S.L., Smith D.J., Ott C.M. NASA ISS Program Science Office; 2013. A Researcher's Guide to International Space Station: Microbial Research.
- Checinska A, Probst AJ, Vaishampayan P, White JR, Kumar D, Stepanov VG, Fox GE, Nilsson HR, Pierson DL, Perry J, Venkateswaran K. Microbiomes of the dust particles collected from the International Space Station and Spacecraft Assembly Facilities. *Microbiome.* 2015 Oct 27;3:50. doi: 10.1186/s40168-015-0116-3.
- ChecinskaSielaff A, Urbaniak C, Mohan GBM, Stepanov VG, Tran Q, Wood JM, Minich J, McDonald D, Mayer T, Knight R, Karouia F, Fox GE, Venkateswaran K. Characterization of the total and viable bacterial and fungal communities associated with the International Space Station surfaces. *Microbiome.* 2019 Apr 8;7(1):50. doi: 10.1186/s40168-019-0666-x.
- Cortês M, Schütze T, Marx R, Moeller R, Meyer V. Grand Challenges in Fungal Biotechnology.
- Crabbé A, Pycke B, Van Houdt R, Monsieus P, Nickerson C, Leys N, Cornelis P. Response of *Pseudomonas aeruginosa* PAO1 to low shear modelled microgravity involves AlgU regulation. *Environ Microbiol.* 2010 Jun;12(6):1545-64. doi: 10.1111/j.1462-2920.2010.02184.x.
- De Boever P, Mergeay M, Ilyin V, Forget-Hanus D, Van der Auwera G, Mahillon J. Conjugation-mediated plasmid exchange between bacteria grown under space flight conditions. *Microgravity Science and Technology.* 2007 Sep;19(5):138-44.
- Decelle JG, Taylor GR. Autoflora in the upper respiratory tract of Apollo astronauts. *Appl Environ Microbiol.* 1976 Nov;32(5):659-65. doi: 10.1128/aem.32.5.659-665.1976.
- Demain AL, Fang A. Secondary metabolism in simulated microgravity. *Chem Rec.* 2001;1(4):333-46. doi: 10.1002/ter.1018.
- Dickson KJ. Summary of biological spaceflight experiments with cells. *ASGSB Bull.* 1991 Jul;4(2):151-260.
- England LS, Gorzelak M, Trevors JT. Growth and membrane polarization in *Pseudomonas aeruginosa* UG2 grown in randomized microgravity in a high aspect ratio vessel. *BiochimBiophys Acta.* 2003 Dec 5;1624(1-3):76-80. doi: 10.1016/j.bbagen.2003.09.012.
- Fang A, Pierson DL, Mishra SK, Demain AL. 2000. Growth of *Streptomyces hygroscopicus* in rotating-wall bioreactor under simulated microgravity inhibits rapamycin production. *Appl. Microbiol.Biotechnol.* 54(1):33-6. doi: 10.1007/s002539900303.
- Gilbert R, Torres M, Clemens R, Hateley S, Hosamani R, Wade W, Bhattacharya S. Spaceflight and simulated microgravity conditions increase virulence of *Serratia marcescens* in the *Drosophila melanogaster* infection model. *NPJ Microgravity.* 2020 Feb 4;6:4. doi: 10.1038/s41526-019-0091-2.
- Gu JD, Ford TE, Berke NS, Mitchell R. Biodeterioration of concrete by the fungus *Fusarium*. *International biodeterioration & biodegradation.* 1998 Jan 1;41(2):101-9.
- Hao Z, Li L, Fu Y, Liu H. The influence of bioregenerative life-support system dietary structure and lifestyle on the gut microbiota: a 105-day ground-based space simulation in Lunar Palace 1. *Environ Microbiol.* 2018 Oct;20(10):3643-3656. doi: 10.1111/1462-2920.14358.
- Heyde KC, Ruder WC. Exploring Host-Microbiome Interactions using an in Silico Model of Biomimetic Robots and Engineered Living Cells. *Sci Rep.* 2015 Jul 16;5:11988. doi: 10.1038/srep11988.
- Higginson EE, Galen JE, Levine MM, Tennant SM. Microgravity as a biological tool to examine host-pathogen interactions and to guide development of therapeutics and preventatives that target pathogenic bacteria. *Pathog Dis.* 2016 Nov;74(8):ftw095. doi: 10.1093/femspd/ftw095.
- Horneck G, Klaus DM, Mancinelli RL. Space microbiology. *Microbiol Mol Biol Rev.* 2010 Mar;74(1):121-56. doi:

- 10.1128/MMBR.00016-09.
- Horneck G, Zell M. Introduction to the EXPOSE-E mission. *Astrobiology*. 2012 May;12(5):373. doi: 10.1089/ast.2012.0831.
- Hotchin J, Lorenz P, Hemenway CL. The survival of terrestrial microorganisms in space at orbital altitudes during Gemini satellite experiments. *Life Sci Space Res*. 1968;6:108-14.
- Hoyle, F., Wickramasinghe, N. Comets – A Vehicle for Panspermia. *Astrophysics and Space Science*. 268, 333–341 (1999). <https://doi.org/10.1023/A:1002428532335>
- Huang B, Li DG, Huang Y, Liu CT. Effects of spaceflight and simulated microgravity on microbial growth and secondary metabolism. *Mil Med Res*. 2018 May 14;5(1):18. doi: 10.1186/s40779-018-0162-9.
- Kacena MA, Merrell GA, Manfredi B, Smith EE, Klaus DM, Todd P. 1999. Bacterial growth in space flight: logistic growth curve parameters for *Escherichia coli* and *Bacillus subtilis*. *Appl.Microbiol.Biotechnol*. Feb;51(2):229-34. doi: 10.1007/s002530051386.
- Kim W, Tengra FK, Young Z, Shong J, Marchand N, Chan HK, Pangule RC, Parra M, Dordick JS, Plawsky JL, Collins CH. Spaceflight promotes biofilm formation by *Pseudomonas aeruginosa*. *PLoS One*. 2013 Apr 29;8(4):e62437. doi: 10.1371/journal.pone.0062437.
- Klaus DM, Howard HN. Antibiotic efficacy and microbial virulence during space flight. *Trends Biotechnol*. 2006 Mar;24(3):131-6. doi: 10.1016/j.tibtech.2006.01.008.
- Klintworth R, Reher HJ, Viktorov AN, Bohle D. Biological induced corrosion of materials II: new test methods and experiences from MIR station. *Acta Astronaut*. 1999 Apr-Jun;44(7-12):569-78. doi: 10.1016/s0094-5765(99)00069-7.
- Knox BP, Blachowicz A, Palmer JM, Romsdahl J, Huttenlocher A, Wang CC, Keller NP, Venkateswaran K. Characterization of *Aspergillus fumigatus* Isolates from Air and Surfaces of the International Space Station. *mSphere*. 2016 Oct 26;1(5):e00227-16. doi: 10.1128/mSphere.00227-16.
- Koenig DW, Pierson DL. Microbiology of the Space Shuttle water system. *Water Sci Technol*. 1997;35(11-12):59-64.
- Lam KS, Gustavson DR, Pirnik DL, Pack E, Bulanlagui C, Mamber SW, Forenza S, Stodieck LS, Klaus DM. The effect of space flight on the production of actinomycin D by *Streptomyces plicatus*. *J Ind MicrobiolBiotechnol*. 2002 Dec;29(6):299-302. doi: 10.1038/sj.jim.7000312.
- Lam KS, Mamber SW, Pack EJ, Forenza S, Fernandes PB, Klaus DM. 1998. The effects of space flight on the production of monorden by *Humicola fuscoatra* WC5157 in solid-state fermentation. *Appl.Microbiol. Biotechnol*. 49(5):579-83. doi: 10.1007/s002530051216.
- Lapchine L, Moatti N, Gasset G, Richoille G, Templier J, Tixador R. Antibiotic activity in space. *Drugs Exp Clin Res*. 1986;12(12):933-8.
- Lencner AA, Lencner CP, Mikelsaar ME, Tjuri ME, Toom MA, Väljaots ME, Silov VM, Liz'ko NN, Legenkov VI, Reznikov IM. Die quantitative Zusammensetzung der Lactoflora des Verdauungstrakts vor und nach kosmischen Flügen unterschiedlicher Dauer (The quantitative composition of the intestinal lactoflora before and after space flights of different lengths). *Nahrung*. 1984;28(6-7):607-13. German. doi: 10.1002/food.19840280608.
- Lynch SV, Brodie EL, Matin A. Role and regulation of sigma S in general resistance conferred by low-shear simulated microgravity in *Escherichia coli*. *J Bacteriol*. 2004 Dec;186(24):8207-12. doi: 10.1128/JB.186.24.8207-8212.2004.
- McLean RJ, Cassanto JM, Barnes MB, Koo JH. Bacterial biofilm formation under microgravity conditions. *FEMS Microbiol Lett*. 2001 Feb 20;195(2):115-9. doi: 10.1111/j.1574-6968.2001.tb10507.x.
- Micke U, Horneck G, Kozubek S. Double strand breaks in the DNA of *Bacillus subtilis* cells irradiated by heavy ions. *Adv Space Res*. 1994 Oct;14(10):207-11. doi: 10.1016/0273-1177(94)90469-3.
- NASA. 2016. Space Biology Science Plan 2016-2025.
- Nickerson CA, Ott CM, Wilson JW, Ramamurthy R, Pierson DL. Microbial responses to microgravity and other low-shear environments. *Microbiol Mol Biol Rev*. 2004 Jun;68(2):345-61. doi:

- 10.1128/MMBR.68.2.345-361.2004.
- Novikova N, De Boever P, Poddubko S, Deshevaya E, Polikarpov N, Rakova N, Coninx I, Mergeay M. Survey of environmental biocontamination on board the International Space Station. *Res Microbiol.* 2006 Jan-Feb;157(1):5-12. doi: 10.1016/j.resmic.2005.07.010.
- Novikova ND. Review of the knowledge of microbial contamination of the Russian manned spacecraft. *Microb Ecol.* 2004 Feb;47(2):127-32. doi: 10.1007/s00248-003-1055-2.
- Roberts MS, Garland JL, Mills AL. Microbial astronauts: assembling microbial communities for advanced life support systems. *Microb Ecol.* 2004 Feb;47(2):137-49. doi: 10.1007/s00248-003-1060-5.
- Romsdahl J, Blachowicz A, Chiang AJ, Chiang YM, Masonjones S, Yaegashi J, Countryman S, Karouia F, Kalkum M, Stajich JE, Venkateswaran K, Wang CCC. International Space Station conditions alter genomics, proteomics, and metabolomics in *Aspergillus nidulans*. *ApplMicrobiolBiotechnol.* 2019 Feb;103(3):1363-1377. doi: 10.1007/s00253-018-9525-0.
- Romsdahl J, Blachowicz A, Chiang YM, Venkateswaran K, Wang CCC. Metabolomic Analysis of *Aspergillus niger* Isolated From the International Space Station Reveals Enhanced Production Levels of the Antioxidant Pyranonigrin A. *Front Microbiol.* 2020 May 21;11:931. doi: 10.3389/fmicb.2020.00931.
- Schafer M, Schmitz C, Bucker H. Heavy ion induced DNA double strand breaks in cells of *E. coli*. *Adv Space Res.* 1994 Oct;14(10):203-6. doi: 10.1016/0273-1177(94)90468-5.
- Schmidt MA, Goodwin TJ. Personalized medicine in human space flight: using Omics based analyses to develop individualized countermeasures that enhance astronaut safety and performance. *Metabolomics.* 2013;9(6):1134-1156. doi: 10.1007/s11306-013-0556-3.
- Senatore G, Mastroleo F, Leys N, Mauriello G. Effect of microgravity & space radiation on microbes. *Future Microbiol.* 2018 Jun 1;13:831-847. doi: 10.2217/fmb-2017-0251.
- Siddiqui R, Akbar N, Khan NA. Gut microbiome and human health under the space environment. *J ApplMicrobiol.* 2021 Jan;130(1):14-24. doi: 10.1111/jam.14789.
- Singh NK, Wood JM, Karouia F, Venkateswaran K. Succession and persistence of microbial communities and antimicrobial resistance genes associated with International Space Station environmental surfaces. *Microbiome.* 2018 Nov 13;6(1):204. doi: 10.1186/s40168-018-0585-2.
- Sobisch LY, Rogowski KM, Fuchs J, Schmieder W, Vaishampayan A, Oles P, Novikova N, Grohmann E. Biofilm Forming Antibiotic Resistant Gram-Positive Pathogens Isolated From Surfaces on the International Space Station. *Front Microbiol.* 2019 Mar 19;10:543. doi: 10.3389/fmicb.2019.00543.
- Song B, Leff LG. Identification and characterization of bacterial isolates from the Mir space station. *Microbiol Res.* 2005;160(2):111-7. doi: 10.1016/j.micres.2004.10.005.
- Taylor G.R., Graves R.C., Brockett R.M., Ferguson J.K., Mieszkuc B.J. NASA Johnson Space Center; 1971. Skylab Environmental and Crew Microbiology Studies.
- Taylor GR, Konstantinova I, Sonnenfeld G, Jennings R. Changes in the immune system during and after spaceflight. *Adv Space Biol Med.* 1997;6:1-32. doi: 10.1016/s1569-2574(08)60076-3.
- Taylor GR. Space microbiology. *Annu Rev Microbiol.* 1974;28(0):121-37. doi: 10.1146/annurev.mi.28.100174.001005.
- Thévenet D, D'Ari R, Bouloc P. The SIGNAL experiment in BIORACK: *Escherichia coli* in microgravity. *J Biotechnol.* 1996 Jun 27;47(2-3):89-97. doi: 10.1016/0168-1656(96)01384-3.
- Urbaniak C, van Dam P, Zaborin A, Zaborina O, Gilbert JA, Torok T, Wang CCC, Venkateswaran K. Genomic Characterization and Virulence Potential of Two *Fusarium oxysporum* Isolates Cultured from the International Space Station. *mSystems.* 2019 Mar 19;4(2):e00345-18. doi: 10.1128/mSystems.00345-18.
- Vaishampayan A, Grohmann E. Multi-resistant biofilm-forming pathogens on the International Space Station. *J Biosci.* 2019 Oct;44(5):125.
- Venkateswaran K, Singh NK, ChecinskaSielaff A,

- Pope RK, Bergman NH, van Tongeren SP, Patel NB, Lawson PA, Satomi M, Williamson CHD, Sahl JW, Keim P, Pierson D, Perry J. Non-Toxin-Producing *Bacillus cereus* Strains Belonging to the *B. anthracis* Clade Isolated from the International Space Station. *mSystems*. 2017 Jun 27;2(3):e00021-17. doi: 10.1128/mSystems.00021-17.
- Venkateswaran K, Vaishampayan P, Cisneros J, Pierson DL, Rogers SO, Perry J. International Space Station environmental microbiome - microbial inventories of ISS filter debris. *Appl.Microbiol.Biotechnol*. 2014;98(14):6453-66. doi: 10.1007/s00253-014-5650-6.
- Wilson JW, Ott CM, HönerzuBentrop K, Ramamurthy R, Quick L, Porwollik S, Cheng P, McClelland M, Tsapralis G, Radabaugh T, Hunt A, Fernandez D, Richter E, Shah M, Kilcoyne M, Joshi L, Nelman-Gonzalez M, Hing S, Parra M, Dumars P, Norwood K, Bober R, Devich J, Ruggles A, Goulart C, Rupert M, Stodieck L, Stafford P, Catella L, Schurr MJ, Buchanan K, Morici L, McCracken J, Allen P, Baker-Coleman C, Hammond T, Vogel J, Nelson R, Pierson DL, Stefanyshyn-Piper HM, Nickerson CA. Space flight alters bacterial gene expression and virulence and reveals a role for global regulator Hfq. *Proc Natl Acad Sci U S A*. 2007 Oct 9;104(41):16299-304. doi: 10.1073/pnas.0707155104.
- Wilson JW, Ott CM, Ramamurthy R, Porwollik S, McClelland M, Pierson DL, Nickerson CA. Low-Shear modeled microgravity alters the *Salmonella enterica* serovar typhimurium stress response in an RpoS-independent manner. *Appl Environ Microbiol*. 2002 Nov;68(11):5408-16. doi: 10.1128/AEM.68.11.5408-5416.2002.
- Zea L, Larsen M, Estante F, Qvortrup K, Moeller R, Dias de Oliveira S, Stodieck L, Klaus D. Phenotypic Changes Exhibited by *E. coli* Cultured in Space. *Front Microbiol*. 2017 Aug 28;8:1598. doi: 10.3389/fmicb.2017.01598.
- Zea L, Nisar Z, Rubin P, Cortesão M, Luo J, McBride SA, Moeller R, Klaus D, Müller D, Varanasi KK, Muecklich F, Stodieck L. Design of a spaceflight biofilm experiment. *Acta Astronaut*. 2018 Jul;148:294-300. doi: 10.1016/j.actaastro.2018.04.039.
- Zea L, Prasad N, Levy SE, Stodieck L, Jones A, Shrestha S, Klaus D. A Molecular Genetic Basis Explaining Altered Bacterial Behavior in Space. *PLoS One*. 2016 Nov 2;11(11):e0164359. doi: 10.1371/journal.pone.0164359.
- Zimmermann H, Schafer M, Schmitz C, Bucker H. Effects of heavy ions on inactivation and DNA double strand breaks in *Deinococcus radiodurans* R1. *Adv Space Res*. 1994 Oct;14(10):213-6. doi: 10.1016/0273-1177(94)90470-7.

How to cite this article:

Rohith, S. and Radhika Oke-Velankar. 2022. Exomicrobiology: The Effects of Outer Space and its Potential Scope for Mankind. *Int.J.Curr.Microbiol.App.Sci*. 11(09): 172-182.
doi: <https://doi.org/10.20546/ijcmas.2022.1109.020>